

***Pseudomonas aeruginosa* infections in Norway**

**An outbreak of *Pseudomonas aeruginosa* infection caused
by contaminated Dent-O-Sept mouth swabs and
invasive *Pseudomonas aeruginosa* infections in Norway
1992-2002**

Bjørn Gunnar Iversen



**Department of Infectious Disease Epidemiology
Division of Infectious Disease Control
Norwegian Institute of Public Health**

Oslo 2009

© **Bjørn Gunnar Iversen, 2009**

*Series of dissertations submitted to the
Faculty of Medicine, University of Oslo
No. 795*

ISBN 978-82-8072-947-7

All rights reserved. No part of this publication may be
reproduced or transmitted, in any form or by any means, without permission.

Cover: Inger Sandved Anfinsen.
Printed in Norway: AiT e-dit AS, Oslo, 2009.

Produced in co-operation with Unipub AS.
The thesis is produced by Unipub AS merely in connection with the
thesis defence. Kindly direct all inquiries regarding the thesis to the copyright
holder or the unit which grants the doctorate.

*Unipub AS is owned by
The University Foundation for Student Life (SiO)*

Contents

Summary	5
Acknowledgements	7
List of papers	9
List of abbreviations.....	10
1. General introduction.....	11
1.1. Microbes and hosts.....	11
1.2. Communicable diseases	11
1.3. Hospital acquired infections.....	14
1.4. Surveillance of infectious diseases.....	15
1.5. Outbreak investigations.....	18
1.6. Medical devices.....	21
1.6.1. Use of medical devices in hospitals and its control	24
1.6.2. Contamination of medical devices	25
1.6.3. Outbreaks caused by medical devices	26
1.7. <i>Pseudomonas aeruginosa</i>	27
1.7.1. Microbiology.....	27
1.7.2. Epidemiology and clinical infection	28
1.8. Biofilm formation.....	28
1.9. Molecular typing methods.....	29
1.10. Causality.....	30
1.10.1. Counterfactual theories	31
1.10.2. Determinism and probabilism	32
2. Background and outline of thesis.....	33
2.1. Background about the outbreak.....	33
2.2. Setting.....	35
2.3. Outline of the thesis.....	36
3. Aims of the thesis.....	37
3.1. Investigating an outbreak of <i>Pseudomonas aeruginosa</i> infections.....	37
3.2. Investigating contamination of the medical device.....	37
3.3. Exploring theories for causality of an outbreak of <i>Pseudomonas aeruginosa</i> infections	37
3.4. Investigating the epidemiology of invasive <i>Pseudomonas aeruginosa</i> infection	38
4. Materials and methods.....	39
4.1. Investigating an outbreak of <i>Pseudomonas aeruginosa</i> infections.....	39
4.2. Investigating contamination of the medical device	40
4.3. Exploring theories for causality of an outbreak of <i>Pseudomonas aeruginosa</i> infections	40
4.4. Investigating the epidemiology of invasive <i>Pseudomonas aeruginosa</i> infection	41
4.5. Data management and statistical analysis	41
4.6. Laboratory analysis	42
4.7. Ethics	43
5. Synopsis of the results of the study	44
5.1. Investigating an outbreak of <i>Pseudomonas aeruginosa</i> infections.....	44
5.2. Investigating contamination of the medical device.....	45
5.3. Exploring theories for causality of an outbreak of <i>Pseudomonas aeruginosa</i> infections	47
5.4. Investigating the epidemiology of invasive <i>Pseudomonas aeruginosa</i> infection	47
6. Discussion.....	50

6.1.	An outbreak of <i>Pseudomonas aeruginosa</i> infections.....	50
6.2.	Contamination of the medical device.....	52
6.3.	Microbial control of moist products.....	54
6.4.	What preservatives were used in the Dent-O-Sept moisturising liquid?	56
6.5.	Medical devices as a source of infection.....	57
6.6.	Claiming causality.....	58
6.7.	Invasive <i>Pseudomonas aeruginosa</i> infection.....	59
6.8.	Methodological weaknesses and limitations	60
6.8.1.	Random error.....	61
6.8.2.	Bias.....	61
6.8.3.	Confounding.....	65
6.8.4.	Effect modification.....	67
6.8.5.	Analysis of causality	67
7.	Main conclusions and further studies.....	69
7.1.	Main conclusions.....	69
7.2.	Proposed actions and further studies	70
8.	References	73
	Appendices.....	89

Tables and figures

<i>Table 1. Categorisation of systems for surveillance of infectious diseases, with three Norwegian systems as examples.</i>	<i>17</i>
<i>Table 2. Critical areas in the production and use of medical devices; problems and possible solutions.</i>	<i>25</i>
<i>Table 3. Time-line of the main events in the Dent-O-Sept case.</i>	<i>33</i>
 <i>Figure 1. The chain of infection</i>	 <i>12</i>
<i>Figure 2. The Dent-O-Sept swab, a non-invasive medical device in Class I.</i>	<i>24</i>
<i>Figure 3. Epidemic curve of the outbreak showing the number of patients cases with the outbreak strain of Pseudomonas aeruginosa isolated from either blood or CSF sample or other sites, by month and year of the first positive culture result.</i>	<i>44</i>
<i>Figure 4. Schematic figure showing the wet part of the production of the Dent-O-Sept swab.</i>	<i>46</i>
<i>Figure 5. The monthly number of cases of invasive Pseudomonas aeruginosa infection in Norway 1992-2002. Forty cases (white bars) belonged to an outbreak caused by a contaminated mouth swab.</i>	<i>48</i>
<i>Figure 6. Association between exposure, outcome and confounder</i>	<i>65</i>

Summary

Infections occurring as a result of stay in hospitals are costly for society and cause much suffering in the patients. A sizeable proportion of these hospital acquired infections are preventable. The hospital patient population is changing with more patients being susceptible to opportunistic infections. The *Pseudomonas* species is ranked among the top ten causes of bacteraemias in hospitals. Medical devices have often been reported to cause outbreaks in hospitals.

The overall aims of this thesis were to investigate a large outbreak of *Pseudomonas* infections and gain knowledge from it, to explore theories for causality and responsibility, and to describe the epidemiology and investigate risk factors for contracting invasive *Pseudomonas aeruginosa* infection.

The research originates from a large outbreak of *P. aeruginosa* infection discovered in 2002 which was caused by a contaminated medical device. From it we explore four areas: 1. the outbreak investigation; 2. the contamination of the medical device involved; 3. theories for causality of the outbreak; and 4. the epidemiology of invasive *Pseudomonas aeruginosa* infection.

Although the research in time moved from the specific *P. aeruginosa* outbreak to explore more general issues the thesis is organised the other way moving from the general to the specific as this gives a better introduction to the subject and is more pedagogical.

Paper I describes an outbreak investigation of *P. aeruginosa* infections, in particular how a nationwide, multicentre investigation was organised and conducted. The team-work and combination of epidemiological and microbiological methods were essential in finding the cause and stopping the outbreak. A total of 231 patients from 24 hospitals were identified with the outbreak strain of *Pseudomonas aeruginosa*; 71 of them died while hospitalised. Genotypically identical strains of the bacterium were isolated from patients, several batches of the Dent-O-Sept swab and from the production plant. We conclude that susceptible patient groups should use only documented quality-controlled, high-level disinfected products and items in the oropharynx.

Paper II describes the investigation of the swabs, the moisturising liquid and the production facility. A total of 76 swabs from 12 different batches produced over a period of 30 weeks were contaminated with the outbreak strain of *Pseudomonas aeruginosa*. Many swabs were

also contaminated with other microbes. More than 250 of 1565 examined swabs were contaminated with one or more microbial species. A system audit revealed serious breaches of production regulations. Biofilm formation in the wet part of the production is proposed as the most plausible explanation for the continuous contamination of the swabs. The legal requirements for microbiological purity of medical devices in Class 1 are not optimal.

Paper III explores the theories for causality of the outbreak of *Pseudomonas aeruginosa* infections. Applying various theories for causality and responsibility from different fields like science, philosophy and law on the actors and acts involved in the outbreak helped elucidating their roles and responsibilities, especially legal theories and counterfactual reasoning. We conclude that many factors contributed to causing the outbreak, but that contamination of a medical device in the production facility was the major necessary condition. The reuse of the medical device in hospitals contributed primarily to the size of the outbreak. In addition there were many errors in the chain from the production of the swabs, through purchasing and storage systems in the health care institutions to the use of the swabs and reporting of defective devices. The unintended error by its producer – and to a minor extent by the hospital practice – was mainly due to non-application of relevant knowledge and skills, and appears to constitute professional negligence. Due to factors outside the discourse of causality, no one was criminally charged for the outbreak.

Paper IV investigates the epidemiology of invasive *Pseudomonas aeruginosa* infection in Norway. Although *P. aeruginosa* usually do not cause infection in healthy persons, it frequently does in patients with certain underlying diseases, and in patients with disrupted barriers, especially in the ICU. Invasive *P. aeruginosa* infection is a rare disease with an incidence rate of 3.16 per 100 000 person-years at risk or 0.20 per 1 000 hospital stays, but very serious for those contracting it with a 30 day case fatality rate of 33%. Patients with malignant neoplasms of lymphoid and haematopoietic tissue and other diseases of blood and blood-forming organs have the highest risk of infection. Prudent antibiotic use is one possible explanation for much lower rates of infection in Norway compared with all other published studies.

Acknowledgements

This thesis is based on work carried out in 2002-2008 at the Department of Infectious Disease Epidemiology at the Norwegian Institute of Public Health. The research all originates from a large outbreak of *Pseudomonas aeruginosa* infection discovered in 2002. I would like to thank the Institute – my working place since 1994 – for providing good working conditions, assisting me when necessary and encouraging me to finish.

First and foremost I would like to thank my supervisor and boss, Preben Aavitsland. Without him there would have been no thesis and scarcely any published articles. He guided the outbreak investigation, initiated the research protocol following up the outbreak investigation, and has encouraged and supported me throughout the process. Despite his hectic daily schedule he has always given me quick and thorough responses to my drafts and my many questions. I would also like to thank my contact supervisor at the University of Oslo, Per Nafstad, who came in later in the process. With fresh eyes and long experience he has quality assured the thesis and helped improving it greatly.

Outbreak investigation is team work. In this outbreak investigation more than 50 hospitals were involved, and many persons in each institution: local outbreak investigation teams, people from microbiological laboratories, infection control personnel, clinicians and administrators. We have collaborated closely with the Ministry of Health and Care, the Norwegian Board of Health Supervision, the Directorate of Health and other institutions, and in addition we have received input from patients and their next of kin and from the media. You have all been indispensable parts of this work. Many of you have been named as co-authors or thanked in the acknowledgements in the individual papers. I thank you all again.

I would especially like to thank Trond Jacobsen at St Olavs Hospital in Trondheim who was just as impatient and enthusiastic as I was. We worked well together and our collaboration exemplifies how genotechnological microbiology and epidemiology supplement each other synergistically in outbreak investigations. Bjørn Hofmann aroused my curiosity and renewed interest for philosophy of science and guided me through a jungle of concepts and terms in a new discipline. The crossing of outbreak investigation with causality and tort law has been fascinating. Thank you. Infection control nurses are the core of hospital infection control and prevention. Sissel Berg-Larsen at Feiringklinikken brought the attention to the mouth swab and Berit Bue at Stavanger University Hospital exemplifies the hard-working, helpful and inspiring infection control nurse. Thank you, both.

Of the many people involved in the outbreak investigation at the Norwegian Institute of Public Health I would especially like to thank my close colleague and friend Hanne-Merete Eriksen. She worked shoulder to shoulder with me during the outbreak investigation and has always been supportive, encouraging and a good critic. Of my colleagues who were not involved in my thesis I am especially indebted to my former boss, Professor Arve Lystad who introduced me to the world of epidemiology, taught me all the links in the chain of infection and raised me in a tradition of infection control and prevention which is broad-minded and interdisciplinary, and based on sound and sober judgements.

My parents, Bjørg and Halfdan, have always stimulated me to be curious and have encouraged me to ask questions and to try to find the answers in encyclopaedias, maps and other books. In the family the standard answer to any question has been the same for generations: Go, look it up!

Finally, I would like to thank the love of my life, my husband and best friend, Bjørn, who for periods has had to put up with not seeing me a lot, and who has caringly coaxed me to finish the thesis.

List of papers

This thesis is based on the following published papers. They will be cited by their roman numbers:

- I Iversen BG, Jacobsen T, Eriksen HM, Bukholm G, Melby KK, Nygard K, Aavitsland P: **An outbreak of *Pseudomonas aeruginosa* infection caused by contaminated mouth swabs.** *Clin Infect Dis* 2007; **44**: 794-801.
- II Iversen BG, Eriksen HM, Bo G, Hagestad K, Jacobsen T, Engeset E, Lassen J, Aavitsland P: ***Pseudomonas aeruginosa* contamination of mouth swabs during production causing a major outbreak.** *Ann Clin Microbiol Antimicrob* 2007; **6**: 3.
- III Iversen BG, Hofmann BM, Aavitsland P. **Questions on causality and responsibility arising from an outbreak of *Pseudomonas aeruginosa* infections in Norway.** *Emerg Themes Epidemiol* 2008, **5**:22.
- IV Iversen BG, Brantsæter AB, Aavitsland P. **Nationwide study of invasive *Pseudomonas aeruginosa* infection in Norway: Importance of underlying disease.** *J Infect* 2008; 57: 139-46.

List of abbreviations

AFLP – Amplified fragment length polymorphism
CE – Communauté Européenne
CFR – Case fatality rate
CI – Confidence interval
CSF – Cerebrospinal fluid
DNA – Deoxyribonucleic acid
EU – European Union
HAI – Hospital acquired infection
HELICS – Hospitals in Europe Link for Infection Control through Surveillance
HUS – Haemolytic uremic syndrome
ICD-10 – International Classification of Diseases, 10th Revision
ICU – Intensive care units
IPSE – Improving Patient Safety in Europe
MRSA – Methicillin resistant *Staphylococcus aureus*
MSIS – The Norwegian Notification System for Communicable Diseases
NIPH – The Norwegian Institute of Public Health
NNIS – National Nosocomial Infections Surveillance System (CDC, USA)
NOIS – The national surveillance system for hospital infections
OR – Odds ratio
PFGE – Pulsed-field gel electrophoresis
PIAH – The national point prevalence surveillance system for hospital infections and antibiotic use
PYAR – Person-years at risk
SPC – Statistical process control
USA – The United States of America
UTI – Urinary tract infection
VAP – Ventilator associated pneumonia

1. General introduction

1.1. Microbes and hosts

Infections have always been a serious threat to mankind, causing disease and death. Through much of historic times man has fought a battle against infectious diseases and its causes. Religious and traditional rules were created to prevent, treat and control the diseases and epidemics (1). Some were based on experience like cleanliness and hygienic measures; others were mere superstition (2), like phlebotomy to cure infections. When microbes were discovered as causes of infectious diseases, the search for cures were intensified and with the advent of antimicrobial therapy some voices in the medical community heralded the end of the era of infectious diseases. The emergence of antibiotic resistance and the increase in the number of debilitated persons with increased susceptibility for infections have curbed this optimism.

Microbes play a natural part in the interaction with humans. They colonise the skin, the outer part of certain orifices and are important for food digestion in the colon and distal ileum. Prudent use of antimicrobials and disinfectants are believed to be important to minimise the disturbance of equilibrium between the different microbes and between microbes and hosts (3).

Microbes are categorised in many ways, one is by pathogenicity. Some bacteria, like *Yersinia pestis* or *Vibrio cholera* usually cause disease in the human host and are called pathogenic whereas others like coagulase negative staphylococci under normal circumstances rarely cause disease and are called apathogenic. In between these two groups there is a continuum of pathogenicity of microbes that can cause disease under certain circumstances when one or more defence mechanisms fail, like disruption of barriers introducing bacteria into sterile body sites or the weakening of immunity during cancer treatment. This group of microbes is called opportunistic. *Pseudomonas aeruginosa* is an opportunistic bacterium (4-7). In addition, within each species of microbes there may be great variability. Among *E. coli* for example one may find apathogenic, opportunistic and pathogenic strains depending on presence of virulence factors.

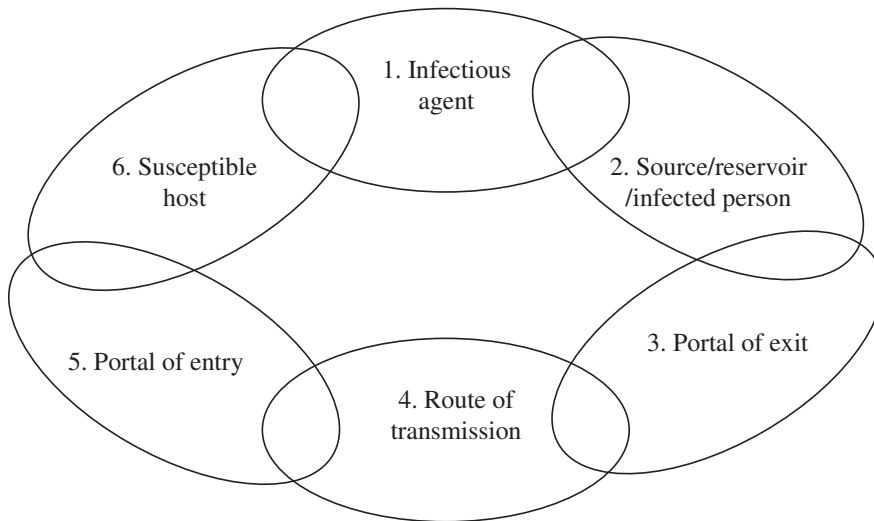
1.2. Communicable diseases

Epidemiology of communicable diseases differs from epidemiology of chronic diseases mainly in that the patient, the case, can become infectious and thus be the source of disease

for new cases. The epidemic potential for an infectious disease is mathematically described through the basic reproductive rate, called R_0 . It is dependent on the risk of transmission per contact (β), the average number of contacts (κ) and the duration of infectivity for an infected person (D), mathematically described as: $R_0 = \beta \times \kappa \times D$ (8). In order for an infectious disease to expand and spread each infected person on average has to infect more than one new person, i.e. $R_0 > 1$. Some non-infectious diseases can be said to spread in populations through altered behaviour patterns. Diet and drinking trends may spread and cause epidemics of obesity and alcoholism. In addition, some infections are caused by microorganisms already present on the patient, the so called endogenous infections. However, in epidemiology communicable diseases and infectious diseases are usually seen as synonyms (9).

A conceptual model for communicable disease spread is the chain of infection (Figure 1) (1, 10). This model consists of six links which all have to be present for an infection to spread. If one link is broken, propagation ceases. Consequently the model is used in infection control and prevention to study where to intervene to prevent the spread of infections and to stop outbreaks.

Figure 1. *The chain of infection*



To break the chain of infection in the hospital setting one needs to analyse each link.

1. **Infectious agent:** All pathogenic microbes, most opportunistic and even some microbes generally considered to be apathogenic, may cause infections due to the susceptibility of many patients. A general reduction of all potentially pathogenic microbes would reduce the risk of infection.

2. **Source/reservoir/infected person:** There are three groups of reservoirs in hospitals, a) patients and personnel, b) the hospital environment, and c) medical devices, pharmaceuticals, food and water.

2 a) patients and personnel: Infectious people can be treated and made non-infectious.

2 b) the hospital environment: Contaminated surfaces can be cleaned and disinfected. Certain areas are more important to control like door knobs, hand rails and switches where many people touch frequently whereas ceilings and floors are less important from an infection control perspective. For moisture-prone bacteria wet areas like sinks, faucets, flasks, other containers and tubes need to be disinfected and controlled regularly. One needs to analyse where bacteria can multiply into numbers posing as a risk for patients and implement preventive measures. In specialised rooms like operating theatres even the air needs to be monitored for bacterial contamination.

2c) medical devices, pharmaceuticals, food and water: Depending on type of use medical devices need to be clean, disinfected or sterilised and there needs to be monitoring systems in place to verify the microbial status of the products. In many hospital outbreaks where medical devices have been involved, deficiencies in the cleaning, disinfecting, and sterilising process of reusable equipment and in the control systems of these processes have been demonstrated. Food and drinking water is not sterile and may contain microbes that can cause infections in susceptible patient groups. Potentially harmful foods have to be removed for these selected groups.

3. **Portal of exit:** The main portals of exit of microbes from a patient are the mouth and airways, urethra, anus, damaged skin (where blood and pus can emerge), and intact skin. To prevent microbe containing body fluids, solid body parts or excretions from being transmitted, the mouth and nose can be covered with a surgical mask, cuts, bruises and abscesses covered with bandages, diapers used and intact skin can be covered to prevent the spread of exfoliations.

4. **Route of transmission:** There are mainly three transmission routes: a) direct and indirect contact transmission, b) droplet transmission and c) airborne transmission. Direct and indirect contact transmission is by far the most important mode of transmission and the hands of health care workers are in most hospital hygiene publications considered to be the most important vehicle for transport of infectious agents in the hospital. Isolation and quarantine or other ways of physical distancing to prevent the infectious from coming into contact with the non-infected are other methods for breaking the route of transmission. The measures instituted depend mainly on the mode of transmission for the particular infectious agent and on the severity of disease that it may cause.
5. **Portal of entry:** This is often the reverse of link 3 where all natural and artificial orifices and the intact skin and other outer surfaces can be a portal. By blocking the portal of entry using a surgical mask or respirator covering the mouth and nose, covering intact or cut skin, eyes etc. and depending on the mode of transmission, one can block the infectious agent from entering. In the hospital setting there are many more entry portals due to artificial openings stemming from surgery, catheters and other medical devices which have broken many of the natural defence barriers. That is why it is so important only to use quality-controlled equipment and to perform all critical procedures with the highest hygienic standards.
6. **Susceptible host:** In the hospital, many patients are especially susceptible for contracting infections. The main means of reducing their susceptibility is through immunisations where a vaccine is available and through improving their general conditions in order to better fight off any intruding microbe. In some instances like for certain surgical procedures short course antibiotic prophylaxis is given to reduce the risk of infection.

1.3. Hospital acquired infections

The risk of contracting an infection is much larger inside a hospital than outside. There are several reasons for this.

Firstly, people in hospitals are already ill. They are often bedridden and pacified making them more susceptible to infections of the skin and airways. Many patients have a reduced capacity to battle infections due to a weakened immune system. Trauma, surgical procedures and medical devices like catheters have disrupted natural defence barriers making it easier for microbes to gain access and cause infection.

Secondly, in hospitals there are many patients with infectious diseases and these may be infectious sources to other patients who consequently more easily may contract new

microbes. And the microbes are easily transferred from patient to patient through direct or indirect contact via health care workers, fixture or medical devices, or for some microbes via droplets or through the air.

Thirdly, the use of antimicrobials per population is much larger inside hospitals than in the general community (although the total consumption is much larger outside) (3). This antibiotic pressure drives a selection for more resistant strains of bacteria causing the bacterial flora in hospitals to be quite different from the one outside. All this makes the risk of contracting an infection much higher in hospitals than outside. It also makes it more difficult to treat due to antibiotic resistance and the susceptibility of the patients.

A hospital acquired infection (HAI) is usually defined as an infection that follows a stay in hospital, but that was not present or incubating at the time of admission to the hospital (11). For bacterial infections a standard incubation period of 48 hours is usually used meaning that infections occurring at least 48 hours after admission to the hospital are considered to be hospital acquired. Hospital acquired is synonymous to nosocomial which is the Greek word pertaining hospital. A wider term often used is health care associated infections which includes all infections that can be associated with hospitals, nursing homes or the outpatient setting in primary or specialist health care. Although an infection may be hospital acquired, this does not necessarily mean the patient acquired the microbe inside the hospital. A large proportion of HAIs results from microbes belonging to the patient's normal bacterial flora. Catheter-related urinary tract infections (UTIs) may be caused by *E. coli* from the patient's intestinal flora and a surgical site infection from the patient's normal skin flora.

1.4. Surveillance of infectious diseases

Public health surveillance is defined as the ongoing systematic collection, analysis, and interpretation of outcome-specific data essential to the planning, implementation and evaluation of public health practice, closely integrated with the timely dissemination of these data to those who need to know (12). The final link in the surveillance chain is the application of these data to prevention and control.

Surveillance of infectious diseases can be categorised in several ways (see Table 1 for categorisation 1a – 6b). In Norway the Norwegian Notification System for Communicable Diseases (in Norwegian abbreviated MSIS) consists of several subsystems. The system for the major part of notifiable diseases in MSIS is a cohort (category 1b) for the whole country (2b) receiving clinical and microbiological information (3a and b) on an individual basis (4b), is mostly passive (5b) but with some active follow-up of missing data (5a) and mainly manual

(6a) but with an aim to increase the electronic transfer, especially from medical microbiological laboratories (6b). Most developed countries have somewhat similar systems. In the period 1975-1991 detection of most bacteria from blood or cerebrospinal fluid (CSF) were individually reportable from medical microbiological laboratories to MSIS. *Pseudomonas aeruginosa* was among the bacteria to be notified but MSIS did not publish tables on a bacterial genus level, only based on the site of infection where the microbes were detected from.

For HAI, Norway has had a tradition since 1979 of repeated national point prevalence studies (in Norwegian abbreviated PIAH). On a given day and time all patients occupying a bed in somatic wards in hospitals are counted (denominator) as are those with one of the four most frequent HAIs: UTIs, lower respiratory tract infections, surgical site infections and blood stream infections (numerator). The infections are not specified by causative microbial agent, so the number of *P. aeruginosa* infections cannot be specified. Numerators and denominators are summed up by ward and hospital and aggregated data sent to the Norwegian Institute of Public Health (NIPH). From 2002 all hospitals and nursing homes have been asked to submit data to NIPH twice-yearly. From 2004 they have had the opportunity to enter the data electronically via Internet. (According to the list this surveillance system is: 1a, 2b, 3a, 4a, 5a, 6b.)

In 2005 NIPH implemented a national surveillance system for hospital infections (in Norwegian abbreviated NOIS). In this system, surgical site infections following certain surgical procedures will be subject to surveillance during a given 3-month period each year. The system is in accordance with the European hospital surveillance network (Hospitals in Europe Link for Infection Control through Surveillance (HELICS) / Improving Patient Safety in Europe (IPSE)) which in turn is based on the National Nosocomial Infections Surveillance System (NNIS) from The United States of America (USA). Participation in NOIS is mandatory for all hospitals. (According to the list this surveillance system is: 1b, 2b, 3a, 4b, 5a, 6b.) In the NOIS system the causative microbial agent is not specified.

Table 1. Categorisation of systems for surveillance of infectious diseases, with three Norwegian systems as examples.

Category	Description	MSIS	PIAH	NOIS
1. By study type	a. Repeated cross-sectional study (prevalence study or survey)		X	
	b. Cohort study (incidence study)	X		X
2. By selection of reporters	a. Sentinel reporting			
	b. Regional or total coverage	X	X	X
3. By source of information	a. Clinical information	X	X	X
	b. Microbiological detection and information	X		
	c. Serological marker of infection			
	d. Surrogate markers of infection (Hospital economical reimbursements, mortality statistics, work force absenteeism etc.)			
4. By type of data	a. Aggregated data		X	
	b. Individual data	X		X
5. By type of data collection	a. Active surveillance	(X)	X	X
	b. Passive surveillance	X		
6. By mode of data transfer	a. Manual, paper-based system	X		
	b. Electronic, automated system	(X)	X	X

In some Norwegian hospitals with on-site microbiological laboratories infection control personnel get regular reports of all detections of certain indicator bacteria. For some they even can get alerted after single findings. There is no national standard for which bacteria to cover or how to report. By selecting *P. aeruginosa* as an indicator bacterium the hospitals can measure the occurrence of detection of this microbe.

Nationally and internationally there is increased emphasis on patient safety and quality assurance systems. Hospitals are increasingly required to measure and report on risks for hazards occurring in hospitals and to set up plans for minimising these risks. Surveillance systems form a basis for these efforts and increasing resources are spent on developing, improving and implementing systems for surveillance of infectious diseases in hospitals.

In Norway, NOIS is currently covering surveillance of surgical site infections following a few surgical procedures. In addition to include more surgical procedures new modules are being developed in other high risk areas for acquiring infections in hospitals as in intensive care units (ICU). Modules are also being developed to more detailed measure the consumption of

antimicrobial drugs in hospitals in order to detect overuse and misuse, and consequently to be able to suggest alterations and improvements. Development of better systems for on-site microbiological laboratory surveillance will also be a priority in the coming years. Improved and better accessible databases will ease this development. In addition surveillance systems measuring the incidence of infections and of antibiotic use in nursing homes are being piloted.

To date, only results from national prevalence studies have been published in scientific papers (13-21). Results from NOIS have only been published on NIPH's Internet pages but several papers will be submitted for publication in the near future.

1.5. Outbreak investigations

Outbreak investigations in hospitals and elsewhere follow the same general structure and regardless of causative agent. An outbreak can be defined in several ways, the simplest being “an event involving more cases than usual”. A more elaborate definition is: An event involving more cases than expected of a certain disease within a given time and area. Another definition is two or more cases of the same disease and with a presumed common source (22).

Outbreaks in hospitals are either common source outbreaks or caused by local spread via patients, personnel, equipment or environment, or to a lesser degree, through air. One can also find mixed-pattern outbreaks where a common source introduces the microbe which then in turn spreads locally.

1. Common source

- a. Medical devices, medicine or other remedy produced locally or procured
- b. Food, drinks or water produced locally or procured
- c. A fixed, maintained source in the hospital (contaminated sink, faucet, ice machine, flower pots and vases, ventilator system or other)
- d. An environmental condition that enables microbiological growth, e.g. moist walls or ceilings where fungus or bacteria can grow
- e. A chronic carrier among the personnel (e.g. a chronic MRSA (Methicillin resistant *Staphylococcus aureus*) nasal carrier or a surgeon with a chronic blood-borne viral infection transmitting to some of her patients during surgery)

2. Local spread

- a. Contact transmission from person to person (patients and personnel), either directly or indirectly via the environment

- b. Droplet transmission from person to person being in close proximity to each other (usually less than 1 meter)
- c. Airborne transmission where microbes can travel longer distances through air (i.e. more than 1 meter)

Outbreaks caused by *P. aeruginosa* can either be common source or through local spread. As the bacterium has affinity for water, moist products, moist environment, or moist areas of the human body are usually the reservoir for the bacterium. The spread is usually through contact, but droplet transmission can also occur, especially from droplet generating procedures.

There are two main ways of detecting outbreaks

1. Indicator based surveillance: Ongoing routine surveillance systems detect an increased number or cases under surveillance or unusual patterns in the data.
2. Event based surveillance: Outbreaks are detected through unstructured reporting systems like media, international alerts, outbreak reporting, and unusual events reporting from the health services.

It is crucial to have systems in place to detect outbreaks as early as possible. Better and more elaborate surveillance systems for events, diseases and microbes will improve our ability for early detection. However, the more sensitive a system is the more “noise” is also detected. And no matter how elaborate an indicator based surveillance system is in detecting outbreaks, we always will have to appreciate the unease or sixth sense of health care personnel as a valuable, additional alert system.

In outbreak investigations there is a range of tasks to undertake, preferably in a logical, chronological order, although many of the tasks needs to be performed in parallel or repeated several times.

The following major tasks for outbreak investigations can be listed (12, 22):

- Prepare and plan
 - Have a general, structured plan ready.
 - Know your potential collaborators, their legal position and skills.
 - Maintain your skill through training.
- Detect and verify

- Have a system in place to receive and assess warnings and notifications about possible outbreaks in order to determine whether further investigations are necessary.
- Alert and inform stakeholders
 - To mitigate the extent and consequences of an outbreak it is important to notify all relevant stakeholders. Norwegian laws and regulations give detailed instructions on whom to inform and when. For example all suspected and verified outbreaks in health care institutions are to be reported immediately to the chief medical officer in the county and to NIPH. (23, 24)
- Make a case definition, identify and verify cases
 - Keep in mind that the case definition can change over time as the knowledge of the outbreak increases. For example one may in the beginning of an outbreak use a syndromic diagnosis to be replaced by an etiologic diagnosis later.
- Describe the outbreak in terms of time, place and person
 - Use basic, epidemiological tools. Describe also who are at risk of becoming ill.
- Generate hypotheses
 - Base your hypotheses on all available information at the time
- Test hypotheses
 - Once hypotheses are generated they are to be tested against the information gathered. The main tools are:
 1. Epidemiological studies
 2. Microbiological sampling
 3. Performing environmental investigations and assessments
 - Based on the preliminary findings from the hypothesis testing, decide on whether to plan for a more systematic study.
- Implement control and preventive measures
 - If the outbreak is sufficiently serious it may be necessary to implement measures on limited knowledge
- Communicate findings
 - Keep detailed minutes of all actions from the very start
 - Prepare a written report

- Keep the mass media informed. Coordinate the main messages with the other stakeholders. In the past years increasing time is spent on keeping the mass media informed. When the outbreak involves children, deaths, differences of opinion among investigators or stakeholders, errors made or political issues, the media attention can be particularly intense.

In outbreak investigations the lack of time is in conflict with the need to be precise, systematic and deliberate. This urgency is the main difference between outbreak investigations and prospectively planned epidemiological studies. The ideal epidemiological study is prospective, well planned, with unambiguous definitions and clear and profound hypotheses to be tested. This would be ideal for outbreak investigations as well but is not feasible most of the time, and one needs to make compromises. For some outbreaks the number of cases is few and the statistical power may be low. For outbreaks with serious outcomes like death and debilitating disease the need for a quick result may force the investigators to compromise on the accuracy of the protocol. As a consequence all results from an outbreak investigation need to be interpreted with caution. When the media pressure is high and the public outcry to come up with an explanation is loud, it is tempting to conclude prematurely and too confidently.

Another contrast with planned epidemiological studies is that the hypotheses, definitions and the protocol may change over time. At the start of an outbreak investigation there is little knowledge so the investigation needs to begin broadly. As knowledge is gained, hypotheses can be more specific, definitions narrower and the protocol more structured. In addition it may be necessary to implement control measures before the investigation is complete which can make it more difficult to come up with clear results. However, whereas the aim of many planned epidemiological studies is to detect small differences between various exposed groups, the main aim of an outbreak investigation is to detect the reasons for the outbreak in order to prevent further cases.

1.6. Medical devices

Prior to 1995 unsterile medical devices were poorly regulated in Norway. Products like mouth swabs were only regulated through general regulations on product control. In 1995 the Act on medical devices and its regulation were introduced (24, 25). The purpose of the Act and its regulation is to prevent harmful effects, mishaps and accidents and to ensure that medical

devices is tested and used in a professional and ethically justifiable way (24). When in doubt the Ministry of health and care defines whether a product is to be called medical device.

Through the European Economic Area Agreement Norway abides by much of the legislation within the European Union (EU), including European Council Directive 93/42/EEC concerning medical devices (26). Norwegian jurisdiction on medical devices today is to a large extent, direct translations of EU council directives.

A medical device is defined in the Council Directive as:

“‘medical device’ means any instrument, apparatus, appliance, material or other article, whether used alone or in combination, including the software necessary for its proper application intended by the manufacturer to be used for human beings for the purpose of:

- diagnosis, prevention, monitoring, treatment or alleviation of disease,
- diagnosis, monitoring, treatment, alleviation of or compensation for an injury or handicap,
- investigation, replacement or modification of the anatomy or of a physiological process,
- control of conception,

and which does not achieve its principal intended action in or on the human body by pharmacological, immunological or metabolic means, but which may be assisted in its function by such means; “ (26)

The Directive classifies medical devices into four classes, I, IIa, IIb and III. The classification rules are based on the vulnerability of the human body taking account of the potential risks associated with the technical design and manufacture of the devices. Annex IX of the Directive gives detailed rules for classification of medical devices into these four classes.

In order to mark a medical device with CE (Communauté Européenne) the Council Directive states that the producer must produce a declaration of conformity. The declaration of conformity is the procedure whereby the manufacturer ensures and declares that the products concerned meet the provisions of the Directive which apply to them. The list of provisions is quite detailed.

For non-invasive medical devices in Class I, there are no demands for sterility. The devices must, when used, “not compromise the clinical condition or the safety of patients”. “The devices and manufacturing processes must be designed in such a way as to eliminate or

reduce as far as possible the risk of infection to the patient, user and third parties.” Beyond this, the directive does not specify the microbial quality of the product.

In comparison, pharmaceutical preparations for use in the respiratory tract are according to the European Pharmacopoeia classified in a Category 2 where – in addition to other microbiological requirements – the absence of *Pseudomonas aeruginosa* needs to be documented (27).

In the aftermath of the outbreak of *Pseudomonas aeruginosa* described in this thesis there were public discussions on how to classify the Dent-O-Sept swab (Figure 2). Was it a medical device or a cosmetic product? The Act (28) and Regulation on cosmetics is rather general and defines cosmetics and body care products as: “Any product intended for use on the bodily surface (like skin, head hair and other hair growth, nails, lips and external genitals) or on the teeth and the mucosa of the oral cavity, in order to exclusively or mainly cleanse, scent or change their appearance or influence bodily odours or protect them or maintain them in good condition.” (29). The Regulation also states that the producer shall produce and have available a dossier which describes “The physico-chemical and microbiological specifications for the raw materials and the finished product and the purity and microbiological control criteria of the cosmetic product”. This is in accordance with EU legislation (30). Although the legislation is rather general the European Commission has several scientific committees to provide more detailed guidelines and opinions. The DG Health and Consumer Protection’s Scientific Committee on Consumer Products has issued Notes of guidance for the testing of cosmetic ingredients and their safety evaluation, currently in its 6th revision. These “Notes of Guidance” should not be seen as a checklist, but have been compiled to provide assistance in the complex process of the testing and safety evaluation of cosmetic ingredients. In its chapter 6-4: Guidelines on microbiological quality of the finished cosmetic product it is – among other requirements – specifically stated that *Pseudomonas aeruginosa* must not be detectable in cosmetic products (31). In addition it recommends challenge testing with *Pseudomonas aeruginosa*, *Staphylococcus aureus* and *Candida albicans* to test the efficacy of the preservation. However, this challenge testing does not take into account the altered microbiological properties of biofilm formation as described in chapter 1.8 of the thesis. It is also worth noting that the requirements for microbiological purity and product documentations are stricter for cosmetics and for comparable pharmaceutical products than for medical devices, class I. The producer of the Dent-O-Sept swab as well as the national health authorities agreed on classifying the Dent-O-Sept swab as a medical device, class I and not a cosmetic product.

Figure 2. The Dent-O-Sept swab, a non-invasive medical device in Class I.



1.6.1. Use of medical devices in hospitals and its control

The industry of medical devices is big business. According to the European commission when it launched the revision to the Medical Device Directives in 2005, there are some 10 000 types of medical devices from some 7 000 business entities in Europe employing upwards of 350 000 Europeans in the EU. The products range from simple bandages and spectacles, through life maintaining implantable devices, equipment to screen and diagnose disease and health conditions, to the most sophisticated diagnostic imaging and minimal invasive surgery equipment (32).

There are no national statistics on the use of medical equipment in Norway. A network organisation supporting suppliers to the health sector (Leverandørforeningen for helsesektoren) has indicated that the total sales of equipment to the hospitals amounts to 8-10 billion NOK annually where approximately half the sum concerns medical devices.

Infection control personnel in hospitals have indicated increasing complexity of highly advanced, technical machinery, including ventilators. Many of them are difficult to clean and disinfect and some are not possible to check whether cleaning and disinfection have been effective. The personell also claim an increasing use of single-use equipment all over the

western world. Many of these devices are expensive and for some years there has been an international debate on reprocessing of expensive single-use devices for reuse.

According to national regulations all hospitals must have written guidelines on the purchase and control of medical devices (33). It is recommended that infection control personnel take part in all purchases of medical devices when relevant for infection control. The purchasing process is not always performed according to guidelines in most hospitals.

The producer, seller, owner, and user of medical devices are all obliged to report errors of medical devices according to previous and current legislation (25). Depending on type of equipment and type of error (product error, electrical error, error concerning radiation, error in use of product) there are different reporting systems. The efficiency and completeness of the reporting systems have been questioned (34). For years there has been ongoing work to improve the error reporting systems from the health care services.

1.6.2. Contamination of medical devices

Medical devices may be contaminated in each of the steps from ingredients and building materials, through production, packing and transport to storage, use and reuse of the final product (Table 2).

Table 2. *Critical areas in the production and use of medical devices; problems and possible solutions.*

Area	Problem	Solutions
Ingredients, building materials	Any of the components used to make up the product or which are used in the production process can contain microbes. When introduced into the production process the microbes can establish and remain without new introductions.	Risk assessment and microbiological quality control of all identified components used in the production. When the final product is sterilised, this is less relevant.
Production facilities	The facilities can be contaminated from any of the components, from personnel or other environmental factors during the production	Risk assessment and microbiological quality control of all identified critical points. When the final product is not sterilised, microbiological quality control of the final product is warranted.

Area	Problem	Solutions
Packing	Packing material and environmental conditions (e.g. moisture) during the packing process may contaminate the outer surfaces.	Risk assessment and microbiological quality control of the final, packed product.
Transport and storage	Packaging and products can be damaged and contaminated during transport and storage.	Control of product and packing upon arrival and before use. Control of any product declarations and expiry dates.
Use	Devices can be contaminated during use, and whether it was sterile or not at commencement, bacterial growth can occur.	Risk assessment of critical points. Implementation of preventive measures to reduce risk. Follow user guidelines.
Reprocessing of reusable devices	Errors can occur in any of the steps of cleaning, disinfection and sterilisation of reusable devices. Many devices can be difficult to reprocess due to ruffled surfaces, small lumina, unreachable inner areas etc.	Follow detailed general and specific guidelines. Microbiological quality control after the finished process. Surveillance and tracking systems for the use of the devices.

1.6.3. Outbreaks caused by medical devices

A range of microbes can cause outbreaks in hospitals. When an association with medical devices or environmental reservoirs is described, often the bacteria in question are Gram-negative water-prone bacteria (7, 35-73). *Pseudomonas aeruginosa* is often the causative agent of hospital outbreaks (35-73); other common bacterial sources are *Serratia marcescens* (65, 74-78), *Acinetobacter baumannii* and other *Acinetobacter* spp. (79-83), other species of *Pseudomonas* (84-87), *Stenotrophomonas maltophilia* (87, 88), *Burkholderia cepacia* (89-92), *Klebsiella pneumonia* (93) and enterococci (94).

Medical devices can either introduce the causative bacterium of the outbreak or also maintain the outbreak due to wrongful use of the product or faulty cleaning and disinfection procedures between patients (35-37, 41, 43, 46, 54, 57, 64-67, 72, 78, 86, 87, 89, 95-98). In other

outbreaks personnel and environmental reservoirs are important (38, 45, 51, 59, 60, 62, 99). Cross-colonisation and cross-contamination within hospitals has been documented and can maintain outbreaks for longer periods (47, 52, 56, 59, 61, 63, 71, 74, 81-83, 93, 94, 100-102). Liquid or moist pharmaceuticals can cause or maintain outbreaks (37, 85, 89, 90, 103) as can moist cosmetics and water (35, 42, 67, 70, 84, 92, 99, 103, 104). Even transplanted organs can transmit bacteria like *P. aeruginosa* and cause an outbreak (58).

Many outbreaks are linked to ICUs and ventilator treatment (35, 38, 42, 45, 47, 50, 53, 78-80, 88, 96, 99). *Pseudomonas aeruginosa* is the most common gram-negative bacteria causing ventilator associated pneumonia (VAP) (105). Oropharyngeal colonisation is important for the development of VAP (106) and oral care may prevent pneumonia (107). For many of the outbreaks reported no definite source introducing the bacteria into the hospital has been verified. Nonetheless, the outbreaks have been brought to an end following the introduction or enforcement of strict infection control routines regarding personnel behaviour, standard precautions, cleaning, disinfection and sterilisation of medical devices, usage of only sterile or high-level disinfected moist products and thorough disinfection of all moist environmental surfaces (36-38, 41, 49-51, 53, 54, 59, 61-63, 65, 66, 70, 71, 74, 77-79, 82, 84, 86, 89-93, 95, 96, 98, 99).

1.7. *Pseudomonas aeruginosa*

The name *Pseudomonas* meaning “false single units” was given to this group of bacteria when detected in the late 19th century in water. *Aeruginosa* means “copper rust” and denotes the blue-green pigment seen in laboratory cultures.

1.7.1. Microbiology

Pseudomonas aeruginosa is a gram-negative, non-spore-forming, rod-shaped bacterium with one polar flagellum. It is almost exclusively aerobic, have minimal nutritional requirements and can utilise carbon from a variety of sources. In the laboratory it is easily identifiable (4-6).

P. aeruginosa produces several virulence factors. Polysaccharides and lipopolysaccharides serve as a barrier between the cell wall and the external environment and form a basis for the biofilm that the bacteria can produce. The bacteria also produce pigments that can act as virulence factors, and different exotoxins and proteases (5). In addition, *P. aeruginosa* produces several signal molecules, important for biofilm formation (108-112).

Pseudomonas is naturally detected in a variety of environments like soil, water, plants and animals, including humans. The bacterium has a predilection for moist environments.

Consequently, in humans it is usually detected in moist areas like the ear, perineum and axilla. Likewise it is detected in moist areas of the hospital, like sinks, taps, mops, water containers, humid medical devices, medicines, food, and in any non-sterilised water.

1.7.2. Epidemiology and clinical infection

P. aeruginosa is often characterised as an opportunistic bacterium which denotes that it rarely causes infection in healthy humans but may do so following disruption of physical barriers and in patients with certain underlying illnesses. Outside the hospital setting, skin infections, especially after skin burns and external otitis in frequent swimmers are the most common clinical manifestations (4, 5, 7).

In hospitals the clinical *P. aeruginosa* infection often reflects the patient's underlying diseases. In addition to bacteraemia and endocarditis, infection of the urinary tract, respiratory tract, central nervous system, ear, eye, bone, joints and skin are most often reported (4, 5, 7, 113, 114). Immunocompromised patients are vulnerable for infections in most body sites. Burns and disruption of the skin barrier can cause severe infections of the skin. Burn wound infections progressing to septicaemia are in $\frac{3}{4}$ of patients caused by *P. aeruginosa* and have a case fatality rate (CFR) of more than 50% (5). In patients receiving mechanical ventilation, other patients in ICUs and other patients with manipulation of airways *Pseudomonas* pneumonia is common. Patients with cystic fibrosis have an especially high risk for chronic colonisation and infection of the airways. Biofilm formation is an important factor in disease persistence for this patient group (115). *Pseudomonas* septicaemia and UTIs are also common clinical manifestations of *P. aeruginosa* infections.

Pseudomonas species is ranked among the top ten causes of bacteraemias in hospitals (116-121). In-hospital crude case-fatality from invasive disease is high, ranging from 18% to 61% (113, 122-132). In a Norwegian single-hospital study of bacteraemia in patients with malignant blood diseases, *P. aeruginosa* ranks number five in frequency as causative agent and these patients had a CFR of 21% within 30 days after bacteraemia diagnosis (133). *Pseudomonas* species other than *Pseudomonas aeruginosa* infrequently cause infection (6).

1.8. Biofilm formation

Bacteria exhibit two distinct modes of behaviour, a planktonic mode with free floating single bacteria and as a biofilm where the bacteria appear as structured communities (108, 111, 134). Most of our knowledge about bacteria stem from studies on planktonic bacteria. Studies in

recent years indicate that biofilm is an important – if not the most important – mode that bacteria appear in.

P. aeruginosa is well known to form biofilms (111, 134, 135). Biofilms are structured, specialised communities of adherent microorganisms encased in a complex extrapolymeric substance matrix (134) which can form on any surface although some surfaces are known to retard adherence (111). When a biofilm is formed and reaches a critical mass the quorum sensing molecules excreted alter many of the functions of the bacteria, including slowing its metabolism and increasing the production of a glycocalyx matrix (108, 112). These and other factors reduces the bacteria's susceptibility to antibiotics and disinfectants (111, 135). It has been shown that *P. aeruginosa* can reappear after biofilms on polyvinylchloride pipes have been exposed to a variety of disinfectants for seven days (136). To eradicate the viable bacteria in a biofilm, heat is preferred. Alternatively mechanical removal or the use of oxidative biocides to slowly dissolve the biofilm matrix (135) are suggested. Once a biofilm has formed and matured it can spread to new locations either through single cell dispersal or the shedding of clumps of biofilm (111, 112, 134).

1.9. Molecular typing methods

In epidemiology and outbreak investigation support from microbiological investigations often is essential in order to delimit an outbreak and determine who is a case and who is not. It is not sufficient to be able to identify *Pseudomonas* spp. or *Salmonella* enteritidis in patients to determine whether they are part of the same outbreak. More specific methods are needed. This is similar to criminal investigations by the police where different so-called fingerprint methods are used to link persons to locations.

In classical microbiology phenotypical and serotypical methods are used to differentiate between bacteria of the same species. Most of these methods are technologically simple and the results easy to compare between laboratories, but the methods are time consuming due to much manual work. Bacteria can further be differentiated on the presence of toxins and virulence factors and on antibiotic resistance patterns (5, 6).

The genome of bacteria has certain areas that are conserved within a species and certain areas that show different degrees of variability. The degree of variability varies considerably between bacterial species. To be able to distinguish between bacteria one needs to identify areas of the bacterial genome that are sufficiently variable to be able to discriminate between

clones, but not too variable so all bacteria appears different. The different molecular typing methods are developed to fit the specific properties of the different bacteria.

There is a whole array of molecular typing methods used for bacteria. All are based on the same principle, which is to identify and extract specific areas of the bacterial genome, amplify them and display them in ways to be able to compare the different bacterial samples.

For *Pseudomonas aeruginosa* mainly two molecular typing methods were used in the period when the Dent-O-Sept outbreak occurred. One is called pulsed-field gel electrophoresis (PFGE) where the bacterial genome is amplified, cut and spread in an agar by an electric field creating bands of different sizes in a specific pattern (137). By visually comparing, one can determine the degree of similarity between isolates. Due to uncontrollable differences in the processing between laboratories, it is not advisable to compare results between different laboratories. Another disadvantage of the method is that it is labour-intensive.

The other method commonly used is amplified fragment length polymorphism (AFLP). AFLP uses restriction enzymes to cut genomic DNA (deoxyribonucleic acid), followed by ligation of complementary double stranded adaptors to the ends of the restriction fragments. A subset of the restriction fragments are then amplified using two primers complementary to the adaptor and restriction site fragments. The fragments are visualised on denaturing polyacrylamide gels either through autoradiography or fluorescence methodologies (38, 101, 138, 139) .

In investigations of hospital outbreaks different molecular typing techniques are commonly used to distinguish cases from non-cases. The first challenge is to identify the best typing method for the microbe in question. Another is to define the genotypical criteria for including and excluding a bacterial isolate among the cases. There may be some variability among the bacteria at the onset of the outbreak and the bacteria may change during the course of the outbreak due to random mutations, antibiotic pressure, transfers of plasmids between bacteria etc. Most of the outbreaks referred to in chapter 1.6.3 have used one or more molecular typing methods to define the cases belonging to the outbreak and to link the cases to sources among medical devices, other equipment, the environment and personnel.

1.10. Causality

In complex situations many factors influence each other and it is rare to find simple cause-effect episodes like “the person died (effect) because he was shot through the heart (cause)”.

There is a definite association between smoking and lung cancer, but not a one-to-one relationship. Not all smokers contract lung cancer and not all with lung cancer have smoked.

The philosophical basis of the dominant approach for testing theories in medicine is the hypothetico-deductive model as described by for example David Hume and Karl Popper. According to this model it is impossible to achieve absolute proof for a scientific hypothesis; tests performed can only corroborate or falsify the hypothesis. Consequently one can never prove causality between factors and an outcome, only strengthen or weaken a proposed association. In this tradition, Sir Austin Bradford Hill listed nine viewpoints from which to study the association of two variables in order to claim causation (140).

Classic epidemiology has been mainly backward looking, seeking an explanation to an event. In much of the 19th century there was a profound debate on what caused many of the major diseases of the time, being it miasmata (stench or bad air) or contagions (141). For a disease like cholera, John Snow, the father of epidemiology, was in favour of the theory of a contagion which he called "morbid matter" (142). Late in the 19th century, a prominent German microbiologist, Robert Koch, formulated a set of postulates that needed to be fulfilled in order to claim that a micro-organism caused a specific disease (143, 144). According to his postulates we need both necessary and the sufficient conditions to claim causal relationship between a microbe and a disease.

1.10.1. Counterfactual theories

A century later MacMahon stated that there are two ways of classifying ill persons, either by *manifestational criteria* (grouping ill persons according to symptoms or clinical signs, e.g. common cold, schizophrenia or meningitis) or by *causal criteria* (grouping ill persons with respect to a specified experience believed to be a cause of their illness, e.g. lead poisoning or meningococcal disease) (145). To have a *Pseudomonas aeruginosa* infection implies by name and definition causality of the bacterium.

The central question in counterfactual theories of causation is "What would have happened if not event c had happened?" And the answer is: "If not event c had occurred, then the event e would not have occurred" (146). Counterfactual reasoning can be used both in deterministic and probabilistic models. In daily life and in medicine counterfactual reasoning is extensively used. "If the needle hadn't been contaminated, the patient would not have acquired hepatitis." "If you hadn't been exposed to asbestos, you would not have contracted mesothelioma." "If the producer had adhered to the regulations, the outbreak would not have occurred." Many of

the epidemiological study designs have counterfactual thinking embodied (147). In cohort studies we compare exposed and unexposed individuals for a certain risk factor. The unexposed group can be viewed as “what if this exposure did not occur”. When calculating the attributable risk fraction, also called the etiological fraction, we assume that all association between the exposure studied and the outcome is causal, and in addition imply that if not the exposed group had been exposed, the rate of outcome among them would have been at the same level as among the unexposed.

1.10.2. Determinism and probabilism

Determinism is based on the idea that every event is necessitated by antecedent events and conditions together with the laws of nature (148). According to causal determinism the causal relationships are invariant: Every time a certain configuration of conditions occurs, the outcome will be the same. We may have causal determinism even if the situation is complex and the outcome is hard to predict. Probabilistic causality on the other hand claims that the causal relationship is probabilistic, and not invariant. That is, the outcome (effect) may vary according to probability distribution. Probabilistic theories of causation state that causes raise the probabilities of their effects (149).

In epidemiology, probabilistic approaches are most often used in the conceptual thinking of a relationship and in the statistical testing of the strength of association (149). Here, Hill’s set of nine viewpoints to explain the association between two variables are commonly used (140). Only the one of temporal sequence of association is essential. This list of “Guidelines for causation” is more in tune with modern epidemiological science as they emphasise the strength of association rather than pure mechanical determinism. However, many have criticised Hill’s list and in recent years there has been a resurgence in the debate about causality (150-154). Often in communicable disease epidemiology, including outbreak investigation, it is useful to apply Hill’s nine viewpoints to assess association. However, with the advent of modern microbiological methods where one can detect genotypically identical strains of a bacterium in different locations and thereby more or less confirm the association, the other viewpoints play a lesser role.

2. Background and outline of thesis

2.1. Background about the outbreak

In late February 2002, the NIPH was alerted to a possible increase in the number of *Pseudomonas* infections in the clinical wards of Norwegian hospitals, especially in ICUs (Table 3). Infection control personnel in different hospitals had vague impressions of seeing more *Pseudomonas* infections than normal. On 8 March 2002, investigators at St. Olavs Hospital in Trondheim, Norway, discovered genotypically identical strains of *P. aeruginosa* in patient samples from two hospitals in different regions, and 10 days later, they discovered a genotypically identical strain from a third hospital in yet another region. We launched a national outbreak investigation. In retrospect we have created a timeline or log over the main events:

Table 3. Time-line of the main events in the Dent-O-Sept case.

Time	Event
1977 or 1978	Production of the Dent-O-Sept swab started
1995	New regulations on medical devices made legal for Norway. In order to CE mark a medical device, the producer needs to make a declaration of conformity.
1999	External evaluation of the production of the Dent-O-Sept swabs after complaints about discoloured swabs. The producer complied with some, but not all of the recommendations.
10.04.2000	The producer was certified by an independent body according to the standard NS-EN ISO 9002, 1994.
20.11.2000	The first patient with the <i>P. aeruginosa</i> later to be identified as the outbreak strain was tested.
12.04.2001	The first patient with <i>P. aeruginosa</i> later to be identified as the outbreak strain in blood culture was tested.
17.09.2001	The first swab to be detected contaminated with the outbreak strain was produced this week.
01.11.2001	A rapid increase in new cases with the outbreak strain started.
Nov. 2001	Some clinicians in hospitals started to question whether they were seeing an increase in <i>Pseudomonas</i> infections, especially in ICUs.

Time	Event
27.02.2002	Telephone from a doctor at the hospital in Stavanger to NIPH where she told us about a perceived increase in <i>Pseudomonas</i> infections and requested us to enquire other hospitals about it.
28.02.2002	E-mail from NIPH to all regional centres in hospital infection control and prevention where we ask whether they have noticed an increase. Quick preliminary answers from most regions.
08.03.2002	St. Olavs Hospital in Trondheim detected genotypically identical strains of <i>P. aeruginosa</i> in hospitalised patients from Stavanger and Trondheim.
12.03.2002	General alert of possible outbreak in MSIS-rapport, a newsletter from Norwegian Institute of Public Health.
18.03.2002	The outbreak strain detected in a third hospital, Ullevål in Oslo.
21.03.2002	The outbreak strain detected in a fourth hospital, Ahus, outside Oslo.
21.03.2002	Trawling questionnaire sent to all hospitals with patients with the outbreak strain.
02.04.2002	Telephone from the infection control nurse at Feiringklinikken. They had sent a discoloured Dent-O-Sept mouth swab for microbiological analysis and <i>Pseudomonas</i> sp. was detected. The lab had discarded the culture.
03.04.2002	E-mail sent to all involved hospitals to check the Dent-O-Sept swabs.
08.04.2002	At 5.18 PM, mail from St. Olavs Hospital: A genotypically identical strain of <i>Pseudomonas aeruginosa</i> was detected in Dent-O-Sept swab produced week 47 in 2001.
09.04.2002	All relevant parties were notified. A press conference was held. Large media attention. Production of the Dent-O-Sept swab ceased permanently.
10.04.2002	The Directorate for Health and Social Affairs asked orally NIPH to perform a mapping of all aspects of the outbreak.
12..04.2002	The Directorate for Health and Social Affairs organised a system audit of the manufacturer. The police started an investigation of the producer.

Time	Event
25.04.2002	The minister of Health gave an orientation on the outbreak of <i>Pseudomonas</i> infections in the parliament.
14.06.2002	The Norwegian Board of Health asked NIPH formally and specifically to perform a mapping of the total extent of the outbreak.
28.06.2002	NIPH sent formal letters to all medical microbiological laboratories about the outbreak mapping.
02.09.2002	The police had finished the investigation of the producer and decided not to press charges and closed the case.
01.10.2002	Internal review report by the Ministry of health on the roles and responsibilities of the central health administration in the fields of medical devices, discrepancy report systems and infection control
09.12.2002	The Directorate for Health and Social Affairs appealed the police's decision not to press charges. The time limit for this kind of appeals is three weeks; hence the Attorney-General could not reopen the case.
21.08.2003	Preliminary report from NIPH on the outbreak investigations of patients.
17.01.2004	Final reports from NIPH, The Directorate for Health and Social Affairs and The Norwegian Board of Health.
18.02.2004	Status report from The Norwegian System for Compensation to Patients on the monetary claims and their assessment.
11.10.2005	Out-of-court settlement where the producer agrees to pay The Norwegian System for Compensation to Patients 1.2 million NOK without accepting any responsibility for the outbreak.
19.06.2006	Court settlement between the producer and one large hospital (Ullevål) where the hospital received a compensation of 3.3 million NOK for additional costs incurred.

2.2. Setting

Norway had a population of 4.5 million people in 2001-2002 and approximately 65 (mainly public) hospitals organised in 5 public regional health trusts, each of which had a centre for hospital infection control. The 22 medical microbiological laboratories in the country provided general bacteriological culturing services. There was no national surveillance system

for *P. aeruginosa* infection. There were around 1000 health care institutions for the elderly. Through the European Economic Area Agreement Norway abides by much of the legislation within the EU, including European Council Directive 93/42/EEC concerning medical devices (26).

The main data for this study was collected in 2002-2003 during and after an outbreak of *P. aeruginosa* infection in hospitals.

Most of the research for this study took place at the NIPH during and after the outbreak investigation. NIPH is a governmental non-regulatory institute mandated to conduct surveillance of infectious diseases (epidemiologically and microbiologically) and advice the health services and the public on prevention and control of infectious diseases. As with other outbreak investigations NIPH collaborated extensively with all affected parties: infection control teams and administration in the hospitals, the regional health trusts' centres for hospital infection control, the medical microbiological laboratories, the local Food Safety Authority, some nursing homes and municipal medical officers, some private persons, the national and county Board of Health, the Directorate of Health and Social Affairs and the Department of Health.

2.3. Outline of the thesis

In Chapter 1 I give a general introduction, high-lighting some of the special features of HAIs and outbreak investigation in a hospital setting, medical devices, the bacterium under study and also give a general introduction to the discourse on causality in epidemiology which became important in the public debate of the outbreak.

In Chapter 3 I list the aims of the study and in Chapter 4 I present the data sources and methods used. The main results are presented in Chapter 5. And in Chapter 6 the findings are discussed and strengths, weaknesses and limitations of the methodology used are put under scrutiny. Finally, in Chapter 7 I reiterate the main conclusions, suggest further studies and propose what actions that should be taken from the lessons learned.

3. Aims of the thesis

All parts of this thesis are in some ways related to a large outbreak of *Pseudomonas aeruginosa* infection in Norwegian hospitals that occurred in 2001-2002.

The overall aims were to investigate a large outbreak of *Pseudomonas* infections and gain knowledge from it, to explore theories for causality and responsibility, and to describe the epidemiology and investigate risk factors for contracting invasive *Pseudomonas aeruginosa* infection.

When an infectious disease outbreak of this size occurs in hospitals large resources are spent on the outbreak investigation. In addition, experience is gained and knowledge is acquired in many related areas. The organisational structure and information channels are tested, as are the guidelines and regulations, the behaviours and routines, and the ability of all parties involved to respond to the emergency occurring.

3.1. Investigating an outbreak of *Pseudomonas aeruginosa* infections

When an outbreak occurs the overriding goal is to stop the outbreak to minimise the damage. The specific aims were to

- describe the outbreak,
- identify risk factors for contracting the disease among the patients,
- identify the causes of the outbreak, and
- make recommendations for the prevention of future outbreaks.

3.2. Investigating contamination of the medical device

When the outbreak strain of *P. aeruginosa* was detected in a medical device, it was concluded that this device was the vehicle introducing the bacterium into the hospitals. All aspects of this device, its production and use, was explored and assessed. The specific aims of this part of the investigation were to

- examine how *Pseudomonas aeruginosa* contaminated the product,
- assess the extent of the contamination by *P. aeruginosa* and other microbes, and
- identify critical points in the production process that made the contamination possible.

3.3. Exploring theories for causality of an outbreak of *Pseudomonas aeruginosa* infections

The outbreak received heavy mass media attention at times. Many patients had fallen ill, many died while hospitalised. In the public debate differences of opinion among stakeholders

were visible, politicians took part (an orientation was given in the parliament), large economical resources and even work places were at stake.

With all these actors and acts and different agendas, it was difficult to see who did what and which role and responsibility each participant had. The literature on causality is plentiful and diverse. The specific aims were to

- examine theories of causality from different fields like science, philosophy and law,
- apply the theories on the various participants in the outbreak to examine their role, and
- discuss the responsibility and fallibility for two of the main actors.

3.4. Investigating the epidemiology of invasive *Pseudomonas aeruginosa* infection

Pseudomonas aeruginosa can cause serious disease in susceptible patients even when there is no outbreak. *Pseudomonas* species is ranked among the top ten causes of bacteraemia in hospitals (116-121). In-hospital crude case-fatality from invasive disease is high, ranging from 18% to 61% (113, 122-132). There is extensive knowledge on *P. aeruginosa* infections in general and especially from tertiary care hospitals. But little is known about the epidemiology of invasive *P. aeruginosa* infection in humans in unselected hospitals, and even less from Norway as these infections are not covered by any national surveillance systems. The specific aims of this part of the study were to

- describe the epidemiology of invasive *P. aeruginosa* infection in Norway,
- identify patient groups at increased risk of disease and of death from *P. aeruginosa* infection, and
- estimate national incidence rates and mortality rates of *P. aeruginosa* infections by groups of underlying diseases.

4. Materials and methods

4.1. Investigating an outbreak of *Pseudomonas aeruginosa* infections

Following the alert of a possible increase, the NIPH immediately launched a classical outbreak investigation as outlined in Chapter 2.4: Infection control personnel in hospitals were alerted, preliminary epidemiological and microbiological investigations were performed, environmental sampling were performed, preliminary, and later definite case definitions were made. When the outbreak strain of *P. aeruginosa* was found in a product, systematic sampling of all available batches of the product was carried out. The production plant was inspected and sampled.

The Ministry of Health and its subordinate institutions, the Norwegian Board of Health and the Directorate for Health and Social Affairs, requested the detection of all patients involved in the outbreak. A systematic protocol was developed and approved. The outbreak strain of *P. aeruginosa* was identified. All available clinical, bacterial isolates were genotyped and compared with the outbreak strain. For each patient with the outbreak strain or with another strain of *P. aeruginosa* isolated from blood or CSF samples, the patient's physician were asked to fill in a detailed questionnaire.

We conducted a case-control study to investigate risk factors for having the outbreak strain of *P. aeruginosa* as compared with other strains of *P. aeruginosa*. We did not look for risk factors for having *P. aeruginosa* infection. Data obtained in the descriptive epidemiological investigation was used. To be able to pick comparable controls the source population was defined as only those with invasive *P. aeruginosa* infection. Case patients were persons with the outbreak strain isolated from blood or CSF samples during the period October 2001–December 2002, and control subjects were all the persons included in the study with genotyped strains of *P. aeruginosa* other than the outbreak strain isolated from blood or CSF samples during the same period.

To investigate risk factors for a fatal outcome during the stay in the institution for patients with an invasive *Pseudomonas* infection, we used a cohort design including all of the patients in the case-control study. The same variables as in the case-control study were included as possible risk factors for death, in addition to having the outbreak strain.

To evaluate whether the *Pseudomonas* infection had contributed to the death of individual patients, Bjørn G. Iversen and Preben Aavitsland meticulously assessed all of the available information for each of the dead patients. Among the information assessed was the course of

events for each patient, dates for hospital admission, diagnosis, transfers and death, underlying illnesses, clinical and microbiological information and the clinicians' assessment of a relationship between *P. aeruginosa* infection and death. Due to a large degree of variability between how the different clinicians assessed the relationship, we did not follow their assessments in all instances.

4.2. Investigating contamination of the medical device

Detailed environmental investigations were carried out for the product, the production facility and for the moist ingredients of the product.

When the outbreak strain of *P. aeruginosa* was found in a product, systematic sampling of all available batches of the product was carried out. Up to 10 items of each available batch of the product were asked to be examined. We asked the laboratories to identify and deep freeze monocultures of all findings of certain microbes whereas others were only to be noted and reported.

The Directorate for Health and Social Affairs organised a system audit of the manufacturer on 12 – 15 April 2002 by studying documents, interviewing selected personnel and inspecting the premises, including microbiological sampling which were cultured at the municipal Food Control Authority. On request from the producer, the laboratory at the municipal Food Control Authority performed environmental sampling in addition to what had been performed during the system audit.

Microbiological analysis (155, 156) were performed on each of the ingredients for the moisturising liquid used in the product (except tap water). The total viable aerobic count and specific detection of *P. aeruginosa* were tested in each of the liquids. Then the moisturising liquid undiluted and in 1:10 dilution were tested for their effect on the outbreak strain and a reference strain of *P. aeruginosa* (ATCC 9027 -MicroBioLogics) (157).

4.3. Exploring theories for causality of an outbreak of *Pseudomonas aeruginosa* infections

An analytical approach in the tradition of philosophy of science, and not using a strict epidemiological methodology, was used to discuss causality in the outbreak. Firstly, theories of causality from different disciplines were introduced. Then the *P. aeruginosa* outbreak was used as a case and the different theories were applied. The different theories of causality were put under scrutiny and the roles of the many actors involved were elucidated. Two actors are

especially central in this outbreak and their roles were further discussed to examine their responsibility and fallibility in the outbreak.

4.4. Investigating the epidemiology of invasive *Pseudomonas aeruginosa* infection

We used all information on patients with *P. aeruginosa* or *Pseudomonas* not identified at the species level (*Pseudomonas* spp.) isolated from blood or CSF during the period 1992-2002 collected during the outbreak investigation. We described the whole cohort and we analysed in detail the patients from the recent years (1999-2002), about whom we had collected much more information. Denominator data was collected from a variety of public sources. Population statistics and the number of beds in municipal nursing homes were downloaded from Statistics Norway. The number of stays and the number of days of hospitalisation in somatic hospitals by region, age and discharge diagnoses were supplied by The Norwegian Patient Register. Several disease categories that have been shown to be associated with increased risk of invasive *P. aeruginosa* infection were selected and grouped according to ICD-10 (International Classification of Diseases, 10th Revision) (158).

Descriptive and analytical epidemiological methods were used, and incidence proportions of infection and death for various groups were calculated.

4.5. Data management and statistical analysis

In the data collection, extensive efforts were made to ensure completeness and quality checks of data. Participating microbiological laboratories were given lists of all information they had sent NIPH to check for completeness of records and to fill in missing values and to correct improbable values. Hospitals were contacted by mail and phone to remind them of missing clinical forms and to check missing or improbable values and unreadable text.

Data were checked manually and electronically by listings, cross tabulations and by calculating time between admission, diagnosis, discharge and death. Some patients had been admitted to more than one hospital, and double entries were checked and removed. All patients were checked with an updated National Population Registry to search for deaths.

Paper I

All data were entered into an Epi Info software database, version 6.04d (Centers for Disease Control and Prevention), and analysed data using Epi Info (Centers for Disease Control and Prevention) and Stata 8 (Stata) statistical software. For the case-control study, odds ratios (ORs), 95% confidence intervals (CIs), and *P* values were calculated. In the multivariable

logistic regression analysis, all risk factors were initially included in the model, and the ones with the highest *P* values were removed one by one, until only variables with *P* values <0.05 remained. The variables that remained in the model were assessed and statistically tested for effect modification. In the cohort study examining risk factors for death, a similar binary regression approach was used.

Paper IV

We entered all patient data in an Epi Info version 6.04d database and analysed them in Epi Info, Excel, Episheet and Stata 8 and 9 statistical software. Incidence proportions, incidence rates and a comparison of these (risk ratios and rate ratios of different kinds) with 95% CIs were calculated in Episheet and Stata. To identify risk factors for dying among the cases we performed stepwise multivariable binomial regression analyses in Stata.

4.6. Laboratory analysis

Primary culturing of product and patient samples was performed at local laboratories. Culturing of samples of the product was performed at local laboratories according to our instruction: “Brush the swab against both a lactose and blood agar dish in a rotating manner so all sides of the foam tip touches the agar. It is not necessary to place the swab in a growth broth”. The isolates were identified by standard procedures in use by the laboratories.

Culturing of samples from the system audit and the additional investigation of the production site were performed at the laboratory of the municipal Food Control Authority. The qualitative analysis of the samples was performed by direct seeding (except for dry Dent-O-Sept swabs) and seeding after enrichment overnight in a heart infusion broth on Kings Agar B and on blood agar. The quantitative analysis was performed by direct seeding of 0.1 mL undiluted or – if heavy contamination was expected – diluted liquid on Kings Agar B and for some samples also on blood agar. The plates were incubated at 37°C overnight before reading.

All available isolates of *P. aeruginosa* from patients, product and from the system audit (but not from the additional investigations of the production site) were sent to at least one of five reference laboratories for genotyping and comparison with the outbreak strain (*P. aeruginosa* found in the product batch 47.2001 on 8 April 2002). The reference laboratories reported whether the isolate belonged to the outbreak strain or not. Four of the reference laboratories used a protocol developed at St. Olavs Hospital for a PFGE method. The criteria of Tenover

et al. (137) were used to interpret identical (no band differences) or closely related (≥ 3 band differences) isolates. The fifth laboratory used an AFLP method. The method for genotyping by AFLP is slightly modified from a method described elsewhere (38). Isolates that displayed $\geq 85\%$ similarity were considered to be closely genetically related and to belong to the same clone.

The AFLP and PFGE protocols were compared and found to be equal in detecting and discriminating the outbreak strain. If an isolate was not typeable by PFGE because of excessive activity of endogenous endonucleases, it was genotyped with AFLP.

4.7. Ethics

All public health work has to balance between the rights of the individual and the benefits for society. This is particularly evident in prevention and control of communicable diseases. A person with a communicable disease can have his freedoms restricted in order to protect the society at large. For example, a person with open pulmonary tuberculosis needs to be confined to an isolation room as long as he is contagious to prevent further spread, preferably voluntarily, if necessary by force. The Norwegian Communicable disease control act is build up around these principles (23). There was no need to isolate any patients with *P. aeruginosa* by force.

One regulation to this act is on the surveillance of communicable diseases and immediate notification of serious events and outbreaks. The immediate outbreak notifications shall not contain person identifiable information. When conducting an outbreak investigation it sometimes is necessary to obtain confidential information quickly in order to control the outbreak and minimise the harm and where seeking approval may seriously delay the investigation. This is acceptable and in accordance with consequentialist ethical thinking. The National Committee for Research Ethics in Norway concludes that prior approval is not needed as long as the objective is to control the outbreak (159). Then it is also in accordance with deontological ethics.

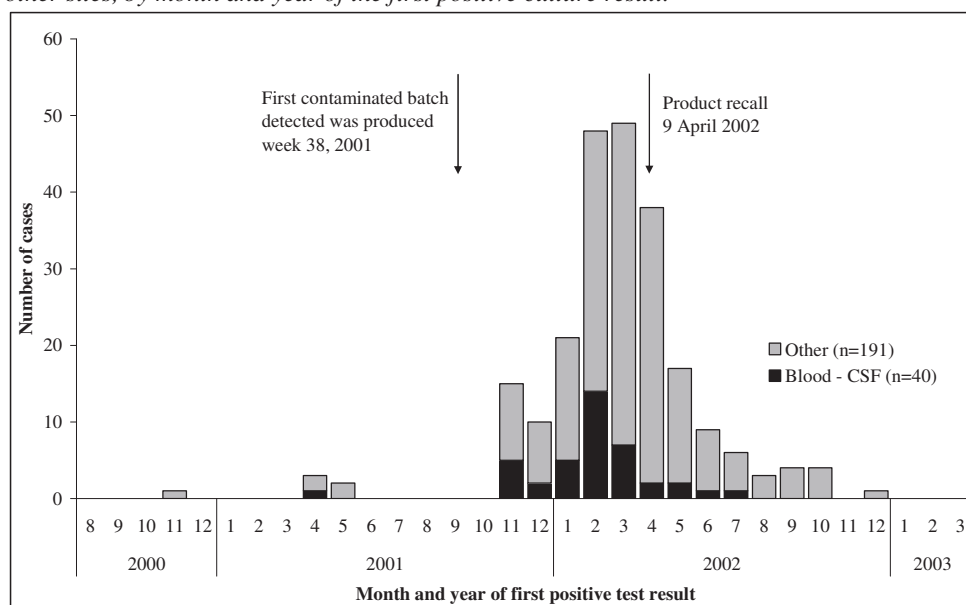
When the outbreak investigation moves beyond the need to control the outbreak, further approval is needed, hence the NIPH was authorised by the Norwegian Board of Health to perform the study. And the Data Inspectorate gave permission to create a database to store the information.

5. Synopsis of the results of the study

5.1. Investigating an outbreak of *Pseudomonas aeruginosa* infections

The outbreak strain of *P. aeruginosa* was isolated from 231 patients from November 2000 through December 2002, with a peak incidence during February–March 2002 (Figure 3). The patients with the outbreak strain were hospitalised at 24 different hospitals in all public regional health trusts (range, 1–39 patients per hospital), whereas 5 lived in other institutions, and 3 were not hospitalised when they received a diagnosis. The median age was 65 years and 61% were men. Of the 231 patients 39 had positive blood culture results. Clinically 42 had sepsis, and 87 had pneumonia, whereas 70 patients were only colonised. Altogether, 156 patients were admitted to an ICU during their hospital stay, and 128 received mechanical ventilation within the previous 3 weeks before receiving a diagnosis of *Pseudomonas* infection.

Figure 3. Epidemic curve of the outbreak showing the number of patients cases with the outbreak strain of *Pseudomonas aeruginosa* isolated from either blood or CSF sample or other sites, by month and year of the first positive culture result.



Seventy-one patients (31%) died while institutionalised; all of the patients who died had severe underlying disease. An assessment of whether the *Pseudomonas* infection contributed to the patient's death concluded that it was probable for 34 patients, improbable for 21,

uncertain for 13, and impossible to evaluate because of a lack of information for 3. A total of 132 patients (69%) with the outbreak strain had definitely or probably used the Dent-O-Sept swab, whereas 58 (31%) had not or probably had not used it.

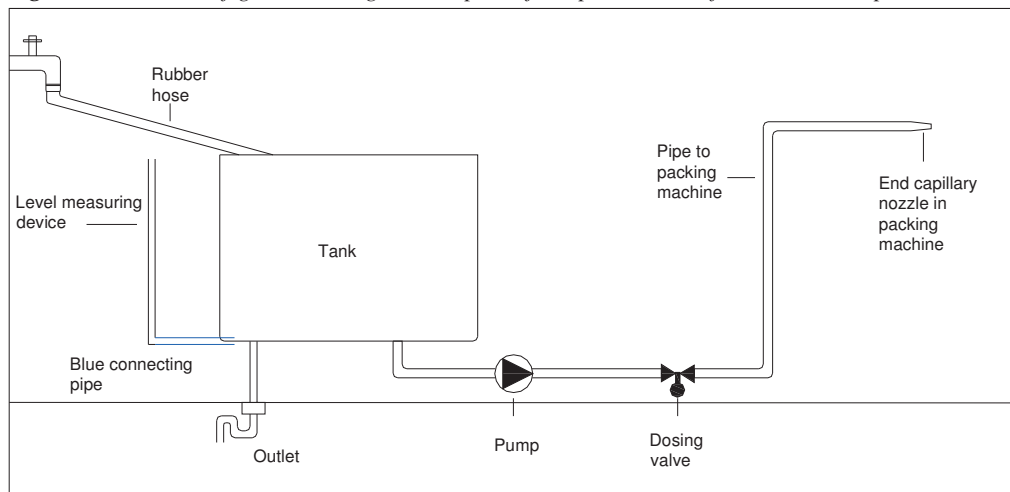
Among 39 case patients and 159 control subjects, use of the moist mouth swab (adjusted OR 5.3; 95% CI 2.0–13.6) and receipt of mechanical ventilation (adjusted OR 6.4; 95% CI 2.3–17.2) were associated with infection due to the outbreak strain.

5.2. Investigating contamination of the medical device

NIPH received information about stored batches of the Dent-O-Sept swab from 59 general hospitals, four other health care services and 20 private persons. A total of 1565 swabs were examined from 149 different batches. Although we asked for up to 10 swabs of each batch to be examined, an average of 18 swabs per available batch were examined for the years 2001 and 2002, ranging from one to 37 swabs per batch. The outbreak strain of *P. aeruginosa* was detected in 76 swabs from 12 different batches of the Dent-O-Sept swab produced from week 38 in 2001 to week 15 in 2002 when production ceased. All genotyped strains of *P. aeruginosa* were identical to the outbreak strain. In total, more than 250 swabs were found to contain one or more species of microorganisms, mainly gram-positive bacteria which were predominantly discovered in the earlier batches. Gram-negative rods including *Acinetobacter baumannii* were isolated in swabs produced in 1999 and 2001.

During the system audit and the additional investigations of the production facilities samples for microbiological examinations were taken from several places along the production line. The outbreak strain of *P. aeruginosa* was detected from the end capillary nozzle in the packing machine (Figure 4). In the additional investigation *P. aeruginosa* (which were not genotyped) were cultured from the blue connecting pipe, the level measuring device and a rubber hose.

Figure 4. Schematic figure showing the wet part of the production of the Dent-O-Sept swab.



The system audit concluded that the production deviated from the existing regulations in several areas:

- The production process, including the recipe for Dent- O-Sept, did not ensure that the product had the qualities and properties stated by the producer nor that the risk of contamination was avoided or reduced to a minimum.
- Neither the boxes nor wraps of the Dent-O-Sept gave the user the necessary information. The CE (Communauté Européenne) marking was unjust because the producer's declaration of conformity with the regulations, including the risk analysis, was poorly based and documented. The technical documentation did not give a third party a basis for assessing whether the device was in accordance with the demands of the regulations.
- The producer did not comply with the obligation to report defects and deficiencies in medical devices to national health officials and had not adequately followed up errors in the production demonstrated in an external review in 1999.

No bacteria were detected in any of the ingredients for the moisturising liquid. When the outbreak strain of *P. aeruginosa* was added to the Dent-O-Sept solution and to the two concentrations of the disinfectant we observed a 6 log reduction in 15 minutes and for the 1:10 diluted Dent-O-Sept solution a 6 log reduction after 3-6 hours. For the reference strain (ATCC 9027) there was a 6 log reduction in 15 minutes for all four liquids.

5.3. Exploring theories for causality of an outbreak of *Pseudomonas aeruginosa* infections

The *P. aeruginosa* outbreak was used as a case and various theories of causality from different disciplines (epidemiology, other sciences, philosophy and law) were applied to discuss the roles and responsibilities of some of the parties involved. Mackie's concept of INUS conditions, Hill's nine viewpoints to study association for claiming causation, deterministic and probabilistic ways of reasoning, all shed light on the issues of causality in this outbreak. Moreover, applying legal theories of causation (counterfactual reasoning and the "but-for" test and the NESS test) proved especially useful, but the case also illustrated the weaknesses of the various theories of causation.

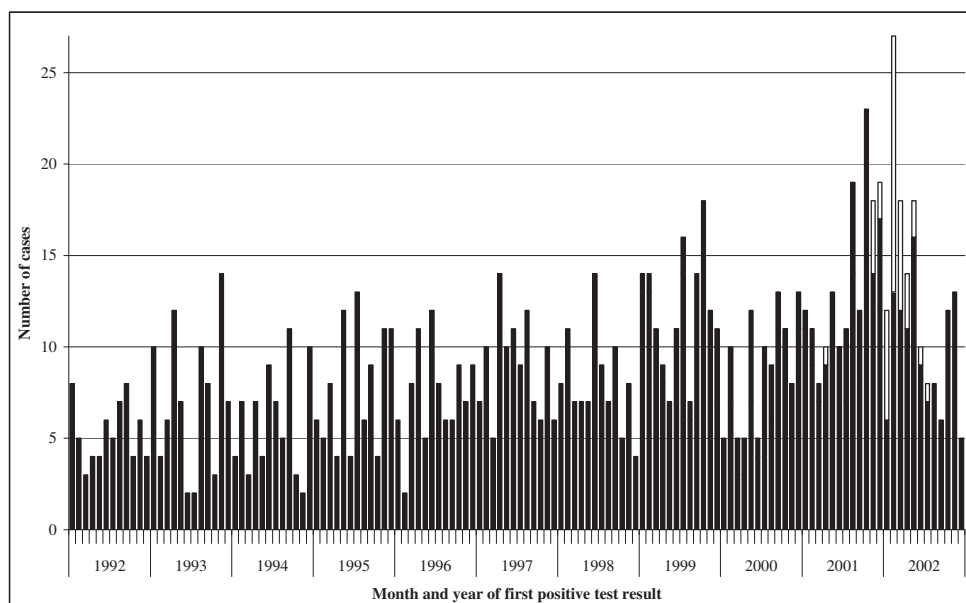
We concluded that many factors contributed to causing the outbreak, but that contamination of a medical device in the production facility was the major necessary condition. The reuse of the medical device in hospitals primarily contributed to the size of the outbreak. The unintended error by its producer – and to a minor extent by the hospital practice – was mainly due to non-application of relevant knowledge and skills, and appears to constitute professional negligence. Due to criminal procedure laws and other factors outside the discourse of causality, no one was criminally charged for the outbreak

5.4. Investigating the epidemiology of invasive *Pseudomonas aeruginosa* infection

In the 11 year period 1992-2002, 1174 patients with invasive disease were identified, of which 1079 (92%) had isolates of *P. aeruginosa* and 95 had *Pseudomonas* not identified at the species level (*Pseudomonas* spp.), resulting in an overall incidence rate of 2.43 per 100 000 person-years at risk (pyar) (Figure 5). The median age of the patients was 72 years and 67% were male.

For the period 1999-2002, 567 incident cases (representing 565 patients) were identified, corresponding to an incidence rate of 3.16 per 100 000 pyar (95% CI, 2.90-3.43). In hospitals the incidence rate was 3.33 per 100 000 person-days (95% CI, 3.06-3.62), or 0.20 per 1000 hospital stays (95% CI, 0.18-0.21).

Figure 5. The monthly number of cases of invasive *Pseudomonas aeruginosa* infection in Norway 1992-2002. Forty cases (white bars) belonged to an outbreak caused by a contaminated mouth swab.



The rate of infection was much higher in males, 5.1 per 100 000 person-days in hospital as compared with 1.9 in females (incidence rate ratio [IRR], 2.6; 95% CI, 2.2-3.1). For hospital-acquired cases the rates were 2.9 and 1.0 per 100 000, respectively (IRR, 3.0; 95% CI, 2.3-3.8).

A total of 55% of the cases were hospital-acquired and an additional 10% were living in or being hospitalised from a nursing home. The remaining 34% were community-acquired and 1% was unknown. For hospital-acquired infection the rate was 671 per 100 000 person-years as compared with 1.13 for community-acquired infection, and 37 in nursing homes.

Of the patients with invasive *P. aeruginosa* infection, 14% received mechanical ventilation within a period of 3 weeks before the sample positive for *P. aeruginosa* was taken, and 30% were admitted to an ICU during their stay in the hospital.

The highest risk for invasive *P. aeruginosa* disease was found in patients with malignant neoplasms of lymphoid and hematopoietic tissue (ICD-10 categories C81-C96) (risk per 1000 hospital stays 1.9; 95% CI 1.5-2.3) and other diseases of blood and blood-forming organs (D70-D77) (2.2; 95% CI 1.2-3.7). The CFR was 35%. The highest CFR was seen among

patients with transplanted organs (Z94) (67%) and patients with certain diseases of the respiratory system (J10–J22; J40–J96) (52%).

In the multivariable regression analysis, the following variables were independently associated with an increased risk of dying (CFR) while hospitalised among the cases: Having one or more underlying risk diagnoses (risk ratio [RR], 2.52; 95% CI, 1.50-4.25), admission to an ICU at some time during the stay (RR 1.55; 95% CI, 1.26-1.91), being 60 years or older (RR, 1.53; 95% CI, 1.14-2.06), and having received immunosuppressive treatment within the past three weeks before the sample with *P. aeruginosa* was taken (RR, 1.39; 95% CI, 1.13-1.73). Having a *Pseudomonas* UTI as the most serious clinical *Pseudomonas* diagnosis was highly protective (RR 0.09; 95% CI, 0.01-0.63).

6. Discussion

The outbreak of *Pseudomonas aeruginosa* infection in 2001-2002 was a major wake-up call for many groups in the society, including hospital managers, hospital infection control personnel, national health service administration, politicians in the parliament and elsewhere and producers of medical devices. Prior to the outbreak few would have believed that a seemingly inconspicuous mouth swab which had been produced for decades, could have caused serious disease in so many patients.

The outbreak reminds us of the change in the patient population in hospitals over the last decades with an increasingly larger part being severely ill and susceptible for an increasing number of opportunistic bacteria. Consequently infection control becomes increasingly important in hospitals.

Questions of causality, responsibility and blame have always been a part of the history of infections. During and after the outbreak investigation, questions of causality, responsibility and liability were raised: Who and what caused the outbreak, who were responsible for the extent of the outbreak, could the damages have been mitigated by acting sooner or differently, should anyone be punished?

The outbreak prompted action in many areas. All actors involved analysed their situation, wrote statements and reviews, a national action plan against HAIs were published; regulations and guidelines for infection control and prevention in hospitals were revised (3, 25, 33, 34, 160-166).

6.1. An outbreak of *Pseudomonas aeruginosa* infections

The *Pseudomonas aeruginosa* outbreak in Norway 2001-2002 is possibly the largest published outbreak of its kind to date with 231 confirmed cases, 161 of whom had a clinical infection. A total of 71 (31%) of the patients died while in an institution. All of those who died had severe underlying illnesses such as terminal cancers, multiple traumas, or severe respiratory or vascular disease. The cause of death was multifactorial for all these patients. To decide whether the *P. aeruginosa* infection contributed to the patients' deaths can only be based on a best judgement as one has to assess all the different factors and estimate the impact of each factor. In an assessment based on all available information of whether the *Pseudomonas* infection contributed to the patient's death, we concluded that it was probable for 34 of the 71 patients, improbable for 21, uncertain for 13, and impossible to evaluate

because of a lack of information for 3. In conclusion at least 161 patients had a clinical infection and 34 died prematurely as a consequence of the outbreak.

The outbreak received much media attention, especially when the link to the Dent-O-Sept swab was documented. Afterwards the attention gradually subsided. Later Norway has experienced other large national outbreaks. In 2006 there was an outbreak with *E. coli* infection causing haemolytic uremic syndrome (HUS) (167). The outbreak – where 10 of 17 cases had HUS as a clinical manifestation and one child died – received much more intense media coverage than the Dent-O-Sept outbreak. In 2005, a legionellosis outbreak in the county of Østfold with 56 confirmed cases and 10 deaths received less media attention than the HUS outbreak but more than the Dent-O-Sept outbreak from an subjective point of view at NIPH (168). There may be several reasons for this. One is a behaviour change of the media being increasingly aggressive and when an event catches on, journalists try to outdo each other in digging up minute details and blowing them out of proportions. Increasingly frequent the media coverage of outbreaks escalates to a level involving the national government and the parliament. The other main reason can be that whereas the Dent-O-Sept and legionellosis outbreaks to a large extent involved elderly and already ill people, the HUS outbreak mainly struck children.

But for the small outbreaks or when the source is obvious, outbreak investigations are both resource and time consuming. When investigating a large, multicentre outbreak like the Dent-O-Sept outbreak it is important to create a flexible central team and a robust network reaching all involved parties. With many participants one of the challenges is to let all voices be heard but at the same time make all pull in the same direction once a matter has been discussed and a decision reached. Not to loose sight when drowned in details, it can be wise to have one or a few “opponents” to evaluate the progress of the outbreak investigation at some distance in order to suggest corrections if needed.

During the *P. aeruginosa* outbreak most of the communication between the central outbreak investigation team at NIPH and the local teams in the hospitals and the laboratories were by e-mail. This was the first large outbreak investigation at NIPH where e-mail was the main mode of communication, and it proved to be quick and very resource efficient. On the day when the source of the outbreak was verified the head investigator sent and received a total of 33 e-mails which was an immense amount in 2002 (but not today). The active use of postings on the NIPH website on the Internet was still in its beginning and would have been used more extensively today. The cooperation between all parties but a few exceptions was exemplary.

Only one local hospital outbreak team worked counter-actively in periods by creating an alternative trawling questionnaire to the one the others had agreed on and sending it to some of the other hospitals, requesting extra information on questionnaires to neighbouring hospitals in addition to what the central investigation team had collected and thereby placing extra burdens on them. At the height of the outbreak investigation we participated in several meetings with the top administration, including the managing director, of this hospital.

Most hospital outbreaks are spread locally via persons, local products or the environment. When three and steadily more hospitals were involved the attention was directed towards a moist product mainly used in the ICU. However which product was difficult to elucidate as there is a huge array of moist devices and pharmaceuticals used in the ICU. A systematic trawling and testing of products had started when an infection control nurse tipped us about discoloured Dent-O-Sept swabs that had yielded *Pseudomonas* sp. but where the bacterial culture had been discarded before it was examined further.

Similar to criminal investigations, genetic fingerprinting has become an important means of connecting cases to sources and causes. We were able to detect genotypically identical bacteria from patients, the product and the production facility thereby beyond doubt confirming the source of the outbreak. An increasing number of outbreaks are investigated with the help of microbiological genotyping (45, 60, 66, 88, 169). In addition, these methods are often used in the case definitions to separate the patients with the outbreak strain from other patients with infections with other strains of the same microbe. Genotyping of the many isolates of *P. aeruginosa* has been an expensive but indispensable part of this outbreak investigation.

6.2. Contamination of the medical device

Medical devices may be contaminated in each of the steps from ingredients and building materials, through production, packing and transport to storage, use and reuse of the final product.

Some, but not all of the Dent-O-Sept swabs were contaminated. How the *P. aeruginosa* bacteria came into the production equipment was never ascertained. One plausible hypothesis is that it entered via the municipal drinking water. Drinking water is not – and is not required to be – sterile. Moisture-prone bacteria like *P. aeruginosa* will naturally occur in municipal drinking water. The misconception of the purity of drinking water is also seen in the health

care setting where tap water is used in areas where only high-level disinfected or sterile water should be used.

In the production facilities the cleaning and disinfection routines of the production equipment did not eradicate the bacteria. And there was no quality control system in place checking the microbiological quality of the final product. This was one of the recommendations given during an external evaluation and not followed up (162).

However, other actors in addition to the producer were not abiding by regulations and guidelines. Many health care institutions found their logistic systems for purchase, storage and use of the swab to be deficient. Examples of reported deficiencies were: no centralised procedure for purchases, infection control personnel not taking part in the purchasing process and boxes of old batches were found in remote storage places. Many also lacked adequate reporting systems for faulty medical devices.

Anecdotal information from several hospitals described that, after use, nurses would store the swab in a glass of tap water on the night stand and later reuse it for the same patient. The extent of this practice is unknown but where it occurred, the bacterial load these patients were exposed to may have increased exponentially as indicated in a report (170). The risk of becoming colonised or infected increases with the dose of bacteria the patient is exposed to.

When a microbe is introduced into a hospital setting for example via a medical device, it may contaminate or colonise patients, personnel, the environment or other medical equipment. Approximately 1/3 of the cases in the *P. aeruginosa* outbreak had probably or definitely not used the swab. And in addition to having used the swab, receipt of mechanical ventilation was an independent risk factor for harbouring the outbreak strain of *P. aeruginosa* compared with having another strain of the same bacterium. We present two possible explanations for this:

1. Injury to the tracheal epithelium may favour adherence of *Pseudomonas*, and *P. aeruginosa* is commonly found in ventilator associated pneumonia (4). When the outbreak strain repeatedly was introduced from the swabs, environment or persons it could easily lead to colonisation or infection in susceptible, ventilated patients.
2. The ventilators themselves posed as a risk implying that the cleaning and disinfection of the ventilators between patients did not sufficiently eradicate the bacteria once it had been introduced. The problem of adequately disinfecting all parts of the ventilator or other medical equipment is well known from other outbreaks (64, 66, 80, 98).

Typical areas prone to *P. aeruginosa* contamination are sinks, faucets, flasks, other containers and tubes; in short, any moist area. When the bacterium first is introduced it needs hardly any nutrition to survive or even to multiply. Many containers with ordinary tap water are left to stand for too long periods without checking, cleaning and disinfection. Health care institutions need to perform risk assessments of all moist environments and to institute guidelines for microbiological quality control of these environments and guidelines for when only to use sterile water.

6.3. Microbial control of moist products

Where there is water there are microbes if not the water is completely sterile. And if bacteria are present they will multiply. A bacterium like *P. aeruginosa* has affinity for moist environments and has minimal nutritional requirements (4, 7). There are several ways of controlling microbial growth in moist products. The first way is to sterilise the product with heat, gas or radiation. For products that are exposed to the environment, microbes will eventually enter the product and some sort of preservative is essential. In cosmetics and pharmaceuticals a range of preservatives have been used to control growth, of which parabens and benzoic acid and its chemical derivatives are common. In foodstuffs salt, sugar, acids, alcohols and gaseous environments are also commonly used. Most disinfectants intended for use as technical disinfection are too toxic to add to these moist products. There are few moist medical devices on the market that does not need to be sterile.

All preservatives and other additives to these kinds of moist products are strictly regulated by law and categorised. For example, all approved food additives in the EU are listed in the Codex alimentarius and given an E-number. All preservatives are E200-E299, and sodium benzoate, for example is E211.

Medical devices: In Norway laws and regulations of medical devices (24, 25) are for all practical purposes identical with those of the EU, including European Council Directive 93/42/EEC concerning medical devices (26). For non-invasive medical devices in Class I, there are no demands for sterility. The devices must, when used, “not compromise the clinical condition or the safety of patients”. “The devices and manufacturing processes must be designed in such a way as to eliminate or reduce as far as possible the risk of infection to the patient, user and third parties.” Beyond this, the directive does not specify the microbial quality of the product, e.g. the absence of *P. aeruginosa*. And without any assistance from a third party the producer can draw a declaration of conformity. The responsibility for control

of medical devices is with the Directorate of Health and Social Affairs (now called the Directorate of Health).

Pharmaceutical preparations: All medicinal products are strictly regulated and needs to be approved by the Norwegian Medicines Agency or a counterpart in another country in the EU in order to receive marketing authorisation (171, 172). According to the European Pharmacopoeia, pharmaceutical preparations for use in the respiratory tract are classified in a Category 2 where – in addition to other microbiological requirements – the absence of *Pseudomonas aeruginosa* needs to be documented (27).

Cosmetics: As for the products mentioned above, cosmetics and body care products are regulated through an Act and a regulation (28, 29). As for medical devices the legislation is almost identical with that of the EU (30). The Regulation states that the producer shall produce and have available a dossier which describes: “The physico-chemical and microbiological specifications for the raw materials and the finished product and the purity and microbiological control criteria of the cosmetic product”. The appendices to the regulation give examples of groups of cosmetics and body care products and list which preservatives can be used. The “Notes of guidance” giving detailed recommendations to the Council Directive states that *Pseudomonas aeruginosa* must not be detectable in cosmetic products (31). The Norwegian Food Safety Authority is responsible for the control of cosmetics.

There seems to be less specific demands for microbiological purity for non-invasive medical devices in Class I compared with other moist products. In their final report the Directorate of Health and Social Affairs stated the need to discuss whether non-sterile, moist medical devices should be reclassified to a higher class than Class I (164, 165). The issue was thoroughly debated at meeting for the authorities on medical devices from the Nordic countries in February 2004. The other Nordic authorities were quite reluctant to reclassify these products. Their main argument was that if the manufacturer had adhered to the current regulations and classifications the outbreak would not have occurred.

The area of medical devices has not been prioritised for resources in the central health administration. This is one of the clear conclusions the Ministry of health drew in an internal review after the outbreak (34). Compared with the other Scandinavian countries, Finland and Great Britain, Norway had spent a lot less resources on the administration and control of medical devices. The resources allocated had mainly been used on controlling medical

devices of higher classes and controlling the technical control organisations. Only on direct enquiries the Board of Health (and later the Directorate of Health and Social Affairs) had followed up on Class I medical devices.

6.4. What preservatives were used in the Dent-O-Sept moisturising liquid?

Two of the major questions after the outbreak were: What preservatives in the moisturising liquid suppressed bacterial growth and how could the bacteria survive in the production facilities and in the individual wrapped swabs?

One batch of moisturising liquid consisted of: Tap water (147 litres), 96% ethanol (3 litres), Glycerol (16 litres) and Vademecum, a commercially available mouth rinse (6 litres). The main ingredients of the mouth rinse are ethanol (44%) and sodium benzoate (5.25%) in addition to water. Of the three major possible preservatives the final concentration in the Dent-O-Sept moisturising liquid was calculated to be 2.3% ethanol; 9.3% glycerol and 0.18% sodium benzoate (173). The producer had used ethanol as an antimicrobial agent, glycerol as a moisturiser and Vademecum for taste and comfort.

It is believed that ethanol and glycerol in such low concentrations have little documentable bacteriostatic effect. Sodium benzoate on the other hand, has antimicrobial effect in concentrations starting from 0.01%. However, the effect is largely dependant on the acidity of the solution, being at its maximum at pH 2.5-4.0 and weak above pH 4.5. In an expert report made for the Directorate of Health and Social Services the acidity measured in a limited number of swabs was approximately pH 7.0 (173). We remade a small batch of the moisturising liquid and measured a pH of 6.8 (174). At this pH level sodium benzoate has minimal antibacterial effect.

In a controlled test two different strains of planktonic *P. aeruginosa* were added to the moisturising liquid. The bacteria were rapidly inactivated even in a 1:10 dilution of the liquid. So far it has not been elucidated what ingredients in the moisturising liquid that have the rapid bactericidal effect on *P. aeruginosa*. Nonetheless, the moisturising liquid inactivated planktonic bacteria even at a pH level of 6.8. Still the outbreak strain of *P. aeruginosa* survived in the production plant and in a number of wrapped swabs.

Several hypotheses have been put forward to explain this apparent discrepancy. Our main hypothesis is biofilm formation. *P. aeruginosa* is well known to form biofilms (111, 134, 135). Bacteria in biofilms have an increased ability to withstand antibiotics and disinfectants

(111, 135). We have shown that the cleaning and disinfection process in the production line probably did not reach all areas in the tank and piping system. A mature biofilm may spread to new locations through single cell dispersal or the shedding of clumps of biofilm (111, 112, 134). Such clumps still have biofilm properties and may well have survived on a wrapped swab surrounded by a liquid with antimicrobial effect.

An alternative hypothesis has been launched in the popular and medical press (175-179). This suggests that the pH may have risen in the liquid inside the wraps due to the chemical influence either from the glue used to attach the foam rubber heads to the stick or other materials inside the wrap. According to this hypothesis an increase in the pH may have lowered the antimicrobial effect of sodium benzoate. It is an interesting hypothesis. However it has one major flaw: The pH of the moisturising liquid and in different approximate remakes of it has been measured to be in the range of 6.4-7.7, and never below 6.4 (174, 178). In this range sodium benzoate has minimal preservative effect and pH alterations within this range do not alter sodium benzoate's bactericidal ability.

In conclusion, we still do not know which ingredients in the Dent-O-Sept liquid that are exerting its rapid bacteriostatic effect. It cannot be singly ethanol, glycerol or sodium benzoate in the concentrations and pH range of the mixed liquid. Whether they may have the tested bacteriostatic effect on planktonic *P. aeruginosa* by working in consort or if there are other unknown preservatives in the Vademecum mouth wash has not been tested. Out of pure curiosity it would be interesting to find the answer. However, the production of Dent-O-Sept swabs ceased in 2002 and any new producer of moist swabs would need to produce a declaration of conformity for their product where the issue is addressed.

6.5. Medical devices as a source of infection

The quantity of published hospital outbreaks is almost innumerable. A quick Internet search on PubMed searching for "hospital" and "outbreak" yielded 9384 articles. In the winter season epidemics of norovirus infections can be rampant as can influenza epidemics and outbreaks with the spread of Methicillin resistant *Staphylococcus aureus*.

Medical devices and other equipment, solutions and liquids are responsible for a considerable number of outbreaks. Often they are related to moisture in one way or another.

Pseudomonas aeruginosa is the dominant causative agent for outbreaks in hospital among the gram-negative opportunistic bacteria (35-37, 39, 41-44, 46, 49, 50, 53-55, 57-59, 64-67, 70, 72, 84, 85, 99, 104, 180-183). Most are caused by medical devices directly or indirectly but

some are related to liquids like flush devices, hydrotherapy, bath toys or even bottled drinking water to name a few (35, 37, 39, 42, 67, 70, 85, 99, 104, 181, 182). Outbreaks have even been caused by lens prosthesis implants (44) and after a multiple organ transplantation (58).

Of the other microbes related to medical devices and other equipment (74-76, 78-80, 86-90, 92, 95-98, 103, 105) many are like *P. aeruginosa* gram-negative bacteria like *Serratia marcescens* (65, 74-78), *Acinetobacter baumannii* and other *Acinetobacter* spp. (79, 80), other species of *Pseudomonas* (84-87), *Stenotrophomonas maltophilia* (87, 88), and *Burkholderia cepacia* (89, 90, 92).

All outbreaks are unique in one or more ways regarding causative agent, mode of transmission, duration, extensiveness, size, consequences, liability, etc. In most of these aspects the Dent-O-Sept outbreak is not unique, encompassing *P. aeruginosa*, a medical device, moisture, hospital setting, ICU, and a mixture of direct transmission from the medical device and indirect via the hospital environment. However, the Dent-O-Sept outbreak is possibly the largest *Pseudomonas* outbreak ever to be published regarding the number affected (231 patients, 161 with infection, 70 colonised), the number of deaths (71 while hospitalised, and where the *Pseudomonas* infection probably contributed to the death for 34 of them) and the number of health care institutions involved (24 hospitals).

6.6. Claiming causality

In the scientific laboratory all variables are known, and the scientist can change one factor at the time and measure its effect on the outcome variables. Modern epidemiology is complex; a range of factors contribute to an outcome, many of which are unknown. And in outbreak investigations, the investigator cannot control the exposure variables because they have already occurred. In addition, there is a time constraint.

The basic concept of causality is a fundamental and integral part of daily language. However, when trying to define the term in philosophy (146, 148, 149), science (140, 144, 145, 147, 152-154, 184-186) or law (187-191), it becomes utterly difficult and complex to a degree that some even discourages the use of the term. Instead euphemisms like 'associated with', 'linked to', 'related to', and 'due to' are used signalling causation but to a somewhat weaker degree.

Tradition on causal theories differs between the different scientific disciplines. Applying the various theories on the major actors and events in the outbreak shed light from different angles and proved to be helpful in the analysis. Especially using counterfactual reasoning which is frequently used in tort law simplified the complex picture. Counterfactual theories

have been used in epidemiology where the term “the counterfactual ideal” has been coined to describe the perfect unexposed experience (192). However, it would be helpful to develop the counterfactual reasoning from tort law further in epidemiology. When used on the Dent-O-Sept outbreak the main actors responsible were easier to identify. Our analysis concluded clearly that the major necessary condition causing the outbreak was the contamination of the swabs in the production facility. Without this contamination, the Dent-O-Sept outbreak would not have happened. Many other factors contributed to the outbreak and the size of it, the reuse of the single use swabs in the hospitals being the most important.

Moral responsibility is related to legal responsibility. As mentioned earlier, outbreak investigations have similarities with police investigations. In large outbreaks, especially where people are injured or die, the police regularly start an investigation. With a clear causal association between the contaminated swabs and *Pseudomonas* infection and death among patients, it would have been interesting to see whether the conclusion would have been the same in a criminal court case.

6.7. Invasive *Pseudomonas aeruginosa* infection

For the period 1999-2002, we found a rate of invasive *P. aeruginosa* infection of 0.20 per 1 000 hospital stays. Studies from other countries have indicated much higher rates, between 0.94 and 1.8 per 1000 hospital stays (114, 120, 123-125, 127, 128, 130) in tertiary referral hospitals and university hospitals and 0.43 and 0.59 per 1000 hospital stays in community hospitals (118, 119). The low incidence in Norway compared with studies from other countries may partly be explained by study design. Our study was nationwide and population-based and included all somatic hospital stays in Norway. Certain hospital departments such as dermatology and gynaecology and obstetrics, and community or specialty hospitals with less than 5000 discharges per year, are known to have low rates of *P. aeruginosa* invasive infection so these departments may have contributed to the low reported overall rates. However, no Norwegian hospital had a higher rate than 0.42 per 1000 hospital stays.

To our knowledge only one other population based study on invasive *P. aeruginosa* infections has been published (193). This study based on the population of a county in Minnesota, USA in 1999-2006, has higher incidence rates than our findings with 10.8 patients per 100 000 pyar in men and 3.7 in women compared with 4.3 and 2.0 respectively per 100 000 pyar in our study.

We suggest that one explanation for the low rates was the prudent use of antibiotics in hospitals in Norway with low overall antibiotic consumption and low use of broad spectrum antibiotics. The use of narrow-spectrum drugs is encouraged (194-196). Indiscriminate use of antibiotics, especially broad-spectrum antibiotics, is known to be associated with development of resistance and selection of resistant bacteria, such as *P. aeruginosa* (4-7, 197).

The use of empiric broad spectrum antibiotics such as most 3rd generation cephalosporins, which may give *Pseudomonas* an advantage, are generally discouraged. For empiric treatment of septicaemia where broad spectrum coverage is necessary, penicillin plus an aminoglycoside is the recommended standard treatment in most hospital departments. In general, aminoglycosides are active against *Pseudomonas* and have a high threshold for development of resistance (tobramycin considered most active), thereby avoiding selection pressure favouring these bacteria (198).

Several risk factors for invasive *P. aeruginosa* infection and death have previously been identified (4, 5, 7, 113, 123, 126, 127, 129, 130, 199). Using complete national discharge statistics, we were able to calculate absolute rates of infection among patients with various underlying diseases. Our study confirms that *P. aeruginosa* infection is a major risk to patients with cancer, immunodeficiency, renal failure and certain other diseases.

Invasive *P. aeruginosa* infection is a serious disease with a high CFR, 35% in our study. Most patients who died from invasive *P. aeruginosa* infection died within a short time of being diagnosed. The patient groups with the following underlying diseases had the highest mortality risk per 1000 discharges: Malignant neoplasms of lymphoid and haematopoietic tissue, other diseases of blood and blood-forming organs, organ transplantation and renal failure. Among the patients with invasive *P. aeruginosa* infection the following factors were independently associated with an increased CFR: Having an underlying risk diagnoses associated with *P. aeruginosa* infection; admission to an ICU; old age and immuno-suppressive treatment prior to infection. A clinical *P. aeruginosa* diagnosis only of UTI was protective, whereas meningitis, pneumonia and septicaemia increased the risk although not significantly. Other studies list similar risk factors (123, 126, 127, 130). As demonstrated by others (124), bacteraemic pneumonia had a high CFR.

6.8. Methodological weaknesses and limitations

An outbreak investigation is getting the best possible results in a chaotic world with limited resources and under heavy time pressure. Once the source of the outbreak is detected and

removed, the time constraint is relieved and a more thorough collection of data can be performed. However, if it takes too long the interest of the collaborators may subside and the quality of data may weaken. For the Dent-O-Sept outbreak the source was detected 8 April 2002, the scientific protocol following the outbreak investigation was finalised in late June 2002 and the last information entered into the crude database in August 2003.

There are always four possible explanations to a finding: 1. it can be true; 2. random error; 3. bias; or 4. confounding. In addition, there is always a possibility that the study design is flawed or inadequate to answer the hypothesis raised, or that inferences drawn from the results are invalid.

6.8.1. Random error

Random error is the portion of variation in a measurement that has no apparent connection to any other measurement or variable, generally regarded as due to chance (9). It is the variability in the data that we cannot readily explain after the systematic error is eliminated. A confidence interval is used to indicate the amount of random error in the estimation (192). By convention most results in epidemiology are given with a 95% confidence interval. This means that if the study were repeated many times and in an identical manner, the confidence interval should include the correct measure 95% of the time, on the condition that there are not other errors. For all effect ratios presented in our studies the point estimate with a 95 % confidence interval were given to account for possible random error.

6.8.2. Bias

Bias is defined in several ways. The textbook definition is all deviations of results or inferences from the truths (9). This will include random error, confounding, effect modification, flawed design, prejudices and wrong interpretations. In practical epidemiology the term bias is usually restricted to two forms of bias, selection bias and information bias.

Selection bias is mainly a problem that affects case-control studies where it gives rise to non-comparability between cases and controls when cases are not representative of the population that produced the cases. Selection bias can also occur in cohort studies when completeness of follow-up or case ascertainment differs between exposure categories.

Information bias is a flaw in measuring exposure or outcome data. There are two types of misclassification: 1. Non-differential misclassification where the probability of exposure being misclassified is the same regardless of outcome status. This type of misclassification will principally weaken the measured effect. 2. Differential misclassification where the

probability of exposure being misclassified depends on the outcome status and vice versa. Differential misclassification can either weaken or strengthen a measured effect. There are two main types of differential misclassifications: 1. Recall bias which is typical in case-control studies where cases and controls remember exposures differently. 2. Observer bias where those collecting data classifies data differently for different groups under study.

Case-control-study

This study in Paper I to identify causes of infection with the outbreak strain was restricted to severe disease, i.e. where the bacterium had been isolated from blood or CSF. The advantage was that the case definition was clearer and we need not speculate whether for example a skin infection was merely a colonisation. However, by only selecting patients with the most severe disease, the analysis loses generalisability. That said, from a clinical and epidemiological perspective, it is more important to look for risk factors among the most severely ill patients.

The aim of the study was to identify the causes for infection with the outbreak strain as compared to being infected with other strains of *P. aeruginosa*. The aim was not to identify risk factors for *P. aeruginosa* infection by *any* strain. This aim had consequences for the choice of control group, namely patients with non-outbreak strain *P. aeruginosa* identified in blood or CSF. We reasoned that these patients had the same risk factors as cases for getting a severe *P. aeruginosa* infection, for instance immunosuppression. Thus, we avoided the cumbersome task of controlling for the unspecific concept “underlying disease”. In epidemiological terms, we defined the source population as patients with *P. aeruginosa* isolated from blood or CSF. If, on the other hand, we had defined the source population as all hospitalised patients and sampled controls among them, the analysis would have been biased towards severity of disease and not of having the outbreak strain. It would have been hard to differentiate the risk factors of invasive *P. aeruginosa* infection in general from risk factors for outbreak strain infection. Had controls been selected among patients admitted to the ICU, the analysis could have been biased the other way as only 2/3 of the case patients had been admitted to the ICU during their stay.

Choosing as cases patients infected with one certain strain or subtype of a microbe and as controls patients with other strains or subtypes has been named a case-case study (200). This design can be useful for communicable diseases where only a potentially non-representative fraction of the infected patients are identified and thus eligible for being cases. This is the situation for salmonellosis for example where most patients do not seek medical care. To control for this selection process of those identified with the disease, controls are chosen

among those identified with other subtypes of the same microbe and who consequently have gone through the same selection process. In essence the case-control study in Paper I is a case-case study where cases and controls differ only regarding the genotype of *P. aeruginosa*. However, our purpose was not to control for the selection process of the cases. In the hospital setting blood cultures are taken from almost all patients where invasive disease is suspected and the detection rate is high. Hence, few patients with invasive *P. aeruginosa* infection will go undetected. Our choice of the case-case variant of the case-control study was instead guided by the aim of the study, i.e. to identify the cause of the outbreak as efficiently as possible.

Microbiology

The main challenge in outbreak investigations is that of bias. It is crucial to detect all cases and to gather correct information on all of them which can be said to be a form of selection bias. The clear assignment from the Ministry and the politicians was to detect everyone who was colonised or infected with the outbreak strain of *Pseudomonas aeruginosa*. All stored cultures of *Pseudomonas* were genotyped. Laboratories had mainly stored cultures of *P. aeruginosa* from blood and CSF and in the retrospective part of the study we may have missed cases with the outbreak strain from other body sites. In the prospective part of the study we believe that a great majority of the cases were detected. For the whole period we do not know how many with the outbreak strain that were not sampled, especially outside hospitals. In conclusion the real number of affected patients was probably much higher. However, there is good reason to believe that most of the missing cases have only been colonised or had less serious infections. Consequently one can claim a selection bias towards more serious cases, but none of the studies intended to give a numerical distribution of the clinical presentation of *Pseudomonas* infection.

For the study of invasive *Pseudomonas aeruginosa* infection the same dataset for the cases was used. As the laboratories keep records and store samples from blood and CSF we believe few cases were missed. Some samples were not available for genotyping which could have influenced the comparison between the outbreak strain and the others.

Five laboratories genotyped the bacteria to identify the outbreak strain, four using the same PFGE typing method, the fifth AFLP. The methods had been compared in their ability to detect the outbreak strain and no discrepancies in the results were detected.

Environmental sampling

All initial environmental sampling from the production facilities and from the swabs were done by direct seeding on a lactose and blood agar dish. Without prior enrichment in a growth broth we may have some false negative results. This was indicated when the laboratory at the municipal Food Control Authority some weeks after the system audit performed repeated environmental sampling of the production facilities and used an enrichment broth. Then they detected the *Pseudomonas* bacterium in several places where the system audit had not (170). Whether to use an enrichment broth or not is a trade off between spending resources and increasing the sensitivity somewhat. More than 1500 swabs were examined in addition to patients and environmental samples.

Clinical information

For all included patients the clinicians were obliged to fill in a one page questionnaire; and we received it for all but two patients. Although great efforts were made to ensure that the outbreak investigation was given highest priority, the quality of the returned questionnaires varied. Extensive efforts were made to ensure completeness and quality of the data, including contacting the clinicians and linkage with the National Population Registry to search for deaths among patients. Consequently, we believe most of the collected patient data is accurate, but some information may have been missed, especially regarding some of the subordinate discharge diagnoses. However, all but 3 of the 567 patients had at least one main underlying disease recorded other than the *Pseudomonas* infection. There were few patients in several of the risk diagnosis groups (as listed in Paper IV, table 3). Missing information in either of these groups could have influenced the incidence and mortality rates and the case fatality, and the results need to be interpreted with caution. However, all numerators and denominators are given in the table, making it possible for the reader to judge the results.

Most of the variables are factual (e.g. whether the patient had been admitted to the ICU during the hospital stay) and are not much influenced by observation bias. However, one variable especially was vulnerable for subjective assessment: If the patient died, “May the patient’s death be related to the detection of *Pseudomonas* infection?”. To control for that two of the paper’s authors meticulously assessed all available information for each of the dead patients, including the clinician’s assessment, underlying illnesses, dates of onset, diagnosis and death, and other clinical and microbiological information. There was varying degree of missing values for the variables, most for the question asking whether the patient had used the Dent-O-Sept swab during the stay. This variable was also the one most subjected to observation

bias as most clinicians knew whether the patient had harboured the outbreak strain or not. However, for 1/3 of the patient with the outbreak strain the clinician wrote that that patient had not or probably not used the swab.

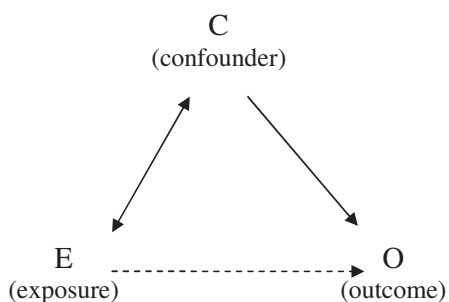
Denominator data

Denominator data in Paper IV were all collected from quality controlled national databases like Statistics Norway and The Norwegian Patient Register. There is always a challenge in combining data from different datasets as the variable definitions may vary. For the dataset in Paper IV it was most crucial for the definition of underlying disease. From the national register we picked the main and up to seven subordinate discharge diagnoses whereas on the clinician's forms there was free space to record as many as they wanted. No more than 10 underlying diagnoses were detected and recorded.

6.8.3. Confounding

The word confounding is derived from Latin *confundere* meaning to mix together (9). A confounder (C) is a variable that is associated with the exposure variable (E) and has an effect on the outcome (O) and is not an intermediate factor in the causal pathway between the exposure and outcome variables (figure 6). For example a measured association between coffee drinking and pancreatic cancer may be due to a higher proportion of coffee drinkers among smokers than among non-smokers. A reanalysis stratified by smoking status may show no association between coffee drinking and cancer in either group of smoking status. Then smoking status is confounding the first measured association. However, not all associated variables are confounders (201).

Figure 6. Association between exposure, outcome and confounder



Confounding can be controlled for in several ways: **1. Restriction** of the population or cases studied; **2. Matching** cases and controls on certain variables; **3. Randomisation** of study subjects in experimental-type studies; **4. Stratification** of the study population on the presumed confounding variables; and **5. Multivariable modelling**, a powerful technique to control for several possible confounders. The first three methods are study design strategies and the last two are analysis strategies.

Most of the results from the outbreak investigation are descriptive with no association between variables posed and no statistical analysis performed. As described above, all effect ratios are presented with a point estimate and a 95 % confidence interval to account for possible random error.

In Paper I we showed a clear association between the use of the Dent-O-Sept swab and having the outbreak strain of *Pseudomonas aeruginosa*. When controlling for known possible confounders the strength of association dropped from OR=7.9 to OR=5.3. There may also be other, unknown confounders that we did not control for. In addition, use of the Dent-O-Sept swab was the variable with the highest number of missing values in this case-control analysis (33 of 198 missing). If differential, it may introduce observation bias.

However, the association between having the outbreak strain and the Dent-O-Sept swab was also established in other ways. Genotypically identical strains of the bacterium were detected in patients and in Dent-O-Sept swabs (and in the production plant) and thereby unequivocally establishing how the outbreak strain was brought into the health care services. In complex settings there are usually more than one factor contributing to an outcome. In this outbreak approximately 1/3 of the cases had not used the swab. Consequently, indirect transmission in the hospital setting via health care workers, medical devices or the environment appears to be an important mode of transmission in this outbreak, although not tested specifically in the outbreak investigation.

In two papers we analysed risk factors for dying among patients with invasive *Pseudomonas aeruginosa* infection. In Paper I several factors were identified in the univariable analysis but only having used the Dent-O-Sept swab remained independently associated in the multivariable regression analysis. Logically it does not make sense that using a mouth swab would increase the risk of dying among patients with invasive *P. aeruginosa* infection. Consequently we interpreted the results as residual confounding where having used the swab served as a marker for severe underlying disease.

In Paper IV including a larger number of patients over a longer time interval (567 patients as compared with 198 in Paper I) and several new variables, more variables were found to be independently associated with an increased risk of dying in the multivariable regression analysis. This time, swab use was not independently associated with dying. One explanation for the difference can be controlling for the residual confounding by adding new clinical variables on underlying disease and clinical manifestation of the *P. aeruginosa* infection; another is larger inaccuracies in the recording of Dent-O-Sept swab use outside of the outbreak period.

In Paper IV, except for the analysis of risk factors for dying, only univariable analyses were performed because denominator data were aggregate data. Thus, we were unable to do individual level analyses of relative risks for infection, nor controlling for potential confounders by multivariable regression analysis. As age and gender are the two major possible confounders that influence the incidence risks, we may not easily compare the groups of underlying diseases. As explained above for the CFR we were able to control for confounders by binomial multivariable regression.

6.8.4. Effect modification

Effect modification – sometimes called effect-measure modification – refers to the situation in which a measure of effect changes over values of some other variable (192). Effect modification can be tested statistically by looking for interaction using statistical software programmes. However, eyeballing and thorough assessment is the best way to judge whether there is effect modification. In Paper I we assessed whether there was effect modification with the two variables “use of swab” and “receipt of mechanical ventilation” regarding the outcome of having the outbreak strain of *P. aeruginosa*. One may hypothesise that the presence or absence of swab use in mechanically ventilated patients influenced the variable “receipt of mechanical ventilation” as a risk factor for having the outbreak strain. We did not find clear indications for effect modification. However, one should bear in mind that the figures are small (Paper I, Table 3).

6.8.5. Analysis of causality

Paper III on causality is of a different genre than the other three. Rather than being quantitative and analysing figures it is qualitative and analyses and debates concepts and theories. Consequently there are no figures with confidence intervals but a discourse aiming at

elucidating the concept of causality through a real life example. In its nature the paper is more subjective.

7. Main conclusions and further studies

7.1. Main conclusions

In the outbreak with *Pseudomonas aeruginosa* in Norway, we detected a total of 231 patients with genotypically identical strains of the bacterium from 24 hospitals in 2000-2002. Seventy-one of the patients died while hospitalised, and for 34 the *Pseudomonas* infection probably contributed to the patients' deaths. The same outbreak strain was isolated from 76 mouth swabs called Dent-O-Sept from 12 different batches produced from September 2001 to April 2002 and from the production facility. In total more than 250 of 1565 examined swabs were contaminated with one or more microbial species.

In a complex situation like an outbreak with many acts, actors and factors playing larger or smaller parts, applying various theories for causality and responsibility from different fields like science, philosophy and law – especially legal theories and counterfactual reasoning – helped elucidating their roles and responsibilities. Many factors contributed to causing the outbreak, but contamination of a medical device in the production facility was the major necessary condition. The reuse of the medical device in hospitals primarily contributed to the size of the outbreak. In addition there were many errors and flaws in the chain from the production of the swabs, through purchasing and storage systems in the health care institutions to the use of the swabs and reporting of defective devices. The unintended error by its producer – and to a minor extent by the hospital practice – was mainly due to non-application of relevant knowledge and skills, and appears to constitute professional negligence.

This outbreak is possibly the largest published *P. aeruginosa* outbreak to date. Although *P. aeruginosa* usually do not cause infection in healthy persons, it frequently does in patients with certain underlying diseases, and in patients with disrupted barriers, especially in the ICU. Invasive *P. aeruginosa* infection is a rare disease with an incidence rate of 3.16 per 100 000 pyar or 0.20 per 1 000 hospital stays, but is very serious for those contracting it with a 30 day case fatality rate of 33%. Patients with malignant neoplasms of lymphoid and haematopoietic tissue and other diseases of blood and blood-forming organs have the highest risk of infection. Prudent antibiotic use is one possible explanation for much lower rates of infection in Norway compared with all other published studies from other countries.

Medical devices, moist equipment and solutions and moist environments are frequently associated with outbreaks with *P. aeruginosa* and related moisture-prone bacteria. Lack of

adherence to standard precautions for infection control and prevention by hospital personnel contributes to the propagation of these outbreaks.

Biofilm formation is possibly the more common of the two distinct modes of behaviour for bacteria; the other being the planktonic mode. Biofilm formation is the most plausible explanation for the survival of the bacteria in the production facilities and in the wrapped swabs. Bacterial biofilms are less sensitive to disinfection and make it more difficult to eradicate. Not abiding by the production regulations, e.g. the requirement to have quality assurance systems including an effective microbiological control system, made the contamination possible in the production process.

Outbreak investigations are essential to detect causes of an outbreak and to gain experience in order to prevent their recurrence. Investigating large, multicentre outbreaks is resource demanding and necessitates a defined network structure where everyone know their role and qualifications and try upmost to cooperate. Expertise in a variety of fields is essential. Molecular finger-printing techniques to identify the outbreak strain of the microbe and discriminate against other strains have become an indispensable part of most outbreak investigations.

7.2. Proposed actions and further studies

After the outbreak all parties involved, from producers and private companies to hospitals and all national, administrative bodies for health services, revised their guidelines, made reviews and summaries and proposed action plans. A national “Action plan to prevent hospital acquired infections” was made as a direct consequence of the outbreak (161) and has recently been revised (3).

Medical devices

- The control and audit of producers of medical devices should improve, also of producers of non-sterile medical devices in Class I. Increased resources should be allocated on a national level to support infection control personnel in health care institutions who have enquiries about medical devices.
- The reporting systems for errors in medical devices and other equipment and facilities in the health services are complex and cumbersome. A revision and coordination has started but is still not finalised.
- Moist medical devices are prone to cause infections and outbreaks if not properly manufactured and used. The Council Directive (26) may have been sufficient to prevent

the outbreak if it had been followed by the producer, but it is not optimal. Preservatives with documented effect should be obligatory in all moist, non-sterile medical devices, parallel to what exists for cosmetics and medicinal products.

- A systematic assessment should be made for the level of disinfection or sterility of all devices, other equipment, solutions, food, water and medicines to be used on different groups of infection-prone patients, especially in ICUs. Only documentable quality controlled, high-level disinfected products and items should be used in the oropharynx of susceptible patients.

Outbreak investigations

- Small outbreaks are not very resource-demanding and can in most instances be covered through regular budgets. Large outbreaks can be very costly. The process of securing financing for genotyping of the many hundred strains of *P. aeruginosa* was unnecessarily cumbersome. Streamlining of these processes needs to come in place.
- Infection control personnel in hospitals should have easy access to theoretical and practical training in epidemiology and outbreak investigation.

Infection control and prevention

National action plans to prevent HAIs list many admirable aims and measures and contain descriptions used on festive occasions. However, in contrast to several institutions on a national level, most hospital administrations do not give priority and resources to infection control and prevention. Infection control does not have a high status in the hospital hierarchy, is understaffed, and is not able to fulfil the requirements set up in laws and regulations.

- The four Regional Health Authorities and the administrations of the local Health Trusts need to review the new national strategy, revise their current infection control strategy plans and make detailed plans for the implementation of this new strategy.

Surveillance

There is no national surveillance system for *P. aeruginosa* and other similar opportunistic bacteria. And probably it will be unwise to include them in the current national system (202). Most hospitals have cooperation between infection control and the microbiology laboratories and some have local systems for reporting of microbiological detections in the hospital.

- An assessment should be done at the local or regional level of which microbes that are desirable to watch. Local systems in hospitals need to be developed for efficient surveillance of these microbes.
- On a national level automatic harvesting of data will be possible through the national health net system. When in place NIPH should in collaboration with the microbiology laboratories develop a system for surveillance of indicator microbes.
- One hospital has used a surveillance method from private industry called statistical process control (SPC) on the *Pseudomonas* outbreak to see if the outbreak could have been detected earlier (203). This method and others need to be developed further and implemented if effective.

***Pseudomonas* infections**

- More studies are needed on risk factors for infection caused by opportunistic pathogens among different groups of debilitated patients and how to prevent them from occurring.
- More studies are needed on bacteria forming biofilms and ways of eradicating them from environmental surfaces, medical devices and biofilm forming infections in humans.

8. References

1. Elstrøm P. Smittevern i helseinstitusjoner (Infection control in health care institutions). Oslo: Gyldendal Norsk Forlag; 2002.
2. Douglas M. Routledge & Kegan Paul, ed. Purity and danger: an analysis of the concepts of pollution and taboo. London: 1966.
3. Ministry of Health and Care Services. Nasjonal strategi for forebygging av infeksjoner i helsetjenesten og antibiotikaresistens (2008–2012) (National strategy to prevent infections in the health services and antibiotic resistance (2008-2012)) [Ministry of Health and Care Services]. [updated 2008; cited 2008 July 17]. Available from: <http://www.regjeringen.no/upload/HOD/Dokumenter%20FHA/Nasjonal%20strategi%20infeksjoner-antibiotikaresistens.pdf>.
4. Pollack M. *Pseudomonas aeruginosa*. In: Mandell GL, Bennett JE, Dolin R, eds. Principles and practice of infectious diseases. 5th ed. Philadelphia: Churchill Livingstone; 2000. p. 2310-35.
5. Bergogne-Berezin E. *Pseudomonas* and miscellaneous gram-negative bacilli. In: Cohen J, Powderly WG, eds. Infectious diseases. 2nd ed. Edinburgh: Mosby; 2004. p. 2203-26.
6. Kiska DL, Gilligan PH. *Pseudomonas*. In: Murray PR, ed. Manual of clinical microbiology. 8th ed. Washington DC: ASM Press; 2003. p. 719-28.
7. Arnow PM, Flaherty JP. Nonfermentative Gram-negative bacilli. In: Mayhall CG, ed. Hospital epidemiology and infection control. 2nd ed. Philadelphia: Lippincott Williams & Wilkins; 1999. p. 431-51.
8. Giesecke J. Modern Infectious disease epidemiology. 2nd ed. London: Arnold; 2002.
9. Last JM. A dictionary of Epidemiology. 4th ed. New York: Oxford University Press; 2001.
10. Hovig B, Lystad A. Infeksjonssykdommer - forebygging og kontroll (Infectious diseases - prevention and control). 2nd ed. Oslo: Universitetsforlaget; 1989.
11. Definisjon og klassifikasjon av sykehusinfeksjoner IK-2556 (Definition and classification of hospital infections). Oslo: Statens helsetilsyn; 1996.
12. Gregg M. Field epidemiology. 3rd ed. New York: Oxford University Press; 2008.
13. Hovig B, Lystad A, Opsjon H. A prevalence survey of infections among hospitalized patients in Norway. NIPH Ann 1981;4:49-60.
14. Aavitsland P, Stormark M, Lystad A. Hospital-acquired infections in Norway: a national prevalence survey in 1991. Scand J Infect Dis 1992;24:477-83.

15. Stormark M, Aavitsland P, Lystad A. [Prevalence of hospital infections in Norwegian somatic hospitals]. *Tidsskr Nor Laegeforen* 1993;113:173-7.
16. Scheel O, Stormark M. National prevalence survey on hospital infections in Norway. *J Hosp Infect* 1999;41:331-5.
17. Eriksen HM, Iversen BG, Aavitsland P. [Hospital infections in Norway 1999 and 2000]. *Tidsskr Nor Laegeforen* 2002;122:2440-3.
18. Eriksen HM, Iversen BG, Aavitsland P. Prevalence of nosocomial infections and use of antibiotics in long-term care facilities in Norway, 2002 and 2003. *J Hosp Infect* 2004;57:316-20.
19. Eriksen HM, Elstrom P, Harthug S, Akselsen PE. [Infection control in long-term care facilities for the elderly]. *Tidsskr Nor Laegeforen* 2005;125:1835-7.
20. Eriksen HM, Iversen BG, Aavitsland P. Prevalence of nosocomial infections in hospitals in Norway, 2002 and 2003. *J Hosp Infect* 2005;60:40-5.
21. Eriksen HM, Koch AM, Elstrom P, Nilsen RM, Harthug S, Aavitsland P. Healthcare-associated infection among residents of long-term care facilities: a cohort and nested case-control study. *J Hosp Infect* 2007;65:334-40.
22. Kapperud G, Nygard K. Oppklaring av utbrudd av næringsmiddelbårne sykdommer og zoonoser (Outbreak investigation of foodborne diseases and zoonoses). Oslo: Folkehelseinstituttet; 2006.
23. Lov om vern mot smittsomme sykdommer LOV-1994-08-05-55 (Communicable disease control act) [Lovdata]. [updated 1994 Aug 5; cited 2008 Aug. 15]. Available from: <http://www.lovdata.no/all/nl-19940805-055.html>.
24. Lov om medisinsk utstyr. LOV-1995-01-12-6 (Act on Medical devices) [Lovdata]. [updated 1995 Jan 12; cited 2008 July 15]. Available from: <http://www.lovdata.no/all/nl-19950112-006.html>.
25. Forskrift om medisinsk utstyr FOR-2005-12-15-1690 (Regulation on medical devices) [Lovdata]. [updated 2005 Dec 15; cited 2008 June 18]. Available from: <http://www.lovdata.no/for/sf/ho/ho-20051215-1690.html>.
26. European Council. European Council Directive 93/42/EEC of 14 June 1993 concerning medical devices. Council Directive 93/42/EEC ed. 1993.
27. Ph.Eur 5 (2005: 5.1.4). In: European Pharmacopoeia. 5th ed. Strasbourg: Council of Europe; 2005.
28. Lov om kosmetikk og kroppspfleieprodukt m.m. LOV 2005-12-21 nr 126 (Act on cosmetics and body care products etc.) [Lovdata]. [updated 2005 Dec 21; cited . Available from: <http://www.lovdata.no/all/hl-20051221-126.html>.
29. Generell forskrift for produksjon, import og frambud mv av kosmetikk og kroppspfleieprodukter FOR 1995-10-26 nr 871 (General regulation on production,

- import and sales of cosmetics and body care products) [Lovdata]. [updated 1995 Oct 26; cited 2008 July 15]
30. European Council. Council Directive of 27 July 1976 on the approximation of the laws of the Member States relating to cosmetic products [European Commission]. [updated 2007 Aug 29; cited 2008 Aug. 13]. Available from: <http://eur-lex.europa.eu/LexUriServ/LexUriServ.do?uri=CONSLEG:1976L0768:20070919:EN:PDF>.
 31. The sccp's notes of guidance for the testing of cosmetic ingredients and their safety evaluation. 6th revision [European Commission]. [updated 2006 Dec 19; cited 2008 Aug. 13]. Available from: http://ec.europa.eu/health/ph_risk/committees/04_sccp/docs/sccp_s_04.pdf.
 32. Press release: Commission launches much-awaited revision to the Medical Device Directives [European Commission]. [updated 2005 Dec 22; cited 2008 July 10]. Available from: <http://europa.eu/rapid/pressReleasesAction.do?reference=IP/05/1684&type=HTML&aged=0&language=EN&guiLanguage=en>.
 33. Forskrift om smittevern i helsetjenesten FOR 2005-06-17 nr 610 (Regulation on infection control and prevention in the health care service) [Lovdata]. [updated 2005 Jun 17; cited 2008 July 10]. Available from: <http://www.lovdata.no/for/sf/ho/xo-20050617-0610.html>.
 34. Gjennomgang av den sentrale helseforvaltningens roller og ansvar på områdene medisinsk utstyr, meldeordninger og smittevern i lys av Dent-O-Septsaken (Internal review on the roles and responsibilities of the central health administration in the fields of medical devices, discrepancy report systems and infection control). Oslo: Ministry of Health; 2002.
 35. Becks VE, Lorenzoni NM. *Pseudomonas aeruginosa* outbreak in a neonatal intensive care unit: a possible link to contaminated hand lotion. *Am J Infect Control* 1995;23:396-8.
 36. Bou R, Aguilar A, Perpinan J, Ramos P, Peris M, Lorente L, et al. Nosocomial outbreak of *Pseudomonas aeruginosa* infections related to a flexible bronchoscope. *J Hosp Infect* 2006;64:129-35.
 37. Prospero E, Barbadoro P, Savini S, Manso E, Annino I, D'Errico MM. Cluster of *Pseudomonas aeruginosa* catheter-related bloodstream infections traced to contaminated multidose heparinized saline solutions in a medical ward. *Int J Hyg Environ Health* 2006;209:553-6.
 38. Bukholm G, Tannaes T, Kjelsberg AB, Smith-Erichsen N. An outbreak of multidrug-resistant *Pseudomonas aeruginosa* associated with increased risk of patient death in an intensive care unit. *Infect Control Hosp Epidemiol* 2002;23:441-6.
 39. BATTERY JP, ALABASTER SJ, HEINE RG, SCOTT SM, CRUTCHFIELD RA, BIGHAM A, et al. Multiresistant *Pseudomonas aeruginosa* outbreak in a pediatric oncology ward related to bath toys. *Pediatr Infect Dis J* 1998;17:509-13.

40. Corvec S, Poirel L, Espaze E, Giraudeau C, Drugeon H, Nordmann P. Long-term evolution of a nosocomial outbreak of *Pseudomonas aeruginosa* producing VIM-2 metallo-enzyme. *J Hosp Infect* 2008;68:73-82.
41. Cobben NA, Drent M, Jonkers M, Wouters EF, Vaneechoutte M, Stobberingh EE. Outbreak of severe *Pseudomonas aeruginosa* respiratory infections due to contaminated nebulizers. *J Hosp Infect* 1996;33:63-70.
42. Eckmanns T, Oppert M, Martin M, Amorosa R, Zuschneid I, Frei U, et al. An outbreak of hospital-acquired *Pseudomonas aeruginosa* infection caused by contaminated bottled water in intensive care units. *Clin Microbiol Infect* 2008;14:454-8.
43. Millership SE, Patel N, Chattopadhyay B. The colonization of patients in an intensive treatment unit with gram-negative flora: the significance of the oral route. *J Hosp Infect* 1986;7:226-35.
44. Farmer JJ, III, Weinstein RA, Zierdt CH, Brokopp CD. Hospital outbreaks caused by *Pseudomonas aeruginosa*: importance of serogroup O11. *J Clin Microbiol* 1982;16:266-70.
45. Foca M, Jakob K, Whittier S, Della LP, Factor S, Rubenstein D, et al. Endemic *Pseudomonas aeruginosa* infection in a neonatal intensive care unit. *N Engl J Med* 2000;343:695-700.
46. Fraser TG, Reiner S, Malczynski M, Yarnold PR, Warren J, Noskin GA. Multidrug-resistant *Pseudomonas aeruginosa* cholangitis after endoscopic retrograde cholangiopancreatography: failure of routine endoscope cultures to prevent an outbreak. *Infect Control Hosp Epidemiol* 2004;25:856-9.
47. Gales AC, Torres PL, Vilarinho DS, Melo RS, Silva CF, Cereda RF. Carbapenem-resistant *Pseudomonas aeruginosa* outbreak in an intensive care unit of a teaching hospital. *Braz J Infect Dis* 2004;8:267-71.
48. Gibb AP, Tribuddharat C, Moore RA, Louie TJ, Krulicki W, Livermore DM, et al. Nosocomial outbreak of carbapenem-resistant *Pseudomonas aeruginosa* with a new bla(IMP) allele, bla(IMP-7). *Antimicrob Agents Chemother* 2002;46:255-8.
49. Gillespie JL, Arnold KE, Noble-Wang J, Jensen B, Arduino M, Hageman J, et al. Outbreak of *Pseudomonas aeruginosa* infections after transrectal ultrasound-guided prostate biopsy. *Urology* 2007;69:912-4.
50. Gras-Le Guen C, Lepelletier D, Debillon T, Gournay V, Espaze E, Roze JC. Contamination of a milk bank pasteuriser causing a *Pseudomonas aeruginosa* outbreak in a neonatal intensive care unit. *Arch Dis Child Fetal Neonatal Ed* 2003;88:F434-F435.
51. Grigis A, Goglio A, Parea M, Gneccchi F, Minetti B, Barbui T. Nosocomial outbreak of severe *Pseudomonas aeruginosa* infections in haematological patients. *Eur J Epidemiol* 1993;9:390-5.

52. Hocquet D, Bertrand X, Kohler T, Talon D, Plesiat P. Genetic and phenotypic variations of a resistant *Pseudomonas aeruginosa* epidemic clone. *Antimicrob Agents Chemother* 2003;47:1887-94.
53. Juma P, Chattopadhyay B. Outbreak of gentamicin, ciprofloxacin-resistant *Pseudomonas aeruginosa* in an intensive care unit, traced to contaminated quivers. *J Hosp Infect* 1994;28:209-18.
54. Kayabas U, Bayraktar M, Otlu B, Ugras M, Ersoy Y, Bayindir Y, et al. An outbreak of *Pseudomonas aeruginosa* because of inadequate disinfection procedures in a urology unit: a pulsed-field gel electrophoresis-based epidemiologic study. *Am J Infect Control* 2008;36:33-8.
55. Keene WE, Markum AC, Samadpour M. Outbreak of *Pseudomonas aeruginosa* infections caused by commercial piercing of upper ear cartilage. *JAMA* 2004;291:981-5.
56. Kerr JR, Moore JE, Curran MD, Graham R, Webb CH, Lowry KG, et al. Investigation of a nosocomial outbreak of *Pseudomonas aeruginosa* pneumonia in an intensive care unit by random amplification of polymorphic DNA assay. *J Hosp Infect* 1995;30:125-31.
57. Kolmos HJ, Thuesen B, Nielsen SV, Lohmann M, Kristoffersen K, Rosdahl VT. Outbreak of infection in a burns unit due to *Pseudomonas aeruginosa* originating from contaminated tubing used for irrigation of patients. *J Hosp Infect* 1993;24:11-21.
58. Kumar D, Catral MS, Robicsek A, Gaudreau C, Humar A. Outbreak of *Pseudomonas aeruginosa* by multiple organ transplantation from a common donor. *Transplantation* 2003;75:1053-5.
59. Lyytikäinen O, Golovanova V, Kolho E, Ruutu P, Sivonen A, Tiittanen L, et al. Outbreak caused by tobramycin-resistant *Pseudomonas aeruginosa* in a bone marrow transplantation unit. *Scand J Infect Dis* 2001;33:445-9.
60. Moolenaar RL, Crutcher JM, San Joaquin VH, Sewell LV, Hutwagner LC, Carson LA, et al. A prolonged outbreak of *Pseudomonas aeruginosa* in a neonatal intensive care unit: did staff fingernails play a role in disease transmission? *Infect Control Hosp Epidemiol* 2000;21:80-5.
61. Panzig B, Schroder G, Pitten FA, Grundling M. A large outbreak of multiresistant *Pseudomonas aeruginosa* strains in north-eastern Germany. *J Antimicrob Chemother* 1999;43:415-8.
62. Pena C, Dominguez MA, Pujol M, Verdaguer R, Gudiol F, Ariza J. An outbreak of carbapenem-resistant *Pseudomonas aeruginosa* in a urology ward. *Clin Microbiol Infect* 2003;9:938-43.
63. Richet H, Escande MC, Marie JP, Zittoun R, Lagrange PH. Epidemic *Pseudomonas aeruginosa* serotype O16 bacteremia in hematology-oncology patients. *J Clin Microbiol* 1989;27:1992-6.

64. Schelenz S, French G. An outbreak of multidrug-resistant *Pseudomonas aeruginosa* infection associated with contamination of bronchoscopes and an endoscope washer-disinfector. *J Hosp Infect* 2000;46:23-30.
65. Silva CV, Magalhaes VD, Pereira CR, Kawagoe JY, Ikura C, Ganc AJ. Pseudo-outbreak of *Pseudomonas aeruginosa* and *Serratia marcescens* related to bronchoscopes. *Infect Control Hosp Epidemiol* 2003;24:195-7.
66. Srinivasan A, Wolfenden LL, Song X, Mackie K, Hartsell TL, Jones HD, et al. An outbreak of *Pseudomonas aeruginosa* infections associated with flexible bronchoscopes. *N Engl J Med* 2003;348:221-7.
67. Stephenson JR, Heard SR, Richards MA, Tabaqchali S. Gastrointestinal colonization and septicaemia with *Pseudomonas aeruginosa* due to contaminated thymol mouthwash in immunocompromised patients. *J Hosp Infect* 1985;6:369-78.
68. Talon D, Capellier G, Boillot A, Michel-Briand Y. Use of pulsed-field gel electrophoresis as an epidemiologic tool during an outbreak of *Pseudomonas aeruginosa* lung infections in an intensive care unit. *Intensive Care Med* 1995;21:996-1002.
69. Tassios PT, Gennimata V, Spaliara-Kalogeropoulou L, Kairis D, Koutsia C, Vatopoulos AC, et al. Multiresistant *Pseudomonas aeruginosa* serogroup O:11 outbreak in an intensive care unit. *Clin Microbiol Infect* 1997;3:621-8.
70. Tredget EE, Shankowsky HA, Joffe AM, Inkson TI, Volpel K, Paranchych W, et al. Epidemiology of infections with *Pseudomonas aeruginosa* in burn patients: the role of hydrotherapy. *Clin Infect Dis* 1992;15:941-9.
71. Widmer AF, Wenzel RP, Trilla A, Bale MJ, Jones RN, Doebbeling BN. Outbreak of *Pseudomonas aeruginosa* infections in a surgical intensive care unit: probable transmission via hands of a health care worker. *Clin Infect Dis* 1993;16:372-6.
72. Yardy GW, Cox RA. An outbreak of *Pseudomonas aeruginosa* infection associated with contaminated urodynamic equipment. *J Hosp Infect* 2001;47:60-3.
73. Zawacki A, O'Rourke E, Potter-Bynoe G, Macone A, Harbarth S, Goldmann D. An outbreak of *Pseudomonas aeruginosa* pneumonia and bloodstream infection associated with intermittent otitis externa in a healthcare worker. *Infect Control Hosp Epidemiol* 2004;25:1083-9.
74. Berthelot P, Grattard F, Amerger C, Frery MC, Lucht F, Pozzetto B, et al. Investigation of a nosocomial outbreak due to *Serratia marcescens* in a maternity hospital. *Infect Control Hosp Epidemiol* 1999;20:233-6.
75. Heltberg O, Skov F, Gerner-Smidt P, Kolmos HJ, Dybkjaer E, Gutschik E, et al. Nosocomial epidemic of *Serratia marcescens* septicemia ascribed to contaminated blood transfusion bags. *Transfusion* 1993;33:221-7.
76. Szewzyk U, Szewzyk R, Stenstrom TA. Growth and survival of *Serratia marcescens* under aerobic and anaerobic conditions in the presence of materials from blood bags. *J Clin Microbiol* 1993;31:1826-30.

77. de Boer MG, Brunsveld-Reinders AH, Salomons EM, Dijkshoorn L, Bernards AT, van den Berg PC, et al. Multifactorial origin of high incidence of *Serratia marcescens* in a cardio-thoracic ICU: analysis of risk factors and epidemiological characteristics. *J Infect* 2008;56:446-53.
78. Maragakis LL, Winkler A, Tucker MG, Cosgrove SE, Ross T, Lawson E, et al. Outbreak of multidrug-resistant *Serratia marcescens* infection in a neonatal intensive care unit. *Infect Control Hosp Epidemiol* 2008;29:418-23.
79. Cox TR, Roland WE, Dolan ME. Ventilator-related *Acinetobacter* outbreak in an intensive care unit. *Mil Med* 1998;163:389-91.
80. Dealler S. Nosocomial outbreak of multi-resistant *Acinetobacter* sp. on an intensive care unit: possible association with ventilation equipment. *J Hosp Infect* 1998;38:147-8.
81. Koeleman JG, Parlevliet GA, Dijkshoorn L, Savelkoul PH, Vandenbroucke-Grauls CM. Nosocomial outbreak of multi-resistant *Acinetobacter baumannii* on a surgical ward: epidemiology and risk factors for acquisition. *J Hosp Infect* 1997;37:113-23.
82. Podnos YD, Cinat ME, Wilson SE, Cooke J, Gornick W, Thrupp LD. Eradication of multi-drug resistant *Acinetobacter* from an intensive care unit. *Surg Infect (Larchmt)* 2001;2:297-301.
83. Struelens MJ, Carlier E, Maes N, Serruys E, Quint WG, van Belkum A. Nosocomial colonization and infection with multiresistant *Acinetobacter baumannii*: outbreak delineation using DNA macrorestriction analysis and PCR-fingerprinting. *J Hosp Infect* 1993;25:15-32.
84. Aumeran C, Paillard C, Robin F, Kanold J, Baud O, Bonnet R, et al. *Pseudomonas aeruginosa* and *Pseudomonas putida* outbreak associated with contaminated water outlets in an oncohaematology paediatric unit. *J Hosp Infect* 2007;65:47-53.
85. Update: Delayed onset *Pseudomonas fluorescens* bloodstream infections after exposure to contaminated heparin flush--Michigan and South Dakota, 2005-2006. *MMWR Morb Mortal Wkly Rep* 2006;55:961-3.
86. Weems JJ, Jr. Nosocomial outbreak of *Pseudomonas cepacia* associated with contamination of reusable electronic ventilator temperature probes. *Infect Control Hosp Epidemiol* 1993;14:583-6.
87. Souza Dias MB, Habert AB, Borrasca V, Stempliuk V, Ciolli A, Araujo MR, et al. Salvage of long-term central venous catheters during an outbreak of *Pseudomonas putida* and *Stenotrophomonas maltophilia* infections associated with contaminated heparin catheter-lock solution. *Infect Control Hosp Epidemiol* 2008;29:125-30.
88. Alfieri N, Ramotar K, Armstrong P, Spornitz ME, Ross G, Winnick J, et al. Two consecutive outbreaks of *Stenotrophomonas maltophilia* (*Xanthomonas maltophilia*) in an intensive-care unit defined by restriction fragment-length polymorphism typing. *Infect Control Hosp Epidemiol* 1999;20:553-6.

89. Estivariz CF, Bhatti LI, Pati R, Jensen B, Arduino MJ, Jernigan D, et al. An outbreak of *Burkholderia cepacia* associated with contamination of albuterol and nasal spray. *Chest* 2006;130:1346-53.
90. Ghazal SS, Al Mudaimiegh K, Al Fakihi EM, Asery AT. Outbreak of *Burkholderia cepacia* bacteremia in immunocompetent children caused by contaminated nebulized sulbutamol in Saudi Arabia. *Am J Infect Control* 2006;34:394-8.
91. Lee JK. Two outbreaks of *Burkholderia cepacia* nosocomial infection in a neonatal intensive care unit. *J Paediatr Child Health* 2008;44:62-6.
92. Nasser RM, Rahi AC, Haddad MF, Daoud Z, Irani-Hakime N, Almawi WY. Outbreak of *Burkholderia cepacia* bacteremia traced to contaminated hospital water used for dilution of an alcohol skin antiseptic. *Infect Control Hosp Epidemiol* 2004;25:231-9.
93. Gonzalez-Vertiz A, Alcantar-Curiel D, Cuauhtli M, Daza C, Gayosso C, Solache G, et al. Multiresistant extended-spectrum beta-lactamase-producing *Klebsiella pneumoniae* causing an outbreak of nosocomial bloodstream infection. *Infect Control Hosp Epidemiol* 2001;22:723-5.
94. Harthug S, Digranes A, Hope O, Kristiansen BE, Allum AG, Langeland N. Vancomycin resistance emerging in a clonal outbreak caused by ampicillin-resistant *Enterococcus faecium*. *Clin Microbiol Infect* 2000;6:19-28.
95. Flaherty JP, Garcia-Houchins S, Chudy R, Arnow PM. An outbreak of gram-negative bacteremia traced to contaminated O-rings in reprocessed dialyzers. *Ann Intern Med* 1993;119:1072-8.
96. Gray J, George RH, Durbin GM, Ewer AK, Hocking MD, Morgan ME. An outbreak of *Bacillus cereus* respiratory tract infections on a neonatal unit due to contaminated ventilator circuits. *J Hosp Infect* 1999;41:19-22.
97. Jhung MA, Sunenshine RH, Noble-Wang J, Coffin SE, St John K, Lewis FM, et al. A national outbreak of *Ralstonia mannitolilytica* associated with use of a contaminated oxygen-delivery device among pediatric patients. *Pediatrics* 2007;119:1061-8.
98. Lemaitre D, Elaichouni A, Hundhausen M, Claeys G, Vanhaesebrouck P, Vaneechoutte M, et al. Tracheal colonization with *Sphingomonas paucimobilis* in mechanically ventilated neonates due to contaminated ventilator temperature probes. *J Hosp Infect* 1996;32:199-206.
99. Bert F, Maubec E, Bruneau B, Berry P, Lambert-Zechovsky N. Multi-resistant *Pseudomonas aeruginosa* outbreak associated with contaminated tap water in a neurosurgery intensive care unit. *J Hosp Infect* 1998;39:53-62.
100. Cheng K, Smyth RL, Govan JR, Doherty C, Winstanley C, Denning N, et al. Spread of beta-lactam-resistant *Pseudomonas aeruginosa* in a cystic fibrosis clinic. *Lancet* 1996;348:639-42.
101. Koeleman JG, Stoof J, Biesmans DJ, Savelkoul PH, Vandenbroucke-Grauls CM. Comparison of amplified ribosomal DNA restriction analysis, random amplified polymorphic DNA analysis, and amplified fragment length polymorphism

- fingerprinting for identification of *Acinetobacter* genomic species and typing of *Acinetobacter baumannii*. *J Clin Microbiol* 1998;36:2522-9.
102. Bergmans DC, Bonten MJ, van Tiel FH, Gaillard CA, van der GS, Wiltink RM, et al. Cross-colonisation with *Pseudomonas aeruginosa* of patients in an intensive care unit. *Thorax* 1998;53:1053-8.
 103. Grobner S, Heeg P, Autenrieth IB, Schulte B. Monoclonal outbreak of catheter-related bacteraemia by *Ralstonia mannitolilytica* on two haemato-oncology wards. *J Infect* 2007;55:539-44.
 104. Stephenson JR, Heard SR, Richards MA, Tabaqchali S. Outbreak of septicaemia due to contaminated mouthwash. *Br Med J (Clin Res Ed)* 1984;289:1584.
 105. Alp E, Voss A. Ventilator associated pneumonia and infection control. *Ann Clin Microbiol Antimicrob* 2006;5:7.
 106. Bergmans DC, Bonten MJ, Gaillard CA, Paling JC, van der GS, van Tiel FH, et al. Prevention of ventilator-associated pneumonia by oral decontamination: a prospective, randomized, double-blind, placebo-controlled study. *Am J Respir Crit Care Med* 2001;164:382-8.
 107. Yoneyama T, Yoshida M, Ohnishi T, Mukaiyama H, Okamoto H, Hoshiba K, et al. Oral care reduces pneumonia in older patients in nursing homes. *J Am Geriatr Soc* 2002;50:430-3.
 108. Prince AS. Biofilms, antimicrobial resistance, and airway infection. *N Engl J Med* 2002;347:1110-1.
 109. Singh PK, Parsek MR, Greenberg EP, Welsh MJ. A component of innate immunity prevents bacterial biofilm development. *Nature* 2002;417:552-5.
 110. Singh PK, Schaefer AL, Parsek MR, Moninger TO, Welsh MJ, Greenberg EP. Quorum-sensing signals indicate that cystic fibrosis lungs are infected with bacterial biofilms. *Nature* 2000;407:762-4.
 111. Dunne WM, Jr. Bacterial adhesion: seen any good biofilms lately? *Clin Microbiol Rev* 2002;15:155-66.
 112. Shirtliff ME, Mader JT, Camper AK. Molecular interactions in biofilms. *Chem Biol* 2002;9:859-71.
 113. Kang CI, Kim SH, Kim HB, Park SW, Choe YJ, Oh MD, et al. *Pseudomonas aeruginosa* bacteremia: risk factors for mortality and influence of delayed receipt of effective antimicrobial therapy on clinical outcome. *Clin Infect Dis* 2003;37:745-51.
 114. Sherertz RJ, Sarubbi FA. A three-year study of nosocomial infections associated with *Pseudomonas aeruginosa*. *J Clin Microbiol* 1983;18:160-4.
 115. Häussler S. *Pseudomonas aeruginosa* Biofilms: Impact of small colony variants on chronic persistent infections. In: Cornelis P, ed. *Pseudomonas* Genomics and molecular biology. Norfolk: Caister Academic press; 2008. p. 159-75.

116. NORM/NORM-VET 2006. Usage of Antimicrobial Agents and Occurrence of Antimicrobial Resistance in Norway. Tromsø/Oslo: 2007.
117. National Nosocomial Infections Surveillance (NNIS) report, data summary from October 1986-April 1996, issued May 1996. A report from the National Nosocomial Infections Surveillance (NNIS) System. *Am J Infect Control* 1996;24:380-8.
118. Javaloyas M, Garcia-Somoza D, Gudiol F. Epidemiology and prognosis of bacteremia: a 10-y study in a community hospital. *Scand J Infect Dis* 2002;34:436-41.
119. Scheckler WE, Bobula JA, Beamsley MB, Hadden ST. Bloodstream infections in a community hospital: a 25-year follow-up. *Infect Control Hosp Epidemiol* 2003;24:936-41.
120. Weinstein MP, Reller LB, Murphy JR, Lichtenstein KA. The clinical significance of positive blood cultures: a comprehensive analysis of 500 episodes of bacteremia and fungemia in adults. I. Laboratory and epidemiologic observations. *Rev Infect Dis* 1983;5:35-53.
121. Diekema DJ, Pfaller MA, Jones RN, Doern GV, Winokur PL, Gales AC, et al. Survey of bloodstream infections due to gram-negative bacilli: frequency of occurrence and antimicrobial susceptibility of isolates collected in the United States, Canada, and Latin America for the SENTRY Antimicrobial Surveillance Program, 1997. *Clin Infect Dis* 1999;29:595-607.
122. Blot S, Vandewoude K, Hoste E, Colardyn F. Reappraisal of attributable mortality in critically ill patients with nosocomial bacteraemia involving *Pseudomonas aeruginosa*. *J Hosp Infect* 2003;53:18-24.
123. Bisbe J, Gatell JM, Puig J, Mallolas J, Martinez JA, Jimenez de Anta MT, et al. *Pseudomonas aeruginosa* bacteremia: univariate and multivariate analyses of factors influencing the prognosis in 133 episodes. *Rev Infect Dis* 1988;10:629-35.
124. Gallagher PG, Watanakunakorn C. *Pseudomonas* bacteremia in a community teaching hospital, 1980-1984. *Rev Infect Dis* 1989;11:846-52.
125. Mallolas J, Gatell JM, Miro JM, Marco F, Soriano E. Epidemiologic characteristics and factors influencing the outcome of *Pseudomonas aeruginosa* bacteremia. *Rev Infect Dis* 1990;12:718-9.
126. Micek ST, Lloyd AE, Ritchie DJ, Reichley RM, Fraser VJ, Kollef MH. *Pseudomonas aeruginosa* bloodstream infection: importance of appropriate initial antimicrobial treatment. *Antimicrob Agents Chemother* 2005;49:1306-11.
127. Aliaga L, Mediavilla JD, Cobo F. A clinical index predicting mortality with *Pseudomonas aeruginosa* bacteraemia. *J Med Microbiol* 2002;51:615-9.
128. Tacconelli E, Tumbarello M, Bertagnolio S, Citton R, Spanu T, Fadda G, et al. Multidrug-resistant *Pseudomonas aeruginosa* bloodstream infections: analysis of trends in prevalence and epidemiology. *Emerg Infect Dis* 2002;8:220-1.

129. Marra AR, Bearman GM, Wenzel RP, Edmond MB. Comparison of severity of illness scoring systems for patients with nosocomial bloodstream infection due to *Pseudomonas aeruginosa*. *BMC Infect Dis* 2006;6:132.
130. Vidal F, Mensa J, Almela M, Martinez JA, Marco F, Casals C, et al. Epidemiology and outcome of *Pseudomonas aeruginosa* bacteremia, with special emphasis on the influence of antibiotic treatment. Analysis of 189 episodes. *Arch Intern Med* 1996;156:2121-6.
131. Weinstein MP, Murphy JR, Reller LB, Lichtenstein KA. The clinical significance of positive blood cultures: a comprehensive analysis of 500 episodes of bacteremia and fungemia in adults. II. Clinical observations, with special reference to factors influencing prognosis. *Rev Infect Dis* 1983;5:54-70.
132. Kuikka A, Valtonen VV. Factors associated with improved outcome of *Pseudomonas aeruginosa* bacteremia in a Finnish university hospital. *Eur J Clin Microbiol Infect Dis* 1998;17:701-8.
133. Hammerstrom J, Roym AL, Gran FW. [Bacteremia in hematological malignant disorders.]. *Tidsskr Nor Laegeforen* 2008;128:1655-9.
134. Hall-Stoodley L, Stoodley P. Biofilm formation and dispersal and the transmission of human pathogens. *Trends Microbiol* 2005;13:7-10.
135. Donlan RM, Costerton JW. Biofilms: survival mechanisms of clinically relevant microorganisms. *Clin Microbiol Rev* 2002;15:167-93.
136. Anderson RL, Holland BW, Carr JK, Bond WW, Favero MS. Effect of disinfectants on pseudomonads colonized on the interior surface of PVC pipes. *Am J Public Health* 1990;80:17-21.
137. Tenover FC, Arbeit RD, Goering RV, Mickelsen PA, Murray BE, Persing DH, et al. Interpreting chromosomal DNA restriction patterns produced by pulsed-field gel electrophoresis: criteria for bacterial strain typing. *J Clin Microbiol* 1995;33:2233-9.
138. Amplified fragment length polymorphism [Wikipedia]. [updated 2008 Jul 12; cited . Available from:
http://en.wikipedia.org/wiki/Amplified_fragment_length_polymorphism.
139. Speijer H, Savelkoul PH, Bonten MJ, Stobberingh EE, Tjhi JH. Application of different genotyping methods for *Pseudomonas aeruginosa* in a setting of endemicity in an intensive care unit. *J Clin Microbiol* 1999;37:3654-61.
140. Hill AB. The environment and disease: Association or causation? *Proceedings of the Royal Society of Medicine - London* 1965;58:295-300.
141. Johnson S. *The Ghost Map*. London: Allen Lane, Penguin Books Ltd; 2006.
142. Snow J. *On the mode of communication of cholera*. 2nd ed. London: John Churchill; 1855.

143. Koch R. Die aetiologie der Tuberkulose. In: Schwalbe J, ed. Gesammelte Werke von Koch. Leipzig: Georg Thieme Verlag; 1912. p. 428-55.
144. Evans AS. Causation and disease: the Henle-Koch postulates revisited. *Yale Journal of Biology and Medicine* 1976;49:175-95.
145. MacMahon B, Pugh TF. *Epidemiology Principles and methods*. Boston: Little, Brown and Company; 1970.
146. Dowe P. Counterfactual Theories of Causation [Stanford Encyclopedia of Philosophy]. [updated 2001 Jan 12; cited 2008 Feb. 29]. Available from: <http://plato.stanford.edu/entries/causation-counterfactual/>.
147. Hofler M. Causal inference based on counterfactuals. *BMC Med Res Methodol* 2005;5:28.
148. Hoefer C. Causal Determinism [Stanford Encyclopedia of Philosophy]. [updated 2008 Jan 23; cited 2008 Feb. 28]. Available from: <http://plato.stanford.edu/entries/determinism-causal/>.
149. Hitchcock C. Probabilistic Causation [Stanford Encyclopedia of Philosophy]. [updated 2002; cited 2007 Nov. 6]. Available from: <http://plato.stanford.edu/entries/causation-probabilistic/>.
150. Rothman KJ, Greenland S. Causation and causal inference. In: Rothman KJ, Greenland S, eds. *Modern Epidemiology*. 2nd ed. Philadelphia: Lippincott Williams & Wilkins; 1998. p. 7-28.
151. Phillips CV, Goodman KJ. The missed lessons of Sir Austin Bradford Hill. *Epidemiol Perspect Innov* 2004;1:3.
152. Lipton R, Odegaard T. Causal thinking and causal language in epidemiology: it's in the details. *Epidemiol Perspect Innov* 2005;2:8.
153. Hofler M. The Bradford Hill considerations on causality: a counterfactual perspective. *Emerg Themes Epidemiol* 2005;2:11.
154. Phillips CV, Goodman KJ. Causal criteria and counterfactuals; nothing more (or less) than scientific common sense. *Emerg Themes Epidemiol* 2006;3:5.
155. Ph.Eur 4 (2002: 2.6.12). In: *European Pharmacopoeia*. 4th ed. Strasbourg: Council of Europe; 2002.
156. Ph.Eur 4 (2002: 2.6.13). In: *European Pharmacopoeia*. 4th ed. Strasbourg: Council of Europe; 2002.
157. Ph.Eur 4 (2002: 5.1.3). In: *European Pharmacopoeia*. 4th ed. Strasbourg: Council of Europe; 2002.
158. International Statistical Classification of Diseases and Related Health Problems. 10th Revision [WHO]. [updated 2007; cited 2007 Nov. 29]. Available from: <http://www.who.int/classifications/apps/icd/icd10online/>.

159. Nygard K. Water and infection. Epidemiological studies of epidemic and endemic waterborne disease (Dissertation). Oslo: University of Oslo; 2008.
160. Rapport til Helsedepartementet om Helsetilsynets oppfølging i Dent-O-Sept saken (Report to the Ministry of Health on Follow-up of the Dent-O-Sept Incident by the Norwegian Board of Health). Oslo: Statens helsetilsyn; 2003.
161. Ministry of Health. Handlingsplan for å forebygge sykehusinfeksjoner 2004-2006 (Action plan to prevent hospital acquired infections 2004-2006). Oslo: Ministry of Health; 2004.
162. Sluttrapport fra tilsyn med Snøgg Industri AS (Final report from the audit of Snøgg Industri AS). Oslo: Sosial- og helsedirektoratet; 2003.
163. Dent-O-Septsaken - Kartleggingsundersøkelser i forbindelse med utbruddet av infeksjoner forårsaket av *Pseudomonas aeruginosa* fra Dent-O-Sept munnpensler (The Dent-O-Sept case). Oslo: Folkehelseinstituttet; 2003.
164. Dent-O-Sept saken. Vurderinger fra Sosial- og helsedirektoratet som nasjonal smittevernmyndighet (The Dent-O-Sept case. Considerations by the Directorate of Health and Social Services as national infection control authority). Oslo: Sosial- og helsedirektoratet; 2004.
165. Sosial- og helsedirektoratets oppfølging av Dent-O-Sept saken som forvaltningsansvarlig for medisinsk utstyr (The follow-up of The Dent-O-Sept case by the Directorate of Health and Social Services as administrative authority of medical devices). Oslo: Sosial- og helsedirektoratet; 2004.
166. Forskrift om internkontroll i sosial- og helsetjenesten. FOR 2002-12-20 nr 1731 (Regulation on internal quality control systems in social services and health care) [Lovdata]. [updated 2002 Dec 20; cited 2008 Feb. 26]. Available from: <http://www.lovdata.no/cgi-wift/ldles?doc=/sf/sf/sf-20021220-1731.html>.
167. Schimmer B, Nygard K, Eriksen HM, Lassen J, Lindstedt BA, Brandal LT, et al. Outbreak of haemolytic uraemic syndrome in Norway caused by stx2-positive *Escherichia coli* O103:H25 traced to cured mutton sausages. *BMC Infect Dis* 2008;8:41.
168. Nygard K, Werner-Johansen O, Ronsen S, Caugant DA, Simonsen O, Kanestrom A, et al. An outbreak of legionnaires disease caused by long-distance spread from an industrial air scrubber in Sarpsborg, Norway. *Clin Infect Dis* 2008;46:61-9.
169. Cruciani M, Malena M, Amalfitano G, Monti P, Bonomi L. Molecular epidemiology in a cluster of cases of postoperative *Pseudomonas aeruginosa* endophthalmitis. *Clin Infect Dis* 1998;26:330-3.
170. Bo G. Analyserapport. *Pseudomonas aeruginosa* i Dent-O-Sept (Analysis report. *Pseudomonas aeruginosa* in Dent-O-Sept). Kristiansand: Naeringsmiddeltilsynet i Vest-Agder, Laboratorium; 2002.

171. Lov om legemidler m.v. LOV-1992-12-04-132 (Act on medicinal products) [Lovdata]. [updated 1992 Dec 4; cited 2008 July 15]. Available from: <http://www.lovdata.no/all/nl-19921204-132.html>.
172. Forskrift om legemidler FOR-1999-12-22-1559 (Regulation on medicinal products) [Lovdata]. [updated 1999 Dec 22; cited 2008 July 15]. Available from: <http://www.lovdata.no/for/sf/ho/ho-19991222-1559.html>.
173. Lassen J, Lingaas E. Vurdering om produksjonsprosessen for DENT-O-SEPT munnpensel er forsvarlig (Assessment of whether the production process of the Dent-O-Sept mouthswab is safe). Oslo: Sosial- og helsedirektoratet; 2002.
174. Iversen BG. Dent-O-Sept-utbruddet (The Dent-O-Sept outbreak) [Dagens Medisin]. [updated 2007 Dec 13; cited 2008 July 15]. Available from: <http://www.dagensmedisin.no/debatt/2007/12/13/dent-o-sept-utbruddet/index.xml>.
175. Baastad KL. Farmasøytblikk på munnpenseltragedien (A pharmacist's view on the mouth swab tragedy). Norges apotekerforenings tidsskrift 2007;115:153-5.
176. Henriksen K. Nye påstander om Dent-O-Sept (New assertions about Dent-O-Sept) [Dagens Medisin]. [updated 2007 Sep 13; cited 2008 July 8]. Available from: <http://www.dagensmedisin.no/nyheter/2007/09/13/nye-pastander-om-dent-o-se/>.
177. Baastad KL. Munnpensel-skandalen (The mouth swab scandal) [Dagens Medisin]. [updated 2007 Nov 13; cited 2008 July 8]. Available from: <http://www.dagensmedisin.no/debatt/2007/11/13/munnpensel-skandalen/index.xml>.
178. Baastad KL. Munnpenselsaken (The mouth swab case). Dagens Medisin. 2008 Jan 17.
179. Baastad KL. Contradiction to the author's conclusion (Comment). Ann Clin Microbiol Antimicrob 2007;
180. Alvarado CJ, Stolz SM, Maki DG. Nosocomial infections from contaminated endoscopes: a flawed automated endoscope washer. An investigation using molecular epidemiology. Am J Med 1991;91:272S-80S.
181. Ayliffe GA, Babb JR, Collins BJ, Lowbury EJ, Newsom SW. Pseudomonas aeruginosa in hospital sinks. Lancet 1974;2:578-81.
182. Casewell MW, Slater NG, Cooper JE. Operating theatre water-baths as a cause of pseudomonas septicemia. J Hosp Infect 1981;2:237-47.
183. Cryan EM, Falkiner FR, Mulvihill TE, Keane CT, Keeling PW. Pseudomonas aeruginosa cross-infection following endoscopic retrograde cholangiopancreatography. J Hosp Infect 1984;5:371-6.
184. Hofler M. Getting causal considerations back on the right track. Emerg Themes Epidemiol 2006;3:8.
185. Rothman KJ, Greenland S. Causation and causal inference in epidemiology. Am J Public Health 2005;95 Suppl 1:S144-S150.

186. Vineis P. Causality in epidemiology. *Soz Praventivmed* 2003;48:80-7.
187. Green L. The Causal Relation Issue in Negligence Law. *Michigan Law Review* 1962;60:543-76.
188. Hart HLA, Honoré AM. *Causation in the Law*. 2nd ed. Oxford: Clarendon; 1985.
189. Honoré AM. Causation in the law [Stanford Encyclopedia of Philosophy]. [updated 2005 Oct 12; cited 2008 Feb. 27]. Available from: <http://plato.stanford.edu/entries/causation-law/>.
190. Ozonoff D. Legal causation and responsibility for causing harm. *Am J Public Health* 2005;95 Suppl 1:S35-S38.
191. Stapleton J. Legal Cause: Cause-in-Fact and the Scope of Liability for Consequences. *Vanderbilt Law Review* 2001;54:941-1000.
192. Rothman KJ. *Epidemiology*. New York: Oxford University Press; 2002.
193. Al Hasan MN, Wilson JW, Lahr BD, Eckel-Passow JE, Baddour LM. Incidence of *Pseudomonas aeruginosa* bacteremia: a population-based study. *Am J Med* 2008;121:702-8.
194. Ferech M, Coenen S, Dvorakova K, Hendrickx E, Suetens C, Goossens H. European Surveillance of Antimicrobial Consumption (ESAC): outpatient penicillin use in Europe. *J Antimicrob Chemother* 2006;58:408-12.
195. Ferech M, Coenen S, Malhotra-Kumar S, Dvorakova K, Hendrickx E, Suetens C, et al. European Surveillance of Antimicrobial Consumption (ESAC): outpatient antibiotic use in Europe. *J Antimicrob Chemother* 2006;58:401-7.
196. Vander Stichele RH, Elseviers MM, Ferech M, Blot S, Goossens H. Hospital consumption of antibiotics in 15 European countries: results of the ESAC Retrospective Data Collection (1997-2002). *J Antimicrob Chemother* 2006;58:159-67.
197. Allen KD, Bartzokas CA, Graham R, Gibson MF, Gilbertson AA. Acquisition of endemic *Pseudomonas aeruginosa* on an intensive therapy unit. *J Hosp Infect* 1987;10:156-64.
198. NORM/NORM-VET 2005. Usage of Antimicrobial Agents and Occurrence of Antimicrobial Resistance in Norway. Tromsø/Oslo: 2006.
199. Llopis F, Grau I, Tubau F, Cisnal M, Pallares R. Epidemiological and clinical characteristics of bacteraemia caused by *Aeromonas* spp. as compared with *Escherichia coli* and *Pseudomonas aeruginosa*. *Scand J Infect Dis* 2004;36:335-41.
200. McCarthy N, Giesecke J. Case-case comparisons to study causation of common infectious diseases. *Int J Epidemiol* 1999;28:764-8.
201. Weinberg CR. Toward a clearer definition of confounding. *Am J Epidemiol* 1993;137:1-8.

202. Forskrift om innsamling og behandling av helseopplysninger i Meldingssystem for smittsomme sykdommer og i Tuberkuloseregisteret og om varsling om smittsomme sykdommer (MSIS- og Tuberkuloseregisterforskriften). FOR 2003-06-20 nr 740 (Regulations on collection and processing of health information in The Norwegian Surveillance System for Communicable Diseases and Tuberculosis Registry and on warning of communicable diseases.) [Lovdata]. [updated 2003 Jun 20; cited 2008 Feb. 26]. Available from: <http://www.lovdata.no/cgi-wift/ldles?doc=sf/sf/sf-20030620-0740.html>.
203. Walberg M, Frosli KF, Roislien J. Local Hospital Perspective on a Nationwide Outbreak of *Pseudomonas aeruginosa* Infection in Norway. *Infect Control Hosp Epidemiol* 2008;29:635-41.

Appendices

Paper I – IV

References

1. Arnow PM, Flaherty JP: **Nonfermentative Garm-negative bacilli**. In *Hospital epidemiology and infection control* Volume 27. 2nd edition. Edited by: Mayhall CG. Philadelphia, Lippincott Williams & Wilkins; 1999:431-451.
2. Pollack M: **Pseudomonas aeruginosa**. In *Principles and practice of infectious diseases* Volume 207. 5th edition. Edited by: Mandell GL, Bennett JE and Dolin R. Philadelphia, Churchill Livingstone; 2000:2310-2335.
3. Stephenson JR, Heard SR, Richards MA, Tabaqchali S: **Gastrointestinal colonization and septicaemia with Pseudomonas aeruginosa due to contaminated thymol mouthwash in immunocompromised patients**. *J Hosp Infect* 1985, **6**:369-378.
4. Becks VE, Lorenzoni NM: **Pseudomonas aeruginosa outbreak in a neonatal intensive care unit: a possible link to contaminated hand lotion**. *Am J Infect Control* 1995, **23**:396-398.
5. Silva CV, Magalhaes VD, Pereira CR, Kawagoe JY, Ikura C, Ganc AJ: **Pseudo-outbreak of Pseudomonas aeruginosa and Serratia marcescens related to bronchoscopes**. *Infect Control Hosp Epidemiol* 2003, **24**:195-197.
6. Srinivasan A, Wolfenden LL, Song X, Mackie K, Hartsell TL, Jones HD, Diette GB, Orens JB, Yung RC, Ross TL, Merz W, Scheel PJ, Haponik EF, Perl TM: **An outbreak of Pseudomonas aeruginosa infections associated with flexible bronchoscopes**. *N Engl J Med* 2003, **348**:221-227.
7. Cobben NA, Drent M, Jonkers M, Wouters EF, Vaneechoutte M, Stobberingh EE: **Outbreak of severe Pseudomonas aeruginosa respiratory infections due to contaminated nebulizers**. *J Hosp Infect* 1996, **33**:63-70.
8. Schelenz S, French G: **An outbreak of multidrug-resistant Pseudomonas aeruginosa infection associated with contamination of bronchoscopes and an endoscope washer-disinfector**. *J Hosp Infect* 2000, **46**:23-30.
9. Millership SE, Patel N, Chattopadhyay B: **The colonization of patients in an intensive treatment unit with gram-negative flora: the significance of the oral route**. *J Hosp Infect* 1986, **7**:226-235.
10. Foca M, Jakob K, Whittier S, Della LP, Factor S, Rubenstein D, Saiman L: **Endemic Pseudomonas aeruginosa infection in a neonatal intensive care unit**. *N Engl J Med* 2000, **343**:695-700.
11. Pena C, Dominguez MA, Pujol M, Verdager R, Gudolf F, Ariza J: **An outbreak of carbapenem-resistant Pseudomonas aeruginosa in a urology ward**. *Clin Microbiol Infect* 2003, **9**:938-943.
12. Bukholm G, Tannaes T, Kjelsberg AB, Smith-Erichsen N: **An outbreak of multidrug-resistant Pseudomonas aeruginosa associated with increased risk of patient death in an intensive care unit**. *Infect Control Hosp Epidemiol* 2002, **23**:441-446.
13. Lyytikäinen O, Golovanova V, Kolho E, Ruutu P, Sivonen A, Tiittanen L, Hakanen M, Vuopio-Varkila J: **Outbreak caused by tobramycin-resistant Pseudomonas aeruginosa in a bone marrow transplantation unit**. *Scand J Infect Dis* 2001, **33**:445-449.
14. Moolenaar RL, Crutcher JM, San Joaquin VH, Sewell LV, Hutwagner LC, Carson LA, Robison DA, Smith LM, Jarvis WR: **A prolonged outbreak of Pseudomonas aeruginosa in a neonatal intensive care unit: did staff fingernails play a role in disease transmission?** *Infect Control Hosp Epidemiol* 2000, **21**:80-85.
15. Bert F, Maubec E, Bruneau B, Berry P, Lambert-Zechovsky N: **Multi-resistant Pseudomonas aeruginosa outbreak associated with contaminated tap water in a neurosurgery intensive care unit**. *J Hosp Infect* 1998, **39**:53-62.
16. Grigis A, Goglio A, Parea M, Gneccchi F, Minetti B, Barbui T: **Nosocomial outbreak of severe Pseudomonas aeruginosa infections in haematological patients**. *Eur J Epidemiol* 1993, **9**:390-395.
17. Cheng K, Smyth RL, Govan JR, Doherty C, Winstanley C, Denning N, Heaf DP, van Saene H, Hart CA: **Spread of beta-lactam-resistant Pseudomonas aeruginosa in a cystic fibrosis clinic**. *Lancet* 1996, **348**:639-642.
18. Bergmans DC, Bonten MJ, van Tiel FH, Gaillard CA, van der GS, Wiltling RM, de Leeuw PW, Stobberingh EE: **Cross-colonisation with Pseudomonas aeruginosa of patients in an intensive care unit**. *Thorax* 1998, **53**:1053-1058.
19. Iversen BG, Jacobsen T, Eriksen HM, Bukholm G, Melby KK, Nygard K, Aavitsland P: **An outbreak of Pseudomonas aeruginosa infection caused by contaminated mouth swabs**. *Clin Infect Dis* 2007, **44**:794-801.
20. Council E: **European Council Directive 93/42/EEC of 14 June 1993 concerning medical devices** Council Directive 93/42/EEC edition. 1993 [http://europa.eu.int/smartapi/cgi/sga_doc?smartapi:celexapi:prod?CELEXnumdoc&lg=EN&numdoc=31993L0042&model=gui&etl].
21. **Water quality - Detection and enumeration of Pseudomonas aeruginosa by membrane filtration**; prEN 12780 Brussels, European Committee for Standardization; 2002.
22. **Ph.Eur 4 (2002: 2.6.12)**. In *European Pharmacopoeia* 4th edition. Strasbourg, Council of Europe; 2002.
23. **Ph.Eur 4 (2002: 2.6.13)**. In *European Pharmacopoeia* 4th edition. Strasbourg, Council of Europe; 2002.
24. **Ph.Eur 4 (2002: 5.1.3)**. In *European Pharmacopoeia* 4th edition. Strasbourg, Council of Europe; 2002.
25. Lassen J, Lingaas E: **Vurdering om produksjonsprosessen for DENT-O-SEPT munnpensel er forsvarlig [Assessment of whether the production process of the Dent-O-Sept mouthswab is safe]** Oslo, Sosial- og helsedirektoratet; 2002.
26. Kiska DL, Gilligan PH: **Pseudomonas**. In *Manual of clinical microbiology* Volume 47. 8th edition. Edited by: Murray PR. Washington DC, ASM Press; 2003:719-728.
27. Shirtliff ME, Mader JT, Camper AK: **Molecular interactions in biofilms**. *Chem Biol* 2002, **9**:859-871.
28. Hall-Stoodley L, Stoodley P: **Biofilm formation and dispersal and the transmission of human pathogens**. *Trends Microbiol* 2005, **13**:7-10.
29. Dunne WM Jr.: **Bacterial adhesion: seen any good biofilms lately?** *Clin Microbiol Rev* 2002, **15**:155-166.
30. Donlan RM, Costerton JW: **Biofilms: survival mechanisms of clinically relevant microorganisms**. *Clin Microbiol Rev* 2002, **15**:167-193.
31. Prince AS: **Biofilms, antimicrobial resistance, and airway infection**. *N Engl J Med* 2002, **347**:1110-1111.
32. Anderson RL, Holland BW, Carr JK, Bond WW, Favero MS: **Effect of disinfectants on pseudomonads colonized on the interior surface of PVC pipes**. *Am J Public Health* 1990, **80**:17-21.
33. Bo G: **Analyserapport. Pseudomonas aeruginosa i Dent-O-Sept [Analysis report. Pseudomonas aeruginosa in Dent-O-Sept]** Kristiansand, Naeringsmiddeltilsynet i Vest-Agder, Laboratorium; 2002.
34. **Ph.Eur 5 (2005: 5.1.4)**. In *European Pharmacopoeia* 5th edition. Strasbourg, Council of Europe; 2005.
35. Bergmans DC, Bonten MJ, Gaillard CA, Paling JC, van der GS, van Tiel FH, Beysens AJ, de Leeuw PW, Stobberingh EE: **Prevention of ventilator-associated pneumonia by oral decontamination: a prospective, randomized, double-blind, placebo-controlled study**. *Am J Respir Crit Care Med* 2001, **164**:382-388.
36. Yoneyama T, Yoshida M, Ohri T, Mukaiyama H, Okamoto H, Hoshiba K, Ihara S, Yanagisawa S, Ariumi S, Morita T, Mizuno Y, Ohsawa T, Akagawa Y, Hashimoto K, Sasaki H: **Oral care reduces pneumonia in older patients in nursing homes**. *J Am Geriatr Soc* 2002, **50**:430-433.
37. Tablan OC, Anderson LJ, Besser R, Bridges C, Hajjeh R: **Guidelines for preventing health-care-associated pneumonia, 2003: recommendations of CDC and the Healthcare Infection Control Practices Advisory Committee**. *MMWR Recomm Rep* 2004, **53**:1-36.
38. Alp E, Voss A: **Ventilator associated pneumonia and infection control**. *Ann Clin Microbiol Antimicrob* 2006, **5**:7.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:

http://www.biomedcentral.com/info/publishing_adv.asp



BioMed Central

Analytic perspective

Open Access

Questions on causality and responsibility arising from an outbreak of *Pseudomonas aeruginosa* infections in Norway

Bjørn G Iversen*¹, Bjørn Hofmann^{2,3} and Preben Aavitsland¹

Address: ¹Norwegian Institute of Public Health, Oslo, Norway, ²University College of Gjøvik, Faculty of Health, Care and Nursing, Gjøvik, Norway and ³University of Oslo, Department of General Practice and Community Medicine, Section for Medical Ethics, Oslo, Norway

Email: Bjørn G Iversen* - bjorn.iversen@fhi.no; Bjørn Hofmann - b.m.hofmann@medisin.uio.no; Preben Aavitsland - preben.aavitsland@fhi.no

* Corresponding author

Published: 23 October 2008

Received: 25 April 2008

Emerging Themes in Epidemiology 2008, **5**:22 doi:10.1186/1742-7622-5-22

Accepted: 23 October 2008

This article is available from: <http://www.ete-online.com/content/5/1/22>

© 2008 Iversen et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

In 2002, Norway experienced a large outbreak of *Pseudomonas aeruginosa* infections in hospitals with 231 confirmed cases. This fuelled intense public and professional debates on what were the causes and who were responsible. In epidemiology, other sciences, in philosophy and in law there is a long tradition of discussing the concept of causality. We use this outbreak as a case; apply various theories of causality from different disciplines to discuss the roles and responsibilities of some of the parties involved. Mackie's concept of INUS conditions, Hill's nine viewpoints to study association for claiming causation, deterministic and probabilistic ways of reasoning, all shed light on the issues of causality in this outbreak. Moreover, applying legal theories of causation (counterfactual reasoning and the "but-for" test and the NESS test) proved especially useful, but the case also illustrated the weaknesses of the various theories of causation.

We conclude that many factors contributed to causing the outbreak, but that contamination of a medical device in the production facility was the major necessary condition. The reuse of the medical device in hospitals contributed primarily to the size of the outbreak. The unintended error by its producer – and to a minor extent by the hospital practice – was mainly due to non-application of relevant knowledge and skills, and appears to constitute professional negligence. Due to criminal procedure laws and other factors outside the discourse of causality, no one was criminally charged for the outbreak which caused much suffering and shortening the life of at least 34 people.

Introduction

In 2002, we traced the source of a large outbreak of *Pseudomonas aeruginosa* infections to contaminated mouth swabs extensively used in Norwegian health care [1]. The investigation revealed many weaknesses and errors in the chain from production to use [2].

During and after the outbreak investigation, questions of causality, responsibility and liability were raised: Who

and what caused the outbreak, who were responsible for the extent of the outbreak, could the damages have been mitigated by acting sooner or differently, should anyone be punished? Questions of causality, responsibility and blame have always been a part of the history of infections. Two examples are the debate on where the Spanish flu came from and who was responsible for starting the Aids epidemic.

The concept of causality is intuitively simple and yet so intricately complex. In epidemiology causality has been hotly debated [3-11]. In philosophy of science there is a long tradition of discussing both the content of the term and how to achieve knowledge about the association of events [12]. In law, decisions on responsibility and liability rests on whether a specific action has *caused* specific harm or loss to another, and jurisprudence frequently defers to science in order to settle issues of causality [13-16]. However, not only is the discourse of causality in the philosophy of science interesting for law, reciprocally the debate on legal causation, especially in tort law, is useful for the scientists and the philosophers of science. In all these three disciplines (science, philosophy and law) and in practical life this discourse has implications for placing moral responsibility, blame, honour and dishonour. Consequently the general debate on causality is of interest both for scientists, manufacturers, and lawyers, as for the general public because it influences moral as well as professional norms.

In this article, we will use the outbreak of *P. aeruginosa* infections to illustrate the relevance of various theories of causality and discuss the role of the different participants. Then we will discuss the responsibility and fallibility for two of the main actors in the outbreak.

Setting the scene

Late February 2002, the Norwegian Institute of Public Health (NIPH) was alerted of a possible increase in the number of *Pseudomonas* infections in clinical wards of Norwegian hospitals [1]. After a strenuous outbreak investigation, on 8 April 2002, the outbreak strain was isolated from a domestically produced mouth swab for use in health care, called "Dent-O-Sept" (figure 1). The finding was publicised, the product was recalled, and the production ceased permanently.



Figure 1
The Dent-O-Sept mouth swab.

The outbreak strain was detected in swabs from 12 batches produced in 2001 and 2002 [2] and from the production line in the factory. An audit of the producer revealed several breaches of production regulations [17]. Health care institutions reported some extent of non-proper reuse of the swabs and weaknesses in their purchasing systems.

The strain was detected in 231 patients from 24 hospitals, of whom 71 (31%) died while hospitalised; all had severe underlying disease. For at least 34 patients the investigators concluded that the *P. aeruginosa* infection probably contributed to the patient's death [1]. No one was found criminally liable for the outbreak.

Two of the authors (BGI and PA) were responsible for the outbreak investigation at the Norwegian Institute of Public Health [1,2]. After six years have passed we feel that we can give a balanced review of the causes of the outbreak but will abstain from evaluating our role in it.

So, what was the cause of the outbreak, and who were responsible? Let us first examine the issues of causation from a scientific point of view, and then relate them to the legal issues of responsibility and liability.

Analysis

Before presenting theories on causality, responsibility and liability we need to define what was caused, i.e. what was the epidemiological outcome. We have asked "what caused the outbreak", but "the outbreak" is a rather diffuse concept and consists of the sum of individuals who each had their own set of factors contributing to them being included. Although the attention was brought to the individual cases by clinical manifestations (infections), we included in the outbreak all patients with genotypically identical strains of *P. aeruginosa*, irrespective of survival or severity of disease [1]. For this analysis we will make it clear which of the four different outcomes we have in mind; 1. being a case as defined in the outbreak investigation, 2. having a *P. aeruginosa* infection, irrespective of type of strain, 3. dying from *P. aeruginosa* infection or 4. the outbreak as a whole.

Causality

We often say that one thing causes another, like "rain causes flooding" and "smoking causes cancer", although it is not always true. We don't have a flood every time it is raining, and flooding can have other reasons than rain: a broken water pipe for example.

The philosophical basis of the dominant approach for testing theories in medicine is the hypothetico-deductive model as described by for example David Hume and Karl Popper. According to this model it is impossible to

achieve absolute proof for a scientific hypothesis; tests performed can only corroborate or falsify the hypothesis. Consequently one can never prove causality between factors and an outcome, only strengthen or weaken a proposed association. In this tradition Sir Austin Bradford Hill listed nine viewpoints from which to study the association of two variables in order to claim causation [3].

Causation in epidemiology

Classic epidemiology has been mainly backward looking, seeking an explanation to an event. In much of the 19th century there was a profound debate on what caused many of the major diseases of the time, being it miasmata (stench or bad air) or contagions [18]. For a disease like cholera John Snow, the father of epidemiology, was in favour of the theory of a contagion which he called "morbid matter" [19]. Late in the 19th century, a prominent microbiologist, Robert Koch, formulated a set of postulates that needed to be fulfilled in order to claim that a micro-organism caused a specific disease [20,21]. According to his postulates we need both necessary and the sufficient conditions to claim causal relationship between a microbe and a disease.

A century later MacMahon stated that there are two ways of classifying ill persons, either by *manifestational criteria* (grouping ill persons according to symptoms or clinical signs, e.g. common cold, schizophrenia or meningitis) or by *causal criteria* (grouping ill persons with respect to a specified experience believed to be a cause of their illness, e.g. lead poisoning or meningococcal disease) [22]. To have a *Pseudomonas aeruginosa* infection implies by name and definition causality of the bacterium.

Causation in law

The Norwegian legal system belongs to the French-German legal tradition which differs from the Anglo-American law in placing relatively more emphasis on statutory law than the judiciary legal institutions in the making of the legal framework. However, regarding tort law and causality the principles of the two legal systems are very similar. Likewise, both legal systems have a lower threshold for civil liability than criminal liability. There are several examples from recent history in Norway where the accused was found not guilty in the criminal court case but was convicted to pay economic compensation in a following civil lawsuit.

Causal connection in law is usually divided into two parts, "cause-in-fact" and "proximate cause" [16,23]. "Cause-in-fact" comes closest to what is regarded as causality in science. However, while science mostly deals with causal generalisation, law focuses more on causes of specific events. One standard method of establishing factual causation is the "but-for" test, aiming at excluding those fac-

tors that had no impact on the course of events. Another influential test for causation is the NESS-test, i.e. Necessary Element of a Sufficient Set test [23,24]. "Proximate cause", also called "adequate cause", embodies reasons for limiting the extent of legal responsibility and liability.

Additionally, deciding on legal responsibility and liability involves a counterfactual proposition, i.e., if a condition that in fact occurred had not occurred, then the outcome would have been different. Both the "but-for" test and the NESS test can be part of such counterfactual propositions. The "but-for" test asks: Would the consequences have occurred in these circumstances had the condition not been present? The NESS test asks: In these circumstances, is the condition a necessary member of a set of conditions that are together sufficient to produce the consequence [24]. Over-determination and joint determination are weaknesses of the "but-for" test, whereas lack of determination challenges the NESS test [22].

Counterfactual theories of causation in sciences

The central question in counterfactual theories of causation is "What would have happened if not event c had happened?" And the answer is: "If not event c had occurred, then the event e would not have occurred" [25]. Counterfactual reasoning can be used both in deterministic and probabilistic models. In daily life and in medicine counterfactual reasoning is extensively used. "If the needle hadn't been contaminated, the patient would not have acquired hepatitis." "If you hadn't been exposed to asbestos, you would not have contracted mesothelioma." Many of the epidemiological study designs have counterfactual thinking embodied [5]. In cohort studies we compare exposed and unexposed individuals for a certain risk factor. The unexposed group can be viewed as "what if this exposure did not occur". When calculating the attributable risk fraction, also called the etiological fraction, we assume that all association between the exposure studied and the outcome is causal, and in addition imply that if not the exposed group had been exposed, the rate of outcome among them would have been at the same level as among the unexposed.

Necessary, sufficient and complex conditions (determinism)

Many conditions are **necessary** for an event to occur. Every time the event occurs, the condition is present. A necessary condition for septicaemia is that one has blood; however, we do not say that having blood is the cause of sepsis. Owning the axe, which a person steals to kill a man, does not make you a murderer, even though the axe was a necessary condition for the man's death. This leads to the claim that causation is not given by the necessary conditions, although they are important, because if we can eliminate the necessary conditions, we can eliminate the problem.

Some conditions are **sufficient** to result in another: every time they occur, something else happens. Drinking a cup of hydrogen cyanide is a sufficient condition for death. However, other conditions may also result in the effect; not all deaths result from drinking hydrogen cyanide.

In complex situations many factors contribute to an effect, and there are logistic problems in that an event occurs some but not all times a constellation of factors occurs. To overcome this, Mackie introduced the so-called INUS condition of causation. An INUS condition for some effect is an Insufficient but Non-redundant part of an Unnecessary but Sufficient condition [26]. The NESS test described above is a clarification or specific instance of an INUS condition [23].

Determinism and probabilism

Causal determinism is based on the idea that that every event is necessitated by antecedent events and conditions together with the laws of nature [27]. According to causal determinism the causal relationships are invariant: Every time a certain configuration of conditions occurs, the outcome will be the same. We may have causal determinism even if the situation is complex and the outcome is hard to predict.

Probabilistic causality on the other hand claims that the causal relationship is probabilistic, and not invariant. That is, the outcome (effect) may vary according to probability distribution. Probabilistic theories of causation state that causes raise the probabilities of their effects [9].

In epidemiology, probabilistic approaches are most often used in the conceptual thinking of a relationship and in the statistical testing of the strength of association [9]. Here, Hill's set of nine viewpoints to explain the association between two variables are commonly used [3]. Only the one of temporal sequence of association is essential. This list of "Guidelines for causation" is more in tune with modern epidemiological science as they emphasize the strength of association rather than pure mechanical determinism. However, many have criticised Hill's list and in recent years there has been a resurgence in the debate about causality [6-8,10,28]. Moreover, probabilistic graphical methods, such as Bayesian networks, may also be used in order to represent the probabilistic independencies between variables.

Causation in the Dent-O-Sept case

We have now presented theories for causation, which can now be applied in a specific case, the Dent-O-Sept outbreak in Norway in 2002 [1,2].

Necessary and sufficient conditions

The *P. aeruginosa* bacterium is not a necessary and sufficient condition for the death of people. Neither is its presence in the production plant a necessary and sufficient condition for the presence of the bacteria in the product. Hence, if necessary and sufficient conditions are required for liability and moral responsibility, no one is responsible for the outbreak. Was the *P. aeruginosa* bacterium a necessary condition?

For patients involved in the Dent-O-Sept outbreak, having the outbreak strain of *P. aeruginosa* was **necessary** to be included as a case. If it was not for the *P. aeruginosa*, then there would have been no outbreak (i.e. "but-for"). But this is more a definition criterion for being a case and does not explain *why* the patient harboured this strain.

No single factor was absolutely required to be colonised or infected with the outbreak strain. The use of Dent-O-Sept was not a necessary condition for infection. Approximately one third of the cases had not used the swab directly. The outbreak investigators concluded that they probably were secondarily infected from contaminated environment or health care workers after the contaminated swabs had introduced the strain in the hospital environment. By including this indirect pathway, it is reasonable to claim that the contaminated swabs were a necessary condition for the patients to become infected. This is equivalent to outbreaks of gastroenteritis (e.g. salmonellosis) where the primary cases may be infected by contaminated food, but cases continue to occur by person to person transmission via the fecal-oral route even after the implicated food item has been removed. In these situations, we would usually say that the food contamination caused the whole outbreak, and not only the first cases.

P. aeruginosa is harmless to most people and in most instances. The large majority of patients with the outbreak strain of *P. aeruginosa* and all who died from the infection had severe underlying illnesses. To have a severe underlying illness was in practice a necessary condition to die from the outbreak strain. So, both the presence of *P. aeruginosa* in Dent-O-Sept and having an underlying illness were necessary conditions for dying from the infection. But there were other necessary conditions as well, such as being hospitalised, but this we would hardly call a cause of death. This illustrates the problem with necessary conditions: there are extremely many of them.

In the Dent-O-Sept outbreak no single condition was **sufficient** to result in infection with the outbreak strain. Given the large number of Dent-O-Sept swabs used in the period and the massive contamination, we believe that several thousand patients were exposed to the outbreak strain of *P. aeruginosa*. Only a few of them became last-

ingly colonised or infected. Hence, the contaminated Dent-O-Sept swab was not a sufficient condition for the outbreak of the infection.

We may visualise a chain of unfortunate events necessary for the outbreak to occur: The outbreak investigation discerned the direction of flow of the *P. aeruginosa* bacteria from the production to the patients (figure 2). Can the links of this chain be seen as a series of necessary conditions that together are sufficient for the outbreak? However, the (necessary) conditions, such as the presence of *P. aeruginosa* in the mouth swabs and the patients' susceptibility, are not sufficient for the outbreak. There will not be an outbreak every time these conditions occur. Applying the concept of the INUS condition is helpful [26]. Using contaminated swabs was in itself insufficient but non-redundant, but together with other factors like the susceptibility of the patients, the infectious dose, and underlying illnesses, became sufficient to infect or colonise the patient.

According to this approach to causality we can find the altogether necessary and sufficient conditions for an event. If we know the conditions (making up the cause), the effect will occur. However, the challenge is that we do not know all the conditions and their complex interplay. For instance, we do not know the importance of the water quality during production or the significance of the reuse of the Dent-O-Sept for the outbreak.

Hill's postulates

In the Dent-O-Sept case, most of the points on Hill's list are helpful in order to claim association, at least to a certain degree (table 1). In addition to the traditional epidemiological measures of strength of association modern microbiology has developed tools to demonstrate associations. Various techniques of producing "fingerprints" of bacterial DNA have made it possible to identify identical strains of bacteria. In outbreak investigations these techniques have become very important to connect events and

claim causality, and are similar to detection of human DNA in criminal investigations. When genotypically identical bacterial isolates were detected in the production plant, in some of the swabs and in the affected patients, we concluded that contamination of the production line caused the outbreak.

Key actors

We will now in some more detail analyse key actors (table 2) and events in the light of causation and prepare for the next section on moral responsibility.

The producer, Snøgg AS had a routine for cleaning and disinfecting the production line but had no quality assurance (QA) systems in place to control the production and verifying that the product was safe. Having QA systems is one of several requirements by Norwegian and EU law [29] to be able to mark the product "CE" (Communauté Européenne) which signifies that it complies with relevant EU-regulations and indicates it being safe. Some years prior to the outbreak, customers had periodically complained about discoloured swabs. In 1999, the producer commissioned an external evaluation and implemented some, but not all the advice given; among the latter was the advice to establish microbiological quality control of the final product [17].

An audit of the producer revealed serious breaches of production regulations [17]. Under strict liability, a party breaking the law may be legally responsible irrespective of whether any harm has been caused [24]. For instance, driving under the influence of alcohol is in Norway as in many other countries punishable by law even when no one is harmed.

The presence of the outbreak strain in the production facility was necessary for contamination to occur, but not sufficient as not all swabs were contaminated. This differed even in swabs produced at the same time of day on the same date. The main hypothesis of the outbreak inves-

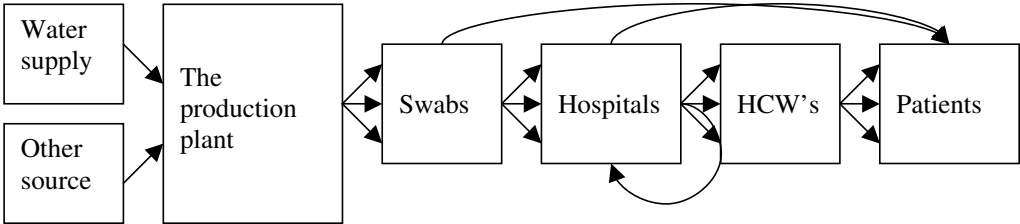


Figure 2
The direction of flow of the *Pseudomonas aeruginosa* bacteria from the production to the patients.

Table 1: Application of Hill's viewpoints on the causal association between the Dent-O-Sept swab and becoming colonised or infected during the outbreak

Hill's viewpoints	Application on the Dent-O-Sept outbreak
1. Strength of association	Strong. Association for having used the swab during hospitalisation and having the outbreak strain of <i>P. aeruginosa</i> , adjusted odds ratio 5.3. Detecting genotypically identical strains of the bacterium in patients, the product and in the production facility [1]
2. Consistency of association	Yes, to a large extent. However, other co-factors also needed to be in place, e.g. contamination of the particular swab and a susceptible patient. Due to secondary spread in the hospitals also patients who did not use the swab were infected.
3. Specificity of association	Yes, mostly. Use of contaminated swabs led to colonisation and some times to infection. Necessary co-factors were as above (2). The clinical manifestations of the <i>P. aeruginosa</i> infection varied widely.
4. Temporal sequence of association	Yes. However, the outbreak strain of the bacteria was found in six patients before the production of the first contaminated batch of swabs was detected [2]. When the swabs were withdrawn from the market the number of cases gradually diminished and disappeared.
5. Biological gradient	This was not tested but assumed. Reuse of the swabs may have increased the bacterial load and hence the risk of becoming infected.
6. Plausibility of association	Yes. The chain from contamination during production to infection is well described.
7. Coherence of association	Yes. There is no other hypothesis of explanations for the outbreak.
8. Experiment (reversibility)	Yes, a natural experiment. When the source was removed the number of cases gradually diminished to zero.
9. Analogy	Yes. There are many other outbreaks caused by medical devices. (References in [1])

tigation team was that bacteria-containing biofilm was randomly shed from the production equipment into the swab wraps [2]. Using the judicial, counterfactual "but-for" test: "But-for the absence of a microbiological quality control of the production equipment and the final product, would the contamination have been detected earlier and the outbreak avoided?" There is good reason to believe so. A total of 76 of 1565 swabs examined during the outbreak investigation contained the outbreak strain of *P. aeruginosa*, and more than 250 swabs (16%) contained one or more microbes (including the outbreak strain), also in swabs produced years before the Dent-O-Sept outbreak started [2]. Consequently, not abiding by the requirement to have QA systems including an effective microbiological control system can be seen as a cause in the legal sense for this outbreak to occur. The same goes if we apply a counterfactual NESS test.

The *P. aeruginosa* bacterium was also detected in a rubber hose leading from a water tap supplied with municipal water to a large steel tank used in the production [2]. In a press release the producer claimed that the municipal water company caused the outbreak [30]. This claim of the origin of the first bacterium could in retrospect not be verified. However, there are no requirements for tap water to be *Pseudomonas* or bacteria free. On the contrary, it is

common microbiological knowledge that *P. aeruginosa* at times can be detected in water and soil [31,32]. The bacteria could have originated from other sources and contaminated the rubber hose. Using the "but-for" test on the water supply fails to show it to be a cause-in-fact due to the uncertainty of the origin of the first bacteria. Likewise, due to the uncertainty of the origin it fails a counterfactual NESS test. Hence, bacteria in the tap water cannot be seen as a legal cause of the outbreak.

Many of the hospitals had several deficiencies in their QA systems, for instance concerning the selection of which products to purchase; the actual procurement of the product; the logistic system for reception, distribution, storage, and use of the product [33]. Many of these deficiencies are in breach of national guidelines and legal regulations but did probably not have any influence on this particular outbreak. The bacterial load inside the wrap probably diminished over time as the bacteria cannot survive without oxygen. Consequently, the deficient logistics systems in the hospitals appear not to be a cause-in-fact of this outbreak. However, competent procurers in hospitals might have detected inferior products or the lack of documentation, such as the declaration of conformity which is required for all CE marked medical devices with the EU directive [29].

Table 2: The main participants in the Dent-O-Sept outbreak and some of their roles, responsibilities and actions.

Participant	Role and responsibilities
The producer "Snøgg AS"	<ul style="list-style-type: none"> • Produced the Dent-O-Sept swab • Did not adhere to the laws and regulations for production of medical devices • Lacked a quality assurance system for the production • Did not implement advice after external evaluation • Stopped the swab production as soon as the connection with the outbreak was established
The water supplier "Kristiansand municipality"	<ul style="list-style-type: none"> • Supplied drinking water to the producer • The <i>P. aeruginosa</i> bacterium may have been introduced into the production plant with the water
The hospitals	<ul style="list-style-type: none"> • Treated patients and procured medical devices • Many lacked quality assured systems for procurement, storage and use of medical devices • Many lacked systems for training of health care workers • Many had inadequate reporting systems for faulty medical devices
The health care workers	<ul style="list-style-type: none"> • Treated and cared for patients • Many reused the "single use" swabs contrary to the text on the wrap • Many did not report faulty medical devices
The patients	<ul style="list-style-type: none"> • Received medical treatment and care • Many were seriously ill and susceptible for contracting infections with the <i>P. aeruginosa</i> bacterium
The surveyor and investigator "Norwegian Institute of Public Health"	<ul style="list-style-type: none"> • Responsible for surveillance of infectious diseases and for outbreak investigations • There is no national surveillance system for <i>P. aeruginosa</i> infections
National administrative body "The Norwegian Directorate for Health and Social Affairs"	<ul style="list-style-type: none"> • Responsible for national administration within certain areas of the health care system • Responsible for the audit of the producer • Ignored the deadline to appeal the police's decision not to press charges.

Health care workers (HCWs) are constantly told to be economical and prudent in the use of medical equipment. In many hospitals it was customary to reuse the swab in the same patient although it was clearly marked as single use equipment. In between cleaning the patient's mouth the swab was sometimes stored in a glass of water on the patient's night stand. This practice allowed rapid multiplication of bacteria on the swab. This unprescribed use did not introduce the bacteria in swabs and hence in patients where it had not already been, but probably increased the bacterial load. An increase in bacterial load increases the risk of infecting a contaminated patient and of causing a more serious disease. "But-for" the improper use of the swab, the same number of patients would be exposed to the *P. aeruginosa* bacteria, but probably fewer would have been colonised and probably fewer colonised patients would have contracted serious infections. Together with the introduction of the outbreak strain into the hospitals

and the susceptibility of the patients, the reuse of the swabs can be seen as a necessary (non-redundant) condition or as a counterfactual conditional. Moreover, the improper use of the swab may fail a but-for-test (due to joint determination), but not a NESS-test. Hence, it is not clear that HCWs behavior caused the outbreak in a legal sense.

Hospital reporting systems for faulty medical equipment are not the same for all types of equipment. A general attitude among HCWs is that reporting is fruitless and not really necessary, especially for minor products like mouth swabs. During the investigation we learnt that several HCWs had detected discoloured or otherwise faulty swabs without reporting the event. It is worth noting that the notification of faulty swabs by an infection control nurse contributed to solving the outbreak quicker.

"But-for" the lack of reporting it is impossible to ascertain whether the outbreak would have been detected earlier as it depends on many other factors like what was reported, how it was reported in the system and what measures were taken following a report. In a probabilistic or risk assessment approach, low threshold reporting systems with appropriate follow-up routines and adequate surveillance systems, make it more likely that the contaminated swabs and the outbreak would have been detected earlier (counterfactual).

Surveillance systems are in place for many infectious diseases, but not for *Pseudomonas* infections [34]. Only the most prevalent or serious diseases are included into the surveillance systems after weighing factors such as costs and preventability. When the computer systems improve, more infections can be included at no or little extra cost. As concluded in the previous paragraph, had an adequate surveillance system for the *P. aeruginosa* bacterium or some of the infections it caused been in place, it is more likely that the outbreak could have been detected earlier. However, the main importance of reporting systems for faulty equipment and surveillance systems is that of preparedness. Had the swabs not been contaminated, imperfect reporting and surveillance systems do not add to the risk of causing outbreaks like unclean production of medical equipment does (counterfactual).

The outbreak investigation was a necessary condition to stop the outbreak. Could the number of patients affected have been smaller if the investigation had been carried out differently? During the investigation there was a tremendous pressure to find the solution quickly. A rushed investigation might have resulted in not detecting the cause or getting the results wrong, whereas a broad, systematic investigation might have taken too long causing unnecessary sufferings and deaths. As two of the authors of this article (BGI and PA) were responsible for the outbreak investigation [1,2] we are not competent to appraise the investigation.

In conclusion, many factors contributed to the outbreak and its eventual dimension. The main necessary condition for the outbreak was the contamination of the swabs in the production facility. The size of the outbreak measured in the number of patients affected and how long it lasted are due to several additional factors. The breaches of regulations by the producer of the swabs play an important role probably together with the reuse of the swabs in the hospital, i.e. they are conditions that are influencing the size of the outbreak. With a regulatory correct production of the swabs in the production facilities there would have been no outbreak (necessary condition). The reuse of the swabs in the hospitals and the non-optimal production probably increased the size of the outbreak (probabilistic

factor). In addition, other factors that might have an influence are the lack of adequate reporting and surveillance systems.

Moral responsibility

Many people suffered in the outbreak. Seventy-one people with the outbreak strain died while hospitalised, and for at least 34 the investigators concluded that the *P. aeruginosa* infection probably contributed to the death. No one was found criminally liable. Several actors were in a position where they could have known and acted differently, and hence, are to be seen as morally responsible. In the discussion on causes for the outbreak two main actors emerged in the discussion on responsibility. One is the producer Snøgg AS. The other is the group of HCWs who reused the swabs and the hospital system permitting these acts or possibly even encouraging them. What is their moral responsibility?

Traditionally, medical errors have been divided into three: Unintentional error, intentional error and random mishaps. In addition, bad outcomes may happen without error [35]. For our discussion we will only focus on **unintentional error** as no one in this outbreak ever was suspected of intentionally wanting to cause harm. Unintentional error can be caused by lack of knowledge, lack of skill or non-application of relevant knowledge or skill.

The producer Snøgg produces a wide range of medical equipment useful for saving lives and reducing suffering. The Dent-O-Sept swab had been produced for decades and was in great demand. Their vision statement is "gjøre det enkelt å hjelpe" [Make it simple to help] <http://www.snogg.no>. In all their appearances the producer gave no impression of intending to harm anyone, and from a virtue-ethical standpoint, the company appeared favourably (see endnote 1). When the connection between the swab and the outbreak was detected, the director of Snøgg was devastated for what his product had caused [36,37]. Some months later the company started to focus more on other factors influencing the outbreak. One of their new initiatives was to partly blame the outbreak on the introduction of the *P. aeruginosa* bacterium into the production facilities through the municipal water pipe [30]. Another was to draw the attention to the incorrect use of the swabs in hospitals [38]. The company is expected to know that they needed to have systems in place to stop bacteria in municipal tap water from reaching the end product.

The impetus not to harm patients ("Primum non nocere" – "First do no harm" ascribed to Hippocrates) and to care for the vulnerable are duties with strong deontologic bearings (see endnote 2). The duty to acquire necessary knowledge for the safe performance of health care services, as

well as being precautionary appear to be part of such a perspective. Hence, the actions of the producer (as well as the health care professionals) appear to breach with basic deontological bearings in health care. Moreover, if the moral norms of the producer's responsibilities are adequately regulated by law, breaking these legal regulations, such as the Act on medical devices [39], would in most cases be a breach with the moral duties of a producer.

The Dent-O-Sept mouth swab belongs to Medical device Class 1, which includes most non-invasive medical devices according to the European Council Directive 93/42/EEC [29]. The directive states that the devices must, when used, "not compromise the clinical condition or the safety of patients". "The devices and manufacturing processes must be designed in such a way as to eliminate or reduce as far as possible the risk of infection to the patient, user and third parties." Accordingly, the Council Directive represents a consequentialist approach (see endnote 3). The producer did not abide by the laws and regulations relevant to him, and thus ignoring relevant norms and relevant consequences.

The unintended error of producing contaminated swabs appears only to a small extent to result from lack of knowledge. The producer knew there had been problems in the production and had received advice on implementing QA systems which the producer had not followed in great detail. By not doing so there appears to be a non-application of relevant knowledge which would normally be characterized as negligent and culpable. However, there probably was a lack of knowledge about how bacteria can contaminate the production equipment. The Dent-O-Sept swab was the only moist item Snøgg produced. Had the microbiological quality control measures been implemented, this would probably have been revealed and harm could have been avoided. In addition, there probably also was some lack of skill in cleaning and disinfecting the wet part of production.

Hence, the outbreak was not a result of wilful or intentional error. However, the non-adherence to norms and regulations and the non-application or non-acquisition of knowledge can be conceived of as malpractice. It is not the case that science has not yet progressed enough, or that there are limitations in the predictive nature of knowledge with regards to the particular case [35].

The health care workers aim at saving lives and alleviate pain and suffering. Their work is legally regulated by laws and regulations, and professionally by guidelines, instructions and training. In addition, their actions are also to a large degree guided by colleagues and the culture of the workplace. One of the traditional Norwegian virtues of is that of austerity. It can partly be ascribed to the nation's

economic poverty up until a few decades ago and to our Lutheran tradition of modesty ("In the sweat of thy face shalt thou eat bread", Genesis 3:19). This demand of being economical is also reflected in the Act on health personnel [40] and in instructions from the hospital management. Single use products are in conflict with being economical. In several hospitals it was accepted or even encouraged to reuse the swabs, possibly considering them to be a variant of the tooth-brush. There is also a consequentialist reasoning for this austerity: reduced cost combined with low risk.

The Norwegian and English texts on the wrap were quite different, and the Norwegian being the most ambiguous: "Antiseptisk engangspensel for munnhygiene" which literally translates to "Antiseptic one-time-swab for mouth hygiene"; whereas the English text read: "Premoistened foam swab for mouth hygiene". Although "engangs" usually is translated to "single use" some can also understand it to be "single period-use" just like a single use syringe can be used for multiple injections in the same patient within a short time frame. To avoid possible misinterpretations a resent amendment (05.09.2007) to the European Council Directive on Medical devices has defined a "single use device" as "a device intended to be used once only for a single patient". The claim of antiseptic properties of the swab (which was never documented by the producers [17]) may have led some health care workers to underestimate the risk this practise posed. Placing a swab coated with an oral cavity bacterial flora in a glass of water with saliva and mucus as nutrition, may lead to extensive bacterial growth up to a concentration which makes it potentially harmful. And no one could presume that the swabs contained *P. aeruginosa*. In addition, there is an active debate within the medical community in Europe whether it can be safe to reuse reprocessed single use medical equipment.

There were no guidelines against this practice and no superiors contradicted it. From a virtue-ethical perspective the act was ambiguous; it was austere, but against professional standards (of following written instructions) and the duty to care for the patient. The main reason for the medical error of reusing the swab was non-application or misunderstanding of relevant information.

According to Norwegian law, hospital management shall provide for making an infection control programme, producing guidelines to prevent hospital acquired infections, having a system for surveillance of infectious diseases and for procurement and control of medical equipment. As there were deficiencies in many of these fields in many hospitals, the hospital management consequently appears to be morally negligent and legally responsible according to the NESS-test. When human error repetitively occurs

within a system it is of interest to discuss whether to have a person approach or a system approach. If preventing future errors is the aim, a system approach appears to be more rewarding [41]. However, even though responsibility of the management does not free the individual employees from responsibility, it would be fruitless to try to identify individual health care workers reusing the swab and place them under moral and legal scrutiny.

After the Dent-O-Sept swab was withdrawn from the market, other similar products have taken its place. Despite all the media attention from this outbreak, we have received anecdotal information that health care workers still reuse single use mouth swabs.

Legal consequences

At least 231 patients contracted the bacterium and for at least 34 patients the investigators concluded that the *P. aeruginosa* infection probably contributed to the patient's death. There was much anxiety and guilt feeling among patients and relatives. Many had cared for their terminally ill relatives and used the Dent-O-Sept swab. Some called the Norwegian Institute of Public Health and asked for example: "Did I kill my mother by using the swab?"

No one was made criminally liable after this outbreak. The police started an investigation of the producer but decided not to press charges. The Norwegian Directorate for Health and Social Affairs appealed the decision several months after the time limit for appeal had expired; hence the Attorney-General could not reopen the case [42,43].

Norsk pasientskadeerstatning (NPE, the Norwegian System of Compensation to Patients) grants monetary compensation mainly for economical loss and to some degree for permanent disablement due to injury inflicted as a result of treatment in public health services in Norway [44,45]. Few patients could document economical loss because they were, among other things, elderly, disabled, not working, had severe chronic diseases or were already severely injured e.g. after serious car accidents.

By 18 February 2004, NPE had received a total of 287 claims. Of 256 claims processed 48 were accepted for compensation and 2.3 million NOK (\approx 290 000 EUR) had been awarded [46]. By June 2007 the total number of processed claims was 291 of 292. NPE sent a claim for re-compensation to the producer. In an out-of court settlement dated 11 October 2005 the producer agreed to pay NPE 1.2 million NOK (\approx 150 000 EUR) as a full and final sum of any regress demand in connection with the mouth swabs and without accepting responsibility for the outbreak (Deputy Director General R. G. Jørstad, NPE, personal communication).

In addition to the claim from NPE other civil claims were made against the producer. One large hospital reached a court settlement on 19 June 2006 with the producer and received compensation amounting to 3.3 million NOK (\approx 410 000 EUR) for additional costs incurred for prolonged hospitalisations of patients and for preventing further spread of the bacterium [47]. There may be other settlements that not have been made public. Hence the total known compensations paid by the producer amounts to 4.5 million NOK (\approx 560 000 EUR).

In jurisprudence responsibility is related to causality. To be negligent in most instances requires to have *caused* the harm. In this article we have argued that the contamination of the Dent-O-Sept swab was a necessary (non-redundant) condition for colonising and infecting the individual patients and by this "caused" the outbreak. Furthermore, there is good reason to believe that if systematic microbiological sampling of the product as part of a QA system had been in place, microbial contamination of the product would have been detected. That is, according to counterfactual probabilistic reasoning, the neglected QA system "caused" the outbreak. This claim uses both deterministic arguments (swab causes outbreak) and probabilistic reasoning (the probability that a microbiological QA system will detect the contamination). In addition many other factors contributed to the number of patients being affected (the extent of the outbreak), like the susceptibility of each individual patient being exposed, the reuse of the swab by HCWs and the hospital attitude for accepting reuse of swabs, to name a few. Several of these factors can be interpreted as INUS conditions and also play a role in applying Hill's nine viewpoints in claiming a causal association between the swab and *Pseudomonas* infection (table 1). The case also illustrates the weaknesses of the "but-for" test (with regards to assessment of cause-in-fact) as there were many concurrent factors that cannot be differentiated as necessary for the event, they were only necessary elements (NESS-test).

After establishing a cause-in-fact relationship, the proximity or adequacy of causes needs to be discussed. Whereas the contamination of the swabs during production appears to be an adequate cause, the possibility of the origin of the first bacterium through the municipal water supply is not because it is neither illegal nor verified. In addition, it precedes a "*novus actus*" which is the breach of regulations in the production. It can be argued that the reuse of the swabs in the hospitals is also proximate, at least for some of the patients becoming infected during the outbreak. Whether the individual HCW or the hospital system is responsible is open for debate [41].

Why was no one criminally charged in this outbreak? We have argued for a causal association between the contaminated swabs and *Pseudomonas* infection and death, and a breach of regulations during production has been established. However, the error was unintentional and due to a non-application of relevant knowledge and skill; a knowledge that isn't intuitively evident to everyone. The police decided not to press charges. In a press release, the police pointed at other circumstances, arguing that the health authorities had not audited the producer prior to the outbreak, and that there were irregularities in the use of the swabs in the hospitals and in the reporting of faulty medical devices. In addition, the fact that the Directorate for Health and Social Affairs did not appeal the decision until too late blocked the possibility for a reinvestigation of the case due to the procedures described in the Criminal Procedure Act.

Conclusion

The major necessary condition causing the outbreak was the contamination of the swabs in the production facility. Without this contamination, the Dent-O-Sept outbreak would not have happened. Hence, there exists a cause-in-fact according to the but-for-test. Many other factors contributed to the outbreak and the size of it, the reuse of the single use swabs being the most important. The unintended error – by the producer of the swabs and to a minor extent by the hospital practice – was mainly due to non-application of relevant knowledge and skills, and breaches with moral duties as professionals, constituting moral negligence.

In epidemiology, other sciences, philosophy and jurisprudence there are plenty of methods and theories to explain causality and responsibility in complex situations like outbreak investigations. Applying different theories from different disciplines on the various necessary and sufficient conditions and the roles and responsibilities of the participants, is useful and important to elucidate the complex from most angles. From an outbreak investigator's viewpoint no theory is the only correct one. Using Mackie's concept of INUS conditions and Hill's nine viewpoints of claiming a causal association, applying deterministic as well as probabilistic ways of reasoning, all shed light on the issues of causality in this outbreak. Medical practice and jurisprudence is closely connected in real life as professional negligence can have legal consequences. Cases in epidemiology, such as outbreak investigation, highlight the tension both in science and jurisprudence between general causality and the causality of specific events. Moreover, applying legal theories of causation (counterfactual reasoning and the "but-for" test or the NESS test) proved important perspectives on the Dent-O-Sept outbreak.

As shown for the outbreak of *P. aeruginosa* infection the issue of causality also serves as a starting point for the debate on legal responsibility. Due to criminal procedure laws and other factors outside the discourse of causality, no one was criminally charged for the outbreak.

Abbreviations

CE: Communauté Européenne; DNA: Deoxyribonucleic acid; EU: European Union; EUR: Euro (currency); HCW: Health care worker; ICU: Intensive care unit; INUS: Insufficient but Non-redundant parts of an Unnecessary but Sufficient condition; NOK: Norwegian kroner (currency); NPE: Norsk pasientskadeerstatning, (The Norwegian System of Compensation to Patients); QA: Quality assurance.

Competing interests

Two of the authors (BGI and PA) were responsible for the outbreak investigation at the Norwegian Institute of Public Health. The authors declare that they have no competing interests.

Endnotes

Endnote 1

Virtue ethics is a branch of moral philosophy that emphasizes character as the key element of ethical thinking, rather than rules or consequences.

Endnote 2

Deontological ethics, deontology or duty-based-ethics is an approach to ethics that focuses on the rightness or wrongness of actions themselves, as opposed to the rightness or wrongness of the consequences of those actions. The term deontology stems from Greek: deon (δὸν) which means obligation or duty.

Endnote 3

Consequentialism refers to those moral theories which hold that the basis for any valid moral judgment about an action is the consequences of the particular action. Accordingly a morally right action is an action that produces good consequences.

Authors' contributions

BGI headed the outbreak investigation and the conception, drafting and revision of the manuscript. BH has contributed with regards to the causality theories and on the relationship between scientific and moral/legal causation, as well as revising the manuscript. PA was over all in charge of the outbreak investigation and participated in the revision of the manuscript. All authors read and approved of the final manuscript.

References

- Iversen BG, Jacobsen T, Eriksen HM, Bukholm G, Melby KK, Nygard K, Aavitsland P: **An outbreak of *Pseudomonas aeruginosa***

- infection caused by contaminated mouth swabs. *Clin Infect Dis* 2007, **44**:794-801.
2. Iversen BG, Eriksen HM, Bo G, Hagestad K, Jacobsen T, Engeset E, Lassen J, Aavitsland P: **Pseudomonas aeruginosa contamination of mouth swabs during production causing a major outbreak.** *Ann Clin Microbiol Antimicrob* 2007, **6**:3.
 3. Hill AB: **The environment and disease: Association or causation?** *Proceedings of the Royal Society of Medicine - London* 1965, **58**:295-300.
 4. Hofer M: **Getting causal considerations back on the right track.** *Emerg Themes Epidemiol* 2006, **3**:8.
 5. Hofer M: **Causal inference based on counterfactuals.** *BMC Med Res Methodol* 2005, **5**:28.
 6. Hofer M: **The Bradford Hill considerations on causality: a counterfactual perspective.** *Emerg Themes Epidemiol* 2005, **2**:11.
 7. Phillips CV, Goodman KJ: **Causal criteria and counterfactuals; nothing more (or less) than scientific common sense.** *Emerg Themes Epidemiol* 2006, **3**:5.
 8. Lipton R, Odegaard T: **Causal thinking and causal language in epidemiology: it's in the details.** *Epidemiol Perspect Innov* 2005, **2**:8.
 9. Hitchcock C: **Probabilistic Causation** [Stanford Encyclopedia of Philosophy]. Updated 2002, cited 2007 Nov 6 [http://plato.stanford.edu/entries/causation-probabilistic/].
 10. Rothman KJ, Greenland S: **Causation and causal inference.** In *Modern Epidemiology* Edited by: Rothman KJ, Greenland S. Philadelphia: Lippincott Williams & Wilkins; 1998:7-28.
 11. Vineis P: **Causality in epidemiology.** *Soz Präventivmed* 2003, **48**:80-87.
 12. Psillos S: **Causation and Explanation (Central Problems of Philosophy)** Chesham: Acumen Publishing Limited; 2002.
 13. Hart HLA, Honoré AM: **Causation in the Law** 2nd edition. Oxford: Clarendon; 1985.
 14. Stapleton J: **Legal Cause: Cause-in-Fact and the Scope of Liability for Consequences.** *Vanderbilt Law Review* 2001, **54**:941-1000.
 15. Green L: **The Causal Relation Issue in Negligence Law.** *Michigan Law Review* 1962, **60**:543-576.
 16. Ozonoff D: **Legal causation and responsibility for causing harm.** *Am J Public Health* 2005, **95**(Suppl 1):S35-S38.
 17. Sluttrapport fra tilsyn med Snøgg Industri AS (Final report from the audit of Snøgg Industri AS) Oslo: Sosial- og helsedirektoratet; 2003.
 18. Johnson S: *The Ghost Map* London: Allen Lane, Penguin Books Ltd; 2006.
 19. Snow J: *On the mode of communication of cholera* 2nd edition. London: John Churchill; 1855.
 20. Koch R: **Die aetiologie der Tuberkulose.** In *Gesammelte Werke von Koch* Edited by: Schwalbe J. Leipzig: Georg Thieme Verlag; 1912:428-455.
 21. Evans AS: **Causation and disease: the Henle-Koch postulates revisited.** *Yale Journal of Biology and Medicine* 1976, **49**:175-195.
 22. MacMahon B, Pugh TF: *Epidemiology Principles and methods* Boston: Little, Brown and Company; 1970.
 23. Honoré AM: **Causation in the law** [Stanford Encyclopedia of Philosophy]. Updated 2005, Oct 12; cited 2008 Feb 27 [http://plato.stanford.edu/entries/causation-law/].
 24. Honoré T: **Necessary and sufficient conditions in tort law.** In *Philosophical foundations of tort law* Edited by: Owen DG. Oxford: Clarendon Press; 1995:363-385.
 25. Dowe P: **Counterfactual Theories of Causation** [Stanford Encyclopedia of Philosophy]. Updated 2001 Jan 12; cited 2008 Feb 29 [http://plato.stanford.edu/entries/causation-counterfactual/].
 26. Mackie J: *The cement of the universe* Oxford: Clarendon Press; 1974.
 27. Hofer C: **Causal Determinism** [Stanford Encyclopedia of Philosophy]. Updated 2008, Jan 23; cited 2008 Feb 28 [http://plato.stanford.edu/entries/determinism-causal/].
 28. Phillips CV, Goodman KJ: **The missed lessons of Sir Austin Bradford Hill.** *Epidemiol Perspect Innov* 2004, **1**:3.
 29. European Council: *European Council Directive 93/42/EEC of 14 June 1993 concerning medical devices, Council Directive 93/42/EEC edn 1993.*
 30. **Drikkevannet årsak til bakterieskandale (Drinking water caused bacteria scandal)** [VG Nett]. Oslo. Updated 2002, Jul 18; cited 2008 Feb 26 [http://www1.vg.no/helse/artikkel.php?artid=7551076].
 31. Pollack M: **Pseudomonas aeruginosa.** In *Principles and practice of infectious diseases* Edited by: Mandell GL, Bennett JE, Dolin R. Philadelphia: Churchill Livingstone; 2000:2310-2335.
 32. Kiska DL, Gilligan PH: **Pseudomonas.** In *Manual of clinical microbiology* Edited by: Murray PR, Washington DC: ASM Press; 2003:719-728.
 33. **Forskrift om internkontroll i sosial- og helsetjenesten. FOR 2002-12-20 nr 1731 (Regulation on internal quality control systems in social services and health care)** [Lovdata]. Updated 2002, Dec 20; cited 2008 Feb 26 [http://www.lovdata.no/cgi-wifit/ldles?doc=/sf/sf/sf-20021220-1731.html].
 34. **Forskrift om innsamling og behandling av helseopplysninger i Meldingssystem for smittsomme sykdommer og i Tuberkuloseregisteret og om varslings om smittsomme sykdommer (MSIS- og Tuberkuloseregisterforskriften). FOR 2003-06-20 nr 740 (Regulations on collection and processing of health information in The Norwegian Surveillance System for Communicable Diseases and Tuberculosis Registry and on warning of communicable diseases.)** [Lovdata]. Updated 2003, Jun 20; cited 2008 Feb 26 [http://www.lovdata.no/cgi-wifit/ldles?doc=/sf/sf/sf-20030620-0740.html].
 35. Gorovitz S, MacIntyre A: **Toward a theory of medical fallibility.** *Hastings Cent Rep* 1975, **5**:13-23.
 36. Klækken B: **Minst 100 smittet av munnpensel (At least 100 infected by mouth swab).** *Fædrelandsvennen* 2002, 10th April.
 37. Altmann C: **Fortvilet direktør stopper produksjonen (Devastated manager stops production).** *Aftenposten* 2002, page 3, 10 April.
 38. Fehr AL von der: **Han gir Snøgg skylden (He blames Snøgg)** [VG]. Oslo. Updated 2003, Feb 25; cited 2008 Feb 26 [http://www.vg.no/pub/vgart.hbs?artid=6006806].
 39. **Lov om medisinsk utstyr. LOV-1995-01-12-6 (Act on Medical devices)** [Lovdata]. Updated 1995 Jan 12; cited 2008 Jul 15 [http://www.lovdata.no/all/nl-19950112-006.html].
 40. **Lov om helsepersonell m.v. LOV 1999-07-02 nr 64 (Act on health personnel etc.)** [Lovdata]. Updated 1999, Jul 2; cited 2008 Sep 2 [http://www.lovdata.no/all/hl-19990702-064.html].
 41. Reason J: **Human error: models and management.** *BMJ* 2000, **320**:768-770.
 42. **Rapport til Helsedepartementet om Helsetilsynets oppfølging i Dent-O-Sept saken (Report to the Ministry of Health on Follow-up of the Dent-O-Sept Incident by the Norwegian Board of Health)** Oslo: Statens helsetilsyn; 2003.
 43. **Dent-O-Sept saken. Vurderinger fra Sosial- og helsedirektoratet som nasjonal smittevernmyndighet (The Dent-O-Sept case. Considerations by the Directorate of Health and Social Services as national infection control authority)** Oslo: Sosial- og helsedirektoratet; 2004.
 44. **Lov om erstatning ved pasientskader mv. (pasient-skadefoven). LOV 2001-06-15 nr 53 (Act relating to compensation for harm on patient (Patient Harm Act))** [Lovdata]. Updated 2001, Jun 15; cited [http://www.lovdata.no/all/hl-20010615-053.html].
 45. Reiersen N: **Compensation to patients after injury.** In *Health legislation in Norway* Edited by: Molven O. Oslo: University of Oslo; 2002:137-143.
 46. Thomsen MV: **Dent-O-Sept - sluttrapport (Dent-O-Sept - Final report)** [Norsk pasientskadeerstatning]. Updated 2007, Apr 26; cited 2007 Nov 6 [http://www.npe.no/domino/npe/cms3603no.nsf/frames/index.html?open4=/(\$All)/899414C4F71774DC1256D03003DA544?OpenDocument].
 47. Ofteidal H: **Fire millioner i erstatning (Four million in compensation)** [Fædrelandsvennen]. Kristiansand. Updated 2006, Jun 22; cited 2008 Feb 26 [http://www.fvn.no/na24/article374985.ece].



ELSEVIER



www.elsevierhealth.com/journals/jinf

Nationwide study of invasive *Pseudomonas aeruginosa* infection in Norway: Importance of underlying disease

Bjørn G. Iversen ^{a,*}, Arne B. Brantsæter ^{a,b}, Preben Aavitsland ^a

^a Norwegian Institute of Public Health, P.O. Box 4404 Nydalen, NO-0403 Oslo, Norway

^b Asker and Bærum Hospital, P.O. Box 83, NO-1309 Rud, Norway

Accepted 28 May 2008

Available online 9 July 2008

KEYWORDS

Pseudomonas aeruginosa;
Bacteraemia;
Nosocomial infection;
Mortality;
Epidemiology;
Central nervous system infections;
Pseudomonas infections;
Gram-negative bacteria;
Hospital-acquired infection;
Community-acquired infection

Summary *Objective:* *Pseudomonas aeruginosa* is an opportunistic pathogen that may cause invasive disease. We describe the epidemiology of invasive *P. aeruginosa* infection in Norway and identify associated clinical factors.

Methods: All patients with invasive *P. aeruginosa* and *Pseudomonas* not identified at the species level (*Pseudomonas* spp.) in Norway 1992–2002 were included. Detailed information was collected for all cases during 1999–2002. Population and health institution statistics were obtained from national databases.

Results: In 1999–2002 the incidence rate was 3.16 per 100 000 person-years at risk or 0.20 per 1000 hospital stays. For hospital-acquired infection the rate was 671 per 100 000 person-years as compared with 1.13 for community-acquired infection, and 37 in nursing homes. The highest risk for invasive *Pseudomonas* disease was found in patients with malignant neoplasms of lymphoid and haematopoietic tissue (risk per 1000 hospital stays 1.9; 95% CI 1.5–2.3) and other diseases of blood and blood-forming organs (2.2; 95% CI 1.2–3.7). The case fatality rate was 35%.

Conclusions: The incidence of invasive *P. aeruginosa* infection in this population-based study was much lower than in most single-hospital studies. The nationwide study design and prudent antibiotic use may explain some of the difference. Infection risk is strongly associated with certain underlying diseases.

© 2008 The British Infection Society. Published by Elsevier Ltd. All rights reserved.

Introduction

Pseudomonas aeruginosa, a gram-negative aerobic bacterium with minimal nutritional requirements, is common in moist environments.^{1,2} It rarely causes infection in healthy humans but may do so following disruption of physical barriers and in patients with certain underlying illnesses.^{1–3}

* Corresponding author. Tel.: +47 21 07 65 16; +47 93 46 03 75 (mobile); fax: +47 21 07 65 13.

E-mail address: bjorn.iversen@fhi.no (B.G. Iversen).

P. aeruginosa produces several virulence-associated factors and can cause a variety of disease manifestations. In addition to bacteraemia and endocarditis, infection of the urinary tract, respiratory tract, central nervous system, ear, eye, bone, joints and skin are most often reported.^{1–5} Biofilm formation is an important factor in disease persistence for example in patients with cystic fibrosis.⁶ *Pseudomonas* species is ranked among the top 10 causes of bacteraemias in hospitals.^{7–12} In-hospital crude case, fatality from invasive disease is high, ranging from 18% to 61%.^{4,13–23} *Pseudomonas* species other than *P. aeruginosa* infrequently cause infection.²⁴

Little is known about the epidemiology of invasive *P. aeruginosa* infection in humans in Norway as these infections are not covered by national surveillance systems. Only one hospital has instituted screening of all patients being admitted to the intensive care unit (ICU) after a local outbreak.²⁵

During the investigation and follow-up of a large national outbreak of *P. aeruginosa* infection in 2001–2002,^{26,27} we collected detailed information on all laboratory-confirmed invasive infections in the country. The objective of this study was to describe the epidemiology of invasive *P. aeruginosa* infection in Norway, and to identify patient groups at increased risk of disease and of death from *P. aeruginosa* infection. In contrast to earlier published studies, this is a population-based study that includes all hospitals in a country. By using complete hospital statistics, we are able to estimate national incidence rates and mortality rates of *P. aeruginosa* infections by groups of underlying diseases.

Materials and methods

A case of invasive *P. aeruginosa* infection was defined as a patient with *P. aeruginosa* or *Pseudomonas* not identified at the species level (*Pseudomonas* spp.) isolated from blood or cerebrospinal fluid (CSF) during the period 1992–2002. In June 2002, we asked all 22 medical microbiological laboratories in Norway to supply lists with name, unique personal identification number and certain other specified information for all cases and to continue prospective reporting until the end of 2002. The laboratories used their standard diagnostic methods; a few based their identification on colony characteristics only. As a recommended routine, two sets of blood cultures are taken from two separate puncture sites. Each set comprises two bottles, usually one aerobic and one anaerobic, each drawing up to 10 ml blood.²⁸ Available cultures from 1999 to 2002 were genotyped to identify cases with the outbreak strain.²⁶

For patients diagnosed during 1999–2002 we asked the patients' physicians to complete a form containing questions on type of *Pseudomonas* infection according to national definitions,²⁹ potential risk factors, discharge diagnoses (free space on the questionnaire, up to 10 diagnoses were recorded in the database), sequelae (including death), and administrative information, including name, the unique personal identification number, name and type of health institution, transfer from other institutions and the dates of admission, discharge, microbiological sampling and death.

Cases, who had been admitted to a hospital ≥ 48 h before collection of the samples that harboured *Pseudomonas*, were defined as having a hospital-acquired infection (HAI). Those living in or having been hospitalised from a nursing home (NH) for the elderly <48 h before the samples were taken, were defined as having nursing home-acquired infection (NHAI). The remainder we had information was defined as having community-acquired infection (CAI).

Population statistics and the number of beds in NHs were downloaded from Statistics Norway (www.ssb.no). The number of stays and the number of days of hospitalisation in somatic hospitals by region, age and discharge diagnoses (main and up to seven subordinate discharge diagnoses) were supplied by The Norwegian Patient Register (www.shdir.no/norsk_pasientregister/), a public service organisation supplying official hospital statistics.

We selected several disease categories that have been shown to be associated with increased risk of invasive *P. aeruginosa* infection^{1,24} and grouped them according to ICD-10 (International Classification of Diseases, 10th Revision).³⁰ We calculated the incidence proportion of in-hospital infection and in-hospital death for these disease groups, using the number of *Pseudomonas* cases and deaths within each diagnosis category as numerator, and the number of patients discharged from hospital with a diagnosis in the same group as denominator. To identify risk factors for dying among the cases we performed stepwise multivariate binomial regression analyses including the following variables in the initial model: age; gender; year and month of diagnosis; clinical *Pseudomonas* diagnosis; surgical, immunosuppressive, antibiotic or ventilator treatment; ICU admittance; place of acquisition and having an underlying risk diagnosis.

We entered all patient data in an Epi Info version 6.04d database and analysed them in Epi Info, Excel, Episheet and Stata 8 and 9 statistical softwares. The Norwegian Institute of Public Health (NIPH) was authorised by the Norwegian Board of Health to perform the study as part of the investigation of a nationwide outbreak.²⁶ The Data Inspectorate gave permission to create a database to store the information. Extensive efforts were made to ensure completeness and quality of data, including linkage with the National Population Registry to search for deaths among the patients.

Results

Incidence 1992–2002

We identified 1174 cases over the 11-year period, of which 1079 (92%) had isolates of *P. aeruginosa* and 95 had *Pseudomonas* not identified at the species level (*Pseudomonas* spp.), resulting in an overall incidence rate of 2.43 per 100 000 person-years at risk (pyar). As other species of *Pseudomonas* only infrequently cause disease,²⁴ we will in the analysis below assume that all were cases of *P. aeruginosa* infection.

The patients' age ranged from 13 days to 100 years (median 72 years) and 67% were male. The age and gender distribution did not change much over the years. Nearly all

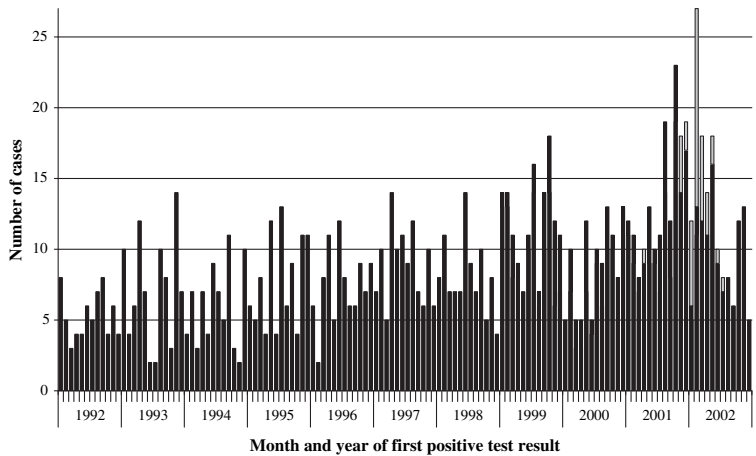


Figure 1 The monthly number of cases (40 cases (gray bars) belonged to an outbreak caused by a contaminated mouth swab^{26,27}) of invasive *P. aeruginosa* infection in Norway 1992–2002.

patients had a blood isolate, four had only a CSF isolate, while two had both.

During the study period there was a small but significant increase in the incidence, averaging 0.18 cases per 100 000 pyar for each year. However, data were not available from four of the 22 laboratories in 1992, three in 1993, two in 1994, and one in the first half of 1995. The monthly number of cases varied from five to 27 (Fig. 1).

The following detailed analysis is restricted to the 567 cases from 1999 to 2002.

Incidence, case fatality and mortality by age and gender, 1999–2002

For the period 1999–2002, 567 incident cases (representing 565 patients) were identified, corresponding to an

incidence rate of 3.16 per 100 000 pyar (95% CI, 2.90–3.43) (Table 1). In hospitals the incidence rate was 3.33 per 100 000 person-days (95% CI, 3.06–3.62), or 0.20 per 1000 hospital stays (95% CI, 0.18–0.21). Both morbidity and mortality increased with age (Table 1).

Two patients were included twice with different episodes of bacteraemia three and nine months apart. Only three patients had isolates from CSF, of whom two had hydrocephalus and one had a lumbar drain. One patient was in a NH located in a hospital building when diagnosed and is included in the numerator for hospital rates. One hundred and ninety nine (35%) cases died while hospitalised (Table 1).

The rate of infection was 5.1 per 100 000 person-days in hospital in males and 1.9 in females (385 males and 182 females) (incidence rate ratio [IRR], 2.6; 95% CI, 2.2–3.1).

Table 1 The incidence and mortality of invasive *P. aeruginosa* infection by place of acquisition and age group in Norway 1999–2002

Age group	Cases (and deaths) by place of acquisition					CFR (%) ^a	Incidence of all cases per 100 000		Mortality of all cases per 100 000	
	CAI	HAI	NHAI	Unknown	Total		Pop. yrs	Hosp. days	Pop. yrs	Hosp. days
0–9 years	3 (1)	4 (2)			7 (3)	43	0.29	0.52	0.12	0.22
10–19 years	0 (0)	6 (0)			6 (0)	0	0.27	1.29	0.00	0.00
20–29 years	6 (0)	9 (2)			15 (2)	13	0.62	1.28	0.08	0.17
30–39 years	1 (0)	12 (0)			13 (0)	0	0.48	0.88	0.00	0.00
40–49 years	8 (1)	18 (5)	1 (0)	1 (0)	28 (6)	21	1.12	2.25	0.24	0.48
50–59 years	11 (4)	49 (20)	2 (0)	1 (0)	63 (24)	38	2.87	3.25	1.10	1.24
60–69 years	41 (12)	60 (22)	1 (0)	3 (1)	105 (35)	33	7.46	4.59	2.49	1.53
70–79 years	70 (26)	89 (49)	17 (3)	2 (1)	178 (79)	44	13.91	4.84	6.17	2.15
80–89 years	44 (15)	57 (22)	24 (6)	1 (0)	126 (43)	34	18.68	4.28	6.38	1.46
≥90 years	8 (1)	9 (2)	9 (4)		26 (7)	27	25.39	5.34	6.84	1.44
Total	192 (60)	313 (124)	54 (13)	8 (2)	567 (199)	35	3.16	3.33	1.11	1.17

^a CFR = case fatality rate; CAI = community-acquired infection; HAI = hospital-acquired infection; NHAI = nursing home-acquired infection.

For hospital-acquired cases (221 males and 92 females) the rates were 2.9 and 1.0 per 100 000, respectively (IRR, 3.0; 95% CI, 2.3–3.8). As with morbidity the overall in-hospital mortality rates were also higher in males with 1.6 per 100 000 person-days in hospital in males and 0.8 in females (122 males and 77 females) (incidence mortality rate ratio [IRR], 2.0; 95% CI, 1.5–2.6). However, the overall case fatality rate was higher in females, 42% vs. 32% in males (fatality risk ratio [FRR] 1.3, 95% CI, 1.1–1.7). In a multivariate analysis, female gender was no longer associated with an increased risk of dying among the cases.

Place of acquisition

Patients were diagnosed at 55 hospitals and one NH (one patient), ranging from 1 to 70 cases per institution for the 4-year period. A total of 55% of the cases were hospital-acquired and an additional 10% were living in or being hospitalised from a NH. The remaining 34% were community-acquired and 1% was unknown. The incidence was 671 per 100 000 pyar in hospital for hospital-acquired infection as compared with 1.13 per 100 000 pyar for community-acquired infection (IRR 627, 95% CI, 524–751). For people living in NHs, the rate was 37 cases per 100 000 pyar compared to 1.13 per 100 000 pyar for community-acquired infection (IRR 18, 95% CI, 14–24).

For all cases, the median length of the total stay in hospital was 15 days and the median time from the *P. aeruginosa* sample was taken to discharge was 13 days. For the HAI cases, the time from admission to the culture was taken varied from 2 to 107 days with a median of 11 days. For those dying, the median time from diagnosis to death was only 3 days.

Rates of community-acquired infection rose sharply from the age of 60 years. For the hospital-acquired cases the increasing infection rate with age started at a lower age, and the age-related rates per 100 000 person-days in hospital showed an almost two-tiered distribution with a break point at 50 years (Fig. 2). The HAI rates for the age groups 50+ years and 0–49 years were 2.3 and 0.86, respectively, per 100 000 person-days in hospital (IRR, 2.7; 95% CI, 2.0–3.7).

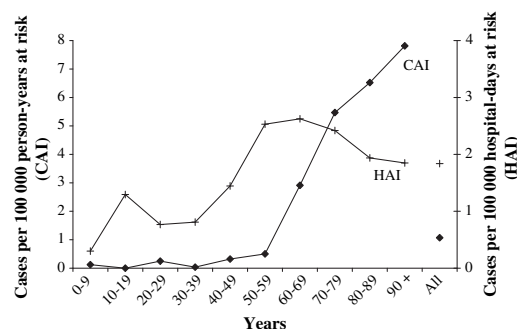


Figure 2 Invasive *P. aeruginosa* infection in Norway 1999–2002 by age groups and place of acquisition (Community-acquired infection (CAI), and hospital-acquired infection (HAI)).

Clinical features

Septicaemia was the most commonly reported clinical manifestation (81%), followed by urinary tract infection and pneumonia (Table 2). The two patients with isolates from CSF had meningitis, and the one with isolates from blood and CSF had septicaemia. Of the other reported diagnoses, seven had infection in the gastrointestinal tract, seven in the biliary system, one had pancreatitis, one endocarditis (in a patient with an artificial mitral valve), and three patients were reported as only being colonised, despite isolation of *P. aeruginosa* from their blood. The primary source of infection for the 460 cases classified as having septicaemia was not available.

Of the patients with invasive *P. aeruginosa* infection, 14% received mechanical ventilation within a period of 3 weeks before the sample positive for *P. aeruginosa* was taken, and 30% were admitted to an ICU during their stay in the hospital.

The 268 patients who received antibiotic treatment during the 3 weeks preceding sampling for *P. aeruginosa*, received a total of 467 courses of antibiotics (mean 1.7 per patient, ranging from one to seven). Eighty-one patients (14%) received a total of 101 systemic antibiotics within the ATC J01-group with activity against *Pseudomonas* [CR 05 (piperacillin tazobactam), DD 02 (ceftazidime), DH 02 (meropenem) and DH 51 (imipenem cilastatin), GB 01, 03 and 07 (tobramycin, gentamicin and netilmicin) and MA 01 and 02 (ofloxacin and ciprofloxacin)]. No patient was reported as having received DF 01 (monobactams) preceding their *Pseudomonas* infection.

Underlying disease and prior therapy

Patients with malignant neoplasms of lymphoid and haematopoietic tissue (ICD-10 categories³⁰ C81–C96) and other diseases of blood and blood-forming organs (D70–D77) had the highest risk of invasive *P. aeruginosa* disease and of dying from this infection while hospitalised (Table 3). The highest case fatality rate was seen among patients with transplanted organs (Z94) and patients with certain diseases of the respiratory system (J10–J22; J40–J96) (Table 3).

Table 2 Clinical manifestations of invasive *P. aeruginosa* infection in Norway 1999–2002

	1999	2000	2001	2002	Total	Deaths	CFR (%) ^a
Septicaemia	118	88	140	114	460	171	37
Pneumonia	7	6	9	11	33	17	52
Meningitis	1	0	0	2	3	2	67
Urinary tract infection	9	7	11	15	42	2	5
Wound infection	4	0	0	3	7	0	0
Unknown	0	0	1	2	3	1	33
Other	5	5	5	4	19	6	32
Total	144	106	166	151	567	199	35

^a CFR = case fatality rate.

Table 3 Number of cases of *P. aeruginosa* invasive infection and in-hospital death and the total number of discharges by certain ICD-10 discharge diagnoses in Norway 1999–2002

ICD-10 categories (codes)	Cases	Deaths	Discharges	Cases		Deaths		
				Risk ^a	95% CI	Risk ^a	95% CI	CFR (%)
Other diseases of blood and blood-forming organs (D70–D77)	14	6	6333	2.2	1.2–3.7	0.9	0.3–2.1	43
Malignant neoplasms of lymphoid and haematopoietic tissue (C81–C96)	94	41	49 685	1.9	1.5–2.3	0.8	0.6–1.1	44
Decubitus ulcer (L89)	7	2	4344	1.6	0.6–3.3	0.5	0.06–1.7	29
Human immunodeficiency virus [HIV] disease (B20–B24)	3	1	1875	1.6	0.4–5.1	0.5	0.01–3.0	33
Injury of nerves and spinal cord (S04; S14; S24; S34)	3	0	1915	1.6	0.3–4.6	0.0	—	0
Immunodeficiency with predominantly antibody defects (D80)	1	0	640	1.6	0.08–7.7	0.0	—	0
Cystic fibrosis (E84)	2	0	1373	1.5	0.2–5.3	0.0	—	0
Renal failure (N17–N19) and Dependence on renal dialysis (Z99.2)	50	25	42 205	1.2	0.9–1.6	0.6	0.4–0.9	50
Transplanted organ and tissue status (Z94)	9	6	8399	1.1	0.5–2.0	0.7	0.3–1.6	67
Burns and corrosions of external body surface (T20–T25)	3	1	4046	0.7	0.2–2.2	0.2	0.01–1.4	33
Urolithiasis (N20–N23)	13	2	24 128	0.5	0.3–0.9	0.08	0.01–0.3	15
Certain diseases of the respiratory system (J10–J22; J40–J96)	180	94	364 014	0.5	0.4–0.6	0.3	0.2–0.3	52
Malignant neoplasms (C00–C80)	137	57	367 817	0.4	0.3–0.4	0.2	0.1–0.2	42
Diabetes mellitus (E10–E14; O24)	46	14	149 299	0.3	0.2–0.4	0.1	0–0.2	30
Other medical care (cancer treatment) (Z51)	16	4	77 268	0.2	0.1–0.3	0.05	0.01–0.1	25
Certain diseases of the circulatory system (I20–I79)	147	69	813 978	0.2	0.1–0.2	0.09	0–0.1	47
Transport accidents (V)	2	0	16 729	0.1	0.02–0.4	0.0	—	0
Injuries, other (S00–S99) (not ICD-10 = S04; S14; S24; S34)	20	8	293 564	0.06	0–0.1	0.03	0–0.05	40
Disorders related to short gestation and low birth weight (P07)	1	0	24 804	0.04	0–0.2	0.0	—	0
Falls (W)	1	0	66 067	0.02	0–0.08	0.0	—	0
The number of risk diagnoses	751	330						
The number of patients with one or more risk diagnoses	462	184						
The number of patients with none of these risk diagnosis	105	15						
All patients	567	199	2 904 940	0.2	0.18–0.21	0.07	0.06–0.08	35

CFR = case fatality rate; 95% CI = 95% confidence interval.

^a Incidence risk and mortality risk per 1000 discharges.

In the multivariate regression analysis, the following variables were independently associated with an increased risk of dying (CFR) among the cases: Having one or more underlying risk diagnoses listed in Table 3 (risk ratio [RR], 2.52; 95% CI, 1.50–4.25), admission to intensive care unit at some time during the stay (RR, 1.55; 95% CI, 1.26–1.91), being 60 years or older (RR, 1.53; 95% CI, 1.14–2.06), and having received immunosuppressive treatment within the past 3 weeks before the sample with *P. aeruginosa* was taken (RR, 1.39; 95% CI, 1.13–1.73). Having a *Pseudomonas* UTI as the most serious clinical

Pseudomonas diagnosis was highly protective (RR, 0.09; 95% CI, 0.01–0.63).

Using other definitions

We have repeated all major calculations for the period 1999–2002 in two alternative data sets: (1) omitting all cases with *Pseudomonas* not identified at the species level (50 cases), and (2) omitting all cases that were part of an outbreak in 2001–2002 (40 cases). The omitted cases are evenly distributed on factors like age groups, gender,

clinical features, place of acquisition and underlying illness. The total incidence rate was then reduced from 0.20 per 1000 hospital stays to 0.18 in both the alternative data sets.

An alternative mortality measure to the "in-hospital death" used above is a "30 day case fatality rate". Counting 30 days from the sampling date, 184 cases died in hospital and an additional two after discharge, reducing the CFR from 35% to 33%. There was no clear pattern difference using either measure.

Discussion

In hospitalised patients and patients with underlying diseases, invasive *P. aeruginosa* infection is an important cause of disease and death. We have described the total number of patients with *P. aeruginosa* isolated from blood or CSF in Norway over an 11-year period. No previous study has reported the risk of acquiring invasive *P. aeruginosa* infection associated with specific underlying diseases at a national level. We found that both incidence of infection and mortality are strongly associated with increasing age and underlying disease.

Low overall incidence

For the period 1999–2002, we found a rate of invasive *P. aeruginosa* infection of 0.20 per 1000 hospital stays. Studies from other countries have indicated rates between 0.94 and 1.8 per 1000 hospital stays^{5,11,14–16,18,19,21} in tertiary referral hospitals and university hospitals and 0.43 and 0.59 per 1000 hospital stays in community hospitals.^{9,10} Apart from our inclusion of patients with positive CSF cultures, our inclusion criteria are very similar to the above-mentioned studies and cannot explain the low rate. The low incidence in Norway compared with studies from other countries may partly be explained by study design. Our study was nationwide and population-based and included all somatic hospital stays in Norway. Certain hospital departments such as dermatology and gynaecology and obstetrics, and community or specialty hospitals with less than 5000 discharges per year, are known to have low rates of *P. aeruginosa* invasive infection so these departments may have contributed to the low reported overall rates. However, no Norwegian hospital had a higher rate than 0.42 per 1000 hospital stays.

Indiscriminate use of antibiotics, especially broad spectrum antibiotics, is known to be associated with development of resistance and selection of resistant bacteria, such as *P. aeruginosa*.^{1–3,24,31} One possible explanation for the low rates of invasive *P. aeruginosa* infection observed in Norway may be prudent use of antibiotics in hospitals. Compared to hospitals in many other countries, Norwegian hospitals have low overall antibiotic consumption and low use of broad spectrum antibiotics. The use of narrow-spectrum drugs is encouraged.^{32–34} For instance, penicillin is the preferred choice for community-acquired uncomplicated pneumonia. The use of empiric broad spectrum antibiotics such as most 3rd generation cephalosporins, which may give *Pseudomonas* a selective advantage, are generally discouraged. For empiric treatment of septicaemia where broad spectrum

coverage is necessary, penicillin plus an aminoglycoside is the recommended standard treatment in most hospital departments. In general, aminoglycosides are active against *Pseudomonas* and have a high threshold for development of resistance (tobramycin considered most active), thereby avoiding selection pressure favouring these bacteria.³⁵

The importance of age and underlying disease

Our finding of 14% of the cases having received mechanical ventilation, and 30% having been admitted to an ICU can be compared with figures from 30 hospitals in the 2002 annual report from The Norwegian Intensive Care Register (Norsk intensivregister). Of the 3.4 million hospital-days, 0.73% was spent on a ventilator and 1.4% in an ICU.³⁶ These figures indicate that being on a ventilator or being admitted to the ICU increases the risk of acquiring invasive *P. aeruginosa* infection by a factor of 20, which is in accordance with other studies.^{4,14,17,18,20,21}

Several risk factors for invasive *P. aeruginosa* infection and death have previously been identified.^{1–4,14,17,18,20,21,37} Using complete national discharge statistics, we were able to calculate absolute rates of infection among patients with various underlying diseases. Our study confirms that *P. aeruginosa* infection is a major risk to patients with cancer, immunodeficiency, renal failure and certain other diseases.

Several studies have shown that the incidence of invasive *P. aeruginosa* infection increases with age, and report mean and median ages between 54 and 70 years^{4,9,15,17,18,20,21}; our findings are in the upper end of that range. The lower age of hospital-acquired cases (Fig. 2) may simply reflect more hospitalised patients have underlying diseases irrespective of age while in the general population the prevalence of underlying illnesses increases sharply from the age of 60.

We found that the rates of hospital-acquired infection are three times higher in males. Most studies show a majority of infection in males, varying from 55% to 72% of the included cases,^{4,9,15–18,20,21} but none of them show rates per hospital population.

In our study, 55% of the cases were hospital-acquired and a further 10% acquired in a NH. One other study report a similar proportion (65%),¹⁵ whereas most studies report a proportion of 80% or higher.^{14,16,18,21} This may be explained by higher incidence in university and referral hospitals.

A deadly disease or a disease of the dying?

Invasive *P. aeruginosa* infection is a serious disease with a high case fatality rate, 35% in our study. Most patients who died from invasive *P. aeruginosa* infection died within a short time of being diagnosed.

The patient groups with the following underlying diseases had the highest mortality risk per 1000 discharges: malignant neoplasms of lymphoid and haematopoietic tissue, other diseases of blood and blood-forming organs, organ transplantation and renal failure. Among the patients with invasive *P. aeruginosa* infection the following factors were independently associated with an increased CFR: having an underlying risk diagnoses associated with

P. aeruginosa infection (listed in Table 3); admission to an ICU; old age and immunosuppressive treatment prior to infection. A clinical *P. aeruginosa* diagnosis only of UTI was protective, whereas meningitis, pneumonia and septicæmia increased the risk although not significantly. Other studies list similar risk factors.^{14,17,18,21} As demonstrated by others,¹⁵ bacteraemic pneumonia had a high CFR. Other studies have examined different combinations of various risk factors, but we found no contradictions to previously published studies.

P. aeruginosa can produce a variety of virulence factors and there is substantial variability among strains.^{1,3,38,39} The presence of certain virulence factors of the Type III secretion system has showed to increase the relative risk of death more than six times.³⁸ In the present study we did not examine the strains for virulence factors.

Limitations

We had access to quality controlled national population and hospital statistics. However, the numerator data were collected during an outbreak investigation of *P. aeruginosa* infection.²⁶ Although great efforts were made to ensure that the outbreak investigation was given highest priority, the quality of the returned questionnaires varied. Extensive efforts were made to ensure completeness and quality of the data, including contacting the clinicians and linkage with the National Population Registry to search for deaths among patients. Consequently, we believe most of the collected patient data are accurate, but some information may have been missed, especially regarding some of the subordinate discharge diagnoses. However, all but three of the 567 patients had at least one main underlying disease recorded. As there were few patients in several of the risk diagnosis groups (Table 3), missing information in either of these groups could have influenced the incidence and mortality rates and the case fatality substantially.

All Norwegian medical microbiological laboratories provided numerator data, and all of them have kept complete records since 1996. The missing data for the first few years may at least partially explain the measured increase in incidence. We consider it unlikely that change in microbiological methods or diagnostic use of blood and CSF culture for detecting *Pseudomonas* has affected our results significantly.

We may have missed invasive infections, either because no blood or CSF was cultured, especially in the out-patient setting, or because culture is not a 100% sensitive. However, we believe this is an unlikely explanation for our comparatively low rates. We included 95 cases with *Pseudomonas* not identified at the species level (*Pseudomonas* spp.). These identifications mainly came from laboratories that based their identification on colony characteristics. Although some isolates may have belonged to other species and closely related genera, it is likely that most of these isolates represented *P. aeruginosa* as other species of *Pseudomonas* rarely causes infection in humans.²⁴

The national hospital statistics on discharge diagnoses were aggregate data. Thus, we were unable to do individual level analyses of relative risks for infection (Table 3), nor controlling for potential confounders by multivariate

regression analysis. As age and gender are the two major possible confounders that influence the incidence risks, we may not easily compare the groups of underlying diseases. However, for the CFR we were able to control for confounders by binomial multivariate regression.

Conclusions

In this population-based study of invasive *P. aeruginosa* infection in Norway, we found a much lower incidence than in single-hospital studies in other countries. The nationwide study design and prudent antibiotic use in Norway may be one possible explanation for some of the difference. Risk of infection is strongly associated with increasing age and certain underlying diseases. Hospitalised patients with malignant or other diseases of blood and blood-forming organs had a 0.2% risk of infection during a hospital stay, which is 10-fold higher than the general hospital population. Around 35% of cases died while hospitalised, either because of the infection or the underlying disease. Invasive *P. aeruginosa* infection remains an important cause of morbidity and mortality.

Conflict of interest

None of the authors have any conflict of interest concerning this work. All primary costs were covered by the health care institutions and national bodies involved.

Acknowledgements

We are indebted to the staff at the medical microbiological laboratories for performing the microbiological analyses and submitting data, infection control nurses and clinicians for filling in the patient forms, Arve Sjølingstad for providing hospital denominator data by discharge diagnoses from The Norwegian Patient Registry, The Norwegian Board of Health and The Directorate for Health and Social Affairs for fruitful cooperation.

References

1. Pollack M. *Pseudomonas aeruginosa*. In: Mandell GL, Bennett JE, Dolin R, editors. *Principles and practice of infectious diseases*. 5th ed. Philadelphia: Churchill Livingstone; 2000. p. 2310–35.
2. Bergogne-Berezin E. *Pseudomonas* and miscellaneous gram-negative bacilli. In: Cohen J, Powderly WG, editors. *Infectious diseases*. 2nd ed. Edinburgh: Mosby; 2004. p. 2203–26.
3. Arnow PM, Flaherty JP. Nonfermentative gram-negative bacilli. In: Mayhall CG, editor. *Hospital epidemiology and infection control*. 2nd ed. Philadelphia: Lippincott Williams & Wilkins; 1999. p. 431–51.
4. Kang CI, Kim SH, Kim HB, Park SW, Choe YJ, Oh MD, et al. *Pseudomonas aeruginosa* bacteremia: risk factors for mortality and influence of delayed receipt of effective antimicrobial therapy on clinical outcome. *Clin Infect Dis* 2003;37:745–51.
5. Sherertz RJ, Sarubbi FA. A three-year study of nosocomial infections associated with *Pseudomonas aeruginosa*. *J Clin Microbiol* 1983;18:160–4.
6. Häussler S. *Pseudomonas aeruginosa* biofilms: impact of small colony variants on chronic persistent infections. In: Cornelis P,

- editor. *Pseudomonas genomics and molecular biology*. Norfolk: Caister Academic Press; 2008. p. 159–75.
7. NORM/NORM-VET 2006. Usage of antimicrobial agents and occurrence of antimicrobial resistance in Norway. Tromsø/Oslo; 2007.
 8. National Nosocomial Infections Surveillance (NNIS). NNIS report, data summary from October 1986–April 1996, issued May 1996. A report from the National Nosocomial Infections Surveillance (NNIS) System. *Am J Infect Control* 1996;24:380–8.
 9. Javaloyas M, Garcia-Somoza D, Gudiol F. Epidemiology and prognosis of bacteremia: a 10-y study in a community hospital. *Scand J Infect Dis* 2002;34:436–41.
 10. Scheckler WE, Bobula JA, Beamsley MB, Hadden ST. Bloodstream infections in a community hospital: a 25-year follow-up. *Infect Control Hosp Epidemiol* 2003;24:936–41.
 11. Weinstein MP, Reller LB, Murphy JR, Lichtenstein KA. The clinical significance of positive blood cultures: a comprehensive analysis of 500 episodes of bacteremia and fungemia in adults. I. Laboratory and epidemiologic observations. *Rev Infect Dis* 1983;5:35–53.
 12. Diekema DJ, Pfaller MA, Jones RN, Doern GV, Winokur PL, Gales AC, et al. Survey of bloodstream infections due to gram-negative bacilli: frequency of occurrence and antimicrobial susceptibility of isolates collected in the United States, Canada, and Latin America for the SENTRY Antimicrobial Surveillance Program. *Clin Infect Dis* 1997;1999(29):595–607.
 13. Blot S, Vandewoude K, Hoste E, Colardyn F. Reappraisal of attributable mortality in critically ill patients with nosocomial bacteraemia involving *Pseudomonas aeruginosa*. *J Hosp Infect* 2003;53:18–24.
 14. Bisbe J, Gatell JM, Puig J, Mallolas J, Martinez JA, Jimenez de Anta MT, et al. *Pseudomonas aeruginosa* bacteremia: univariate and multivariate analyses of factors influencing the prognosis in 133 episodes. *Rev Infect Dis* 1988;10:629–35.
 15. Gallagher PG, Watanakunakorn C. *Pseudomonas* bacteremia in a community teaching hospital, 1980–1984. *Rev Infect Dis* 1989;11:846–52.
 16. Mallolas J, Gatell JM, Miro JM, Marco F, Soriano E. Epidemiologic characteristics and factors influencing the outcome of *Pseudomonas aeruginosa* bacteremia. *Rev Infect Dis* 1990;12:718–9.
 17. Micek ST, Lloyd AE, Ritchie DJ, Reichley RM, Fraser VJ, Kollef MH. *Pseudomonas aeruginosa* bloodstream infection: importance of appropriate initial antimicrobial treatment. *Antimicrob Agents Chemother* 2005;49:1306–11.
 18. Aliaga L, Mediavilla JD, Cobo F. A clinical index predicting mortality with *Pseudomonas aeruginosa* bacteraemia. *J Med Microbiol* 2002;51:615–9.
 19. Tacconelli E, Tumbarello M, Bertagnolio S, Citton R, Spanu T, Fadda G, et al. Multidrug-resistant *Pseudomonas aeruginosa* bloodstream infections: analysis of trends in prevalence and epidemiology. *Emerg Infect Dis* 2002;8:220–1.
 20. Marra AR, Bearman GM, Wenzel RP, Edmond MB. Comparison of severity of illness scoring systems for patients with nosocomial bloodstream infection due to *Pseudomonas aeruginosa*. *BMC Infect Dis* 2006;6:132.
 21. Vidal F, Mensa J, Almela M, Martinez JA, Marco F, Casals C, et al. Epidemiology and outcome of *Pseudomonas aeruginosa* bacteremia, with special emphasis on the influence of antibiotic treatment. Analysis of 189 episodes. *Arch Intern Med* 1996;156:2121–6.
 22. Weinstein MP, Murphy JR, Reller LB, Lichtenstein KA. The clinical significance of positive blood cultures: a comprehensive analysis of 500 episodes of bacteremia and fungemia in adults. II. Clinical observations, with special reference to factors influencing prognosis. *Rev Infect Dis* 1983;5:54–70.
 23. Kuikka A, Valtanen VV. Factors associated with improved outcome of *Pseudomonas aeruginosa* bacteremia in a Finnish university hospital. *Eur J Clin Microbiol Infect Dis* 1998;17:701–8.
 24. Kiska DL, Gilligan PH. *Pseudomonas*. In: Murray PR, editor. *Manual of clinical microbiology*. 8th ed. Washington, DC: ASM Press; 2003. p. 719–28.
 25. Bukholm G, Tannaes T, Kjelsberg AB, Smith-Erichsen N. An outbreak of multidrug-resistant *Pseudomonas aeruginosa* associated with increased risk of patient death in an intensive care unit. *Infect Control Hosp Epidemiol* 2002;23:441–6.
 26. Iversen BG, Jacobsen T, Eriksen HM, Bukholm G, Melby KK, Nygard K, et al. An outbreak of *Pseudomonas aeruginosa* infection caused by contaminated mouth swabs. *Clin Infect Dis* 2007;44:794–801.
 27. Iversen BG, Eriksen HM, Bo G, Hagestad K, Jacobsen T, Engeset E, et al. *Pseudomonas aeruginosa* contamination of mouth swabs during production causing a major outbreak. *Ann Clin Microbiol Antimicrob* 2007;6:3.
 28. Blodkultur. Strategimøte nr. 16, 2002 [Blood culture. Strategy meeting no. 16, 2002]. Oslo: Strategimøte; 2002.
 29. Definisjon og klassifikasjon av sykehusinfeksjoner IK-2556. [Definition and classification of hospital infections]. Oslo: Statens helsetilsyn; 1996.
 30. International statistical classification of diseases and related health problems. Available from: <http://www.who.int/classifications/apps/icd/icd10online/>. 10th Revision [WHO]. Updated 2007, cited Nov 29, 2007.
 31. Allen KD, Bartzokas CA, Graham R, Gibson MF, Gilbertson AA. Acquisition of endemic *Pseudomonas aeruginosa* on an intensive therapy unit. *J Hosp Infect* 1987;10:156–64.
 32. Ferech M, Coenen S, Dvorakova K, Hendrickx E, Suetens C, Goossens H. European Surveillance of Antimicrobial Consumption (ESAC): outpatient penicillin use in Europe. *J Antimicrob Chemother* 2006;58:408–12.
 33. Ferech M, Coenen S, Malhotra-Kumar S, Dvorakova K, Hendrickx E, Suetens C, et al. European Surveillance of Antimicrobial Consumption (ESAC): outpatient antibiotic use in Europe. *J Antimicrob Chemother* 2006;58:401–7.
 34. Vander Stichele RH, Elseviers MM, Ferech M, Blot S, Goossens H. Hospital consumption of antibiotics in 15 European countries: results of the ESAC retrospective data collection (1997–2002). *J Antimicrob Chemother* 2006;58:159–67.
 35. NORM/NORM-VET 2005. Usage of Antimicrobial Agents and Occurrence of Antimicrobial Resistance in Norway. Tromsø/Oslo; 2006.
 36. Flaatten H. Årsrapport fra Norsk Intensivregister (NIR) 2002 [Annual report from The Norwegian Intensive Care Register 2002]. Bergen; 2007.
 37. Llopis F, Grau I, Tubau F, Císnal M, Pallares R. Epidemiological and clinical characteristics of bacteraemia caused by *Aeromonas* spp. as compared with *Escherichia coli* and *Pseudomonas aeruginosa*. *Scand J Infect Dis* 2004;36:335–41.
 38. Roy-Burman A, Savel RH, Racine S, Swanson BL, Revadigar NS, Fujimoto J, et al. Type III protein secretion is associated with death in lower respiratory and systemic *Pseudomonas aeruginosa* infections. *J Infect Dis* 2001;183:1767–74.
 39. Berthelot P, Attree I, Plesiat P, Chabert J, de Bentzmann S, Pozzetto B, et al. Genotypic and phenotypic analysis of type III secretion system in a cohort of *Pseudomonas aeruginosa* bacteremia isolates: evidence for a possible association between O serotypes and exo genes. *J Infect Dis* 2003;188:512–8.